

Antibiotic Susceptibility Pattern of Methicillin-Resistant *Staphylococcus aureus* from the Isolated Wound Culture in the Northwest Region, Kingdom of Saudi Arabia

Ibrahim Al Balawi¹, Palanisamy Amirthalingam^{2*}, Abdullah Abdul Khalig Alyoussef³, Osama Salih Mohammed³, Hyder Oman Mirghani³ and Amgad A. Ezzat⁴

¹Department of Surgery, Faculty of Medicine, University of Tabuk, Tabuk, Kingdom of Saudi Arabia

²Department of Clinical Pharmacy, Faculty of Pharmacy, University of Tabuk, Tabuk, Kingdom of Saudi Arabia; amirpalanisamy15@gmail.com

³Department of Medicine, Faculty of Medicine, University of Tabuk, Tabuk, Kingdom of Saudi Arabia

⁴Department of Microbiology, Faculty of Medicine, University of Tabuk, Tabuk, Kingdom of Saudi Arabia

Abstract

The present study aimed to investigate the antibiotic susceptibility pattern of Methicillin-resistant *Staphylococcus aureus* (MRSA) in isolated wound cultures of the patients admitted in King Khalid Hospital, Tabuk, Kingdom of Saudi Arabia. A retrospective cohort study of 54 patients admitted with wound infections in the surgical department. Ethics committee approval was granted by the University of Tabuk and King Khalid Hospital, Kingdom of Saudi Arabia. Fifty-four adult patients (>18 years old) diagnosed with moderate to severe skin and soft tissue infections were included in the study. 26 patients with isolated cultures of MRSA were compared with 28 patients with Methicillin-sensitive *Staphylococcus aureus* (MSSA) isolated cultures using Graph pad prism 4.0 version statistical databases. Overall, there was no significant difference in sensitivity (P=0.2445) and resistance (P=0.4215) between MRSA and MSSA cultures. However, it is interesting findings that Oxacillin and Fusidic acid had higher resistance in MRSA isolated cultures compared MSSA culture, on the other hand, Linezolid, Tigecycline and Nitrofurantoin shows 100% sensitivity in both MRSA and MSSA isolates. No significant difference between male and female regarding the sensitivity (P=0.0638) and resistance (P=0.3638). The current study emphasizes that Tigecycline, Nitrofurantoin and Fusidic acid were the best drugs in both MRSA and MSSA isolates. While, oxacillin showed 100% resistance to MRSA; but retain its efficacy on MSSA isolates.

Keywords: Antibiotic, Cultures, Sensitive

1. Introduction

Wound infections have been a problem in the field of medicine for a long time. The presence of foreign materials increases the risk of serious infection even with relatively small bacterial inoculums¹. Advances in control of infections have not completely eradicated this problem because of the development of drug resistance². The widespread misuses of antibiotics over a long time have led to emergences of resistant organisms contributing

to morbidity and mortality³⁻⁵. Antimicrobial resistance can increase complications and costs associated with procedures and treatment⁶.

The most common isolated aerobic microorganisms were *Staphylococcus aureus*, Coagulase-negative staphylococci (CoNS), Enterococci, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Enterobacter* species, *Proteus mirabilis*, *Candida albicans* and *Acinetobacter*^{7,8}.

Wound infections can be caused by different groups

* Author for correspondence

of microorganisms like bacteria, fungi, and protozoa. However, different microorganisms can exist in polymicrobial communities especially in the margins of wounds and in chronic wounds⁹. The infecting microorganism may belong to aerobic as most commonly isolated aerobic microorganism include *Staphylococcus aureus*, Coagulase-negative staphylococci (CoNS), Enterococci, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Enterobacter* species, *Proteus mirabilis*, *Candida albicans* and *Acinetobacter*^{7,8}.

The genus *Staphylococcus* includes pathogenic organisms in which *Staphylococcus aureus* is most important. It overcomes most of the therapeutic agents that have been developed in the recent years; hence, the antimicrobial chemotherapy for this species has always been empirical¹⁰. After the emergence of MRSA as a nosocomial pathogen in the early 1960s¹¹, an increasing number of outbreaks due to MRSA infections in hospitals have been reported from many countries, ranging from abscesses to life-threatening sepsis, endocarditis, and osteomyelitis¹².

Early diagnosis of MRSA and treatment by following standard antibiotic guidelines will reduce morbidity and mortality rate in tertiary care hospitals. Understanding the antibiotic susceptibility pattern of MRSA in a specific geographic region is vital in the selection of the appropriate empirical antibiotic therapy. The present study was carried out to determine the antibiotic susceptibility pattern of MRSA from the isolated wound cultures, and update the clinicians about the optimal antibiotics to treat wound infections.

2. Methods

A retrospective study conducted at King Khalid Hospital in Tabuk City during the period from June to December 2014, following the ethical guidelines for patient data privacy fifty-four wound cultures of adult patients (>18 years old) and sensitivity forms referred from the surgeons from the surgical department were reviewed. The research was cleared by the ethical committee of the University

of Tabuk and King Khalid Hospital, Tabuk, Kingdom of Saudi Arabia. Graph pad InStat Prism 4.0 version was used for data analysis. The t-test was performed to compare the sensitivity and resistance pattern between MRSA and Methicillin-sensitive *Staphylococcus aureus* (MSSA) isolates.

Information collected includes: sex, type of specimen included is wounds. Media used for bacterial isolation was carried using the serial dilution technique on the wound swabs to isolate *Staphylococcus aureus*.

2.1 Specimen Collection

Samples were collected from the patients with complaints of wound sepsis. The wound samples were collected by using a sterile cotton swab, the inner surface of the infected area was swabbed gently and then the swabs were transported to the laboratory.

2.2 Bacteriology and Antibiotic Susceptibility Testing

Bacterial isolation according to morphology followed according to Benson *et al.*, 1994¹³. Antibiotic sensitivity test was performed by using the Kirby-Bauer disk diffusion method recommended by the National Committee for Clinical Laboratory Standard (NCCLS, 2000)¹⁴ for the following antibiotics: cefoxitin, penicillin, oxacillin, gentamicin, tobramycin, levofloxacin, moxifloxacin, erythromycin, clindamycin, linezolid, teicoplanin, vancomycin, tetracycline, tigecyclin, fosfomicin, nitrofurantoin, fusidic acid, mupirocin, rifampicin, trimethoprim-sulphamethoxazole etc. The Vitex 12, Phoenix, and Micro scans were used. MRSA test was performed using cefoxitin 30 µg disc on Mueller-Hinton agar with 24 hours incubation at 35°C. The antibiotic discs used for the susceptibility tests were from Hi-Media Laboratories Pvt. Limited, India. A zone of inhibition less than 10 mm or any discernible growth within a zone of inhibition was indicative of methicillin resistance. *Staphylococcus aureus* ATCC 25923 (Manassas, VA, USA) was used as a standard control strain.

Table 1. Minimum inhibitory concentration (MIC) of the antibiotics

| S.No. | Antibiotic | MIC* |
|-------|--------------------------------|-------|
| 1 | Cefoxitin | POS** |
| 2 | Penicillin | ≥0.5 |
| 3 | Oxacillin | ≥4 |
| 4 | Gentamicin | ≥16 |
| 5 | Tobramycin | 8 |
| 6 | Levofloxacin | 4 |
| 7 | Moxifloxacin | 1 |
| 8 | Erythromycin | ≥8 |
| 9 | Clindamycin | ≤0.25 |
| 10 | Linezolid | 2 |
| 11 | Teicoplanin | 2 |
| 12 | Vancomycin | ≤0.5 |
| 13 | Tetracycline | 2 |
| 14 | Tigecyclin | ≤0.12 |
| 15 | Fosfomycin | 32 |
| 16 | Nitrofurantoin | ≤16 |
| 17 | Fusidic acid | ≥32 |
| 18 | Mupirocin | ≤2 |
| 19 | Rifampicin | ≤0.5 |
| 20 | Trimethoprim+Sulphamethoxazole | ≥320 |

*MIC: Minimum Inhibitory Concentration

**POS: Positive

3. Results

Out of 199 wound sepsis samples at the King Khalid Hospital, Tabuk, Kingdom of Saudi Arabia, July to December 2014, predominant *Staphylococcus aureus* (27%; n=54) followed by *Pseudomonas aeruginosa* (20%; n=39), *E. coli* (15%; n=30) etc. (Figure 1). Among the *Staphylococcus aureus*, MRSA resistance (48%) and 28 (52%) with MSSA resistance (Figure 2) observed in the *Staphylococcus aureus* isolates. Gender distribution reveals predominant male population in MRSA (n=25; 96%) and MSSA resistance samples (n=16; 57%) (Figure 3).

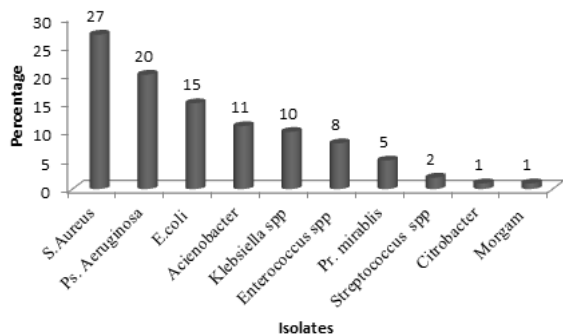


Figure 1. Percentage of isolates from wound sepsis (n=199).

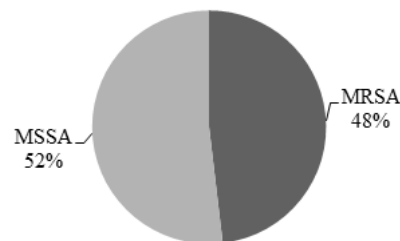


Figure 2. Percentage of MRSA and MSSA among the culture sample (n=54).

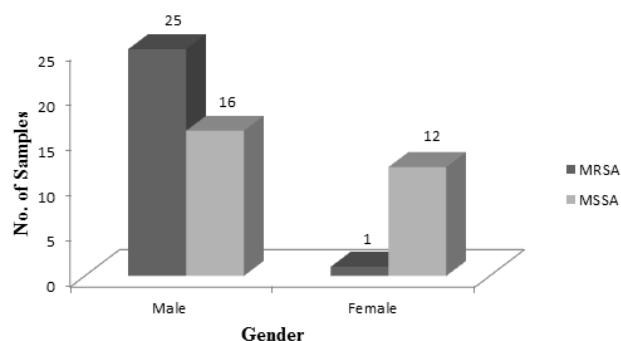


Figure 3. Gender distribution among the culture sample (n=54).

3.1 Association Antibiotic Sensitivity and MRSA

Linezolid, Teicoplanin, Vancomycin, Tigecyclin, Nitrofurantoin and Mupirocin were found to have 100% sensitivity against MRSA isolates; however, Penicillin and Oxacillin were found to be 100% resistant. On the other hand, Gentamycin, Tobramycin, Linezolid, Tetracycline, Fosfomycin and Nitrofurantoin were found to have 100% sensitivity against MSSA isolates. Both sensitivity (P=0.2445) and resistance (P=0.4215) were found statistically not significant. However, a significant difference was observed between isolates of MRSA sensitivity and resistance (P<0.0001). Similarly, MSSA isolates also shows a significant difference (P<0.0001). Interestingly, the present study observed some findings as follows 1. Oxacillin shows 96% sensitivity in MSSA isolates and 0% sensitivity in MRSA isolates, 2. Fusidic acid was 93% sensitivity in MSSA Vs. 46% in MRSA isolates. 3. Linezolid, Tigecyclin, and Nitrofurantoin shows 100% sensitivity in both MRSA and MSSA isolates (Table 2).

Table 3. Antibiotic sensitivity in Methicillin sensitive staphylococcus aureus among the gender (n=54)

| Antibiotic | Male (16) | | | | Female (12) | | | |
|--------------------------------|-----------|-----|----|-----|-------------|-----|----|-----|
| | S | % | R | % | S | % | R | % |
| Cefoxitin | 0 | 0 | 16 | 100 | 0 | 0 | 12 | 100 |
| Penicillin | 0 | 0 | 16 | 100 | 0 | 0 | 12 | 100 |
| Oxacillin | 15 | 93 | 1 | 7 | 12 | 100 | 0 | 0 |
| Gentamicin | 16 | 100 | 0 | 0 | 12 | 100 | 0 | 0 |
| Tobramycin | 16 | 100 | 0 | 0 | 12 | 100 | 0 | 0 |
| Levofloxacin | 13 | 81 | 3 | 19 | 11 | 92 | 1 | 8 |
| Moxifloxacin | 12 | 75 | 4 | 25 | 11 | 92 | 1 | 8 |
| Erythromycin | 14 | 88 | 2 | 12 | 11 | 92 | 1 | 8 |
| Clindamycin | 15 | 93 | 1 | 7 | 11 | 92 | 1 | 8 |
| Linezolid | 16 | 100 | 0 | | 12 | 100 | 0 | 0 |
| Teicoplanin | 15 | 93 | 1 | 7 | 12 | 100 | 0 | 0 |
| Vancomycin | 15 | 93 | 1 | 7 | 11 | 92 | 1 | 8 |
| Tetracycline | 16 | 100 | 0 | 0 | 12 | 100 | 0 | 0 |
| Tigecyclin | 16 | 100 | 0 | 0 | 12 | 100 | 0 | 0 |
| Fosfomycin | 16 | 100 | 0 | 0 | 12 | 100 | 0 | 0 |
| Nitrofurantoin | 16 | 100 | 0 | 0 | 12 | 100 | 0 | 0 |
| Fusidic acid | 15 | 93 | 1 | 7 | 10 | 62 | 2 | 38 |
| Mupirocin | 15 | 93 | 1 | 7 | 12 | 100 | 0 | 0 |
| Rifampicin | 4 | 25 | 12 | 75 | 9 | 56 | 3 | 44 |
| Trimethoprim+Sulphamethoxazole | 15 | 93 | 1 | 7 | 12 | 100 | 0 | 0 |

Male Vs. Female culture sensitivity - P=0.0638; Not significant

Male Vs. Female culture resistance - P=0.3638; Not significant

Male sensitivity Vs. Resistance - P<0.0001; Significant

Female sensitivity Vs. Resistance - P<0.0001; Significant

3.2 Antibiotic Sensitivity in MSSA among the Gender Distribution

In the present study, we thought to investigate the sensitivity of MSSA among the gender. While cefoxitin and penicillin were found to have 100% resistance in both genders, Tetracycline, Tigecyclin, Nitrofurantoin were found to have 100% sensitive in both genders. On the contrary, rifampicin was found to have 25% and 56% sensitivity for male and female respectively. However, no statistical significance between male and female in both culture sensitivity and resistance. Although, a significant difference was noted between sensitivity and resistance in both male and female (P<0.0001) Table 3.

4. Discussion

The aim of the present study is to understand the association of wound culture and MRSA, since the wound is most commonly caused by MRSA, which is well documented already¹⁵. MRSA is a challenging issue in healthcare during the selection of antibiotic across the world. In Saudi Arabia, its prevalence keeps uprising with

significant morbidity and mortality. The present study shows 48% MRSA among the fifty-four isolates, by a study conducted the by Ghazal *et al.*, 2010 in Saudi Arabia¹⁶.

Draghi *et al.*, 2006¹⁷ and Neela *et al.*, 2008¹⁸ already reported the sensitivity of Vancomycin, Linezolid, and Teicoplanin. The present study substantiates the same result and in addition, to that the present study exploring the sensitivity of Nitrofurantoin and Mupirocin. Although Madani *et al.*, 2001¹⁹ reported vancomycin resistance is emerging problem, our study shows 100% sensitivity (Table 2).

Nagwa *et al.*, 2012²⁰ indicates variable resistance towards tetracycline, rifampicin, trimethoprim/sulphamethoxazole, levofloxacin, erythromycin, and clindamycin, etc. and the present study substantiates the same (Table 2). However, the present study indicates levofloxacin found to have 12% resistance in contrast to the result of Nagwa *et al.*, 2012²⁰ showing 66% resistance. This discrepancy mandates further study with larger samples.

Oxacillin was found to have 100% resistance in MRSA isolates and 4% resistance in MSSA isolates. Yasoka Hosaka *et al.*, 2006 already addressed this issue in their

Table 3. Association antibiotic sensitivity and methicillin resistant *Staphylococcus aureus* (n=54)

| Antibiotic | MRSA (26) | | | | MSSA (28) | | | |
|--------------------------------|-----------|-----|----|-----|-----------|-----|----|-----|
| | S | % | R | % | S | % | R | % |
| Cefoxitin | 21 | 81 | 5 | 19 | 0 | 0 | 28 | 100 |
| Penicillin | 0 | 0 | 26 | 100 | 0 | 0 | 28 | 100 |
| Oxacillin | 0 | 0 | 26 | 100 | 27 | 96 | 1 | 4 |
| Gentamicin | 20 | 77 | 6 | 23 | 28 | 100 | 0 | 0 |
| Tobramycin | 18 | 69 | 8 | 31 | 28 | 100 | 0 | 0 |
| Levofloxacin | 23 | 88 | 3 | 12 | 25 | 92 | 3 | 8 |
| Moxifloxacin | 23 | 88 | 3 | 12 | 24 | 85 | 4 | 15 |
| Erythromycin | 20 | 77 | 6 | 23 | 25 | 92 | 3 | 8 |
| Clindamycin | 20 | 77 | 6 | 23 | 26 | 93 | 2 | 7 |
| Linezolid | 26 | 100 | 0 | 0 | 28 | 100 | 0 | 0 |
| Teicoplanin | 26 | 100 | 0 | 0 | 27 | 96 | 1 | 4 |
| Vancomycin | 26 | 100 | 0 | 0 | 26 | 93 | 2 | 7 |
| Tetracycline | 19 | 73 | 7 | 27 | 28 | 100 | 0 | 0 |
| Tigecyclin | 26 | 100 | 0 | 0 | 28 | 100 | 0 | 0 |
| Fosfomicin | 25 | 96 | 1 | 4 | 28 | 100 | 0 | 0 |
| Nitrofurantoin | 26 | 100 | 0 | 0 | 28 | 100 | 0 | 0 |
| Fusidic acid | 12 | 46 | 14 | 54 | 27 | 93 | 2 | 7 |
| Mupirocin | 26 | 100 | 0 | 0 | 27 | 96 | 1 | 4 |
| Rifampicin | 6 | 23 | 20 | 77 | 7 | 26 | 21 | 74 |
| Trimethoprim+Sulphamethoxazole | 20 | 77 | 6 | 23 | 28 | 100 | 0 | 0 |

MRSA Vs. MSSA culture sensitivity - P=0.2445; Not significant

MRSA Vs. MSSA culture resistance - P=0.4215; Not significant

MRSA sensitivity Vs. Resistance - P<0.0001; Significant

Non- MRSA sensitivity Vs. Resistance - P<0.0001; Significant

earlier report that when treating OS-MRSA infections, we should take precautions because treatment with β -lactam antibiotics may cause the emergence of high-resistant MRSA (HA-MRSA type), which is attributable to the presence of the *mecA* gene. Balode *et al.*, 2013²¹ and Yao *et al.*, 2009²² reported the effectiveness of Tigecycline and Linezolid respectively on MRSA and the present study consistent with their statement.

The present study made an attempt to understand the impact of the gender on antibiotic susceptibility of MSSA and the result shows no significant difference between male and female which is in agreement with Kimberly *et al.*, 2013²³. However, due to the small size of females among (3.8%; n=1) MRSA isolates, it was impossible to compare with males. Our study claims more studies with larger sample size to address this issue.

5. Conclusion

The current study recommends Linezolid, Tigecycline and Nitrofurantoin can be used as drugs of choice in both MRSA and MSSA isolates. However, Oxacillin should not be used in MRSA isolates, additionally cefoxitin not

to be used in MSSA. The study is useful to understand antibiotic susceptibility in Tabuk, which is located in the north-west region of Saudi Arabia and also to guide the clinicians choosing empirical antibiotic treatment.

6. Acknowledgement

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7. References

1. Rubin RH. Surgical wound infection: epidemiology, pathogenesis, diagnosis and management. *BMC Infect Dis.* 2006; 6:171–2.
2. Thomas KH. Surgical wound infection: an overview. *Am J Med.* 1981; 7:712–8.
3. Elmer WK, Stephen DA, William MJ, Schreckenberger PC, Winn WC. Antimicrobial susceptibility testing. In: *Colour atlas and textbook of Diagnostic Microbiology*. 5th ed. Philadelphia: Raven Publisher; 1997. p. 69–120.
4. Sani RA, Garba, Oyewole OA. Antibiotic resistance profile

- of Gram negative bacteria isolated from surgical wounds in Minna, Bida, Kontagora and Suleja areas of Niger State. American J Med Medical Sci. 2012; 2(1):20-4.
5. Mulugeta KA, Bayeh AB. Bacteriology and antibiogram of pathogens from wound infections at Dessie Laboratory, North East Ethiopia, Tanzania. J Health Res. 2011; 13(4):1-10.
 6. Anguzu J, Olila D. Drug sensitivity patterns of bacterial isolates from septic post-operative wounds in a regional referral hospital in Uganda. Afr Health Sci. 2007; 7(3):148-54.
 7. Rajendra G, Anju A, Hari PN, Sony S. Antibiotic susceptibility pattern of bacterial isolates from wound infection in Chitwan Medical College Teaching Hospital, Chitwan, Nepal, IJBAR. 2013; 4:4.
 8. Tayfour MA, Al-Ghamdi, Al-Ghamdi, A. Surgical wound infections in King Fahad Hospital at Al-Baha. Saudi Med J. 2005; 26(8):1305-7.
 9. Percevil S, Bowler P. Understanding the effects of bacterial communities and biofilms on wound healing. 2004.
 10. Available from: <http://www.worldwidewounds.com>
 11. Jun IS, Tomoko F, Katsutoshi S, Hisami K, Haruo N, Akihiko K. Prevalence of erythromycin, tetracycline, and aminoglycoside-resistance genes in methicillin-resistant *Staphylococcus aureus* in hospitals in Tokyo and Kumamoto. Jpn J Infect Dis. 2004; 57:75-7.
 12. Gould I. MRSA in Practice. The Royal Society of Medicine Press, 2006; p. 687-8.
 13. Cox RA, Conquest C, Mallaghan C, Marples RR. A major outbreak of methicillin resistant *Staphylococcus aureus* caused by a new phage type (EMRSA-16). J Hosp Infect. 1995; 29:87-106.
 14. Benson HJ. Microbiological applications (complete version): A laboratory manual in general microbiology. 3rd ed. Dubuque, Iowa: William C. Brown Company; 1994.
 15. National Committee for Clinical Laboratory Standards (NCCLS). Performance Standards for Antimicrobial Disk Susceptibility Test-Approved Standard (Document M2-A7). 7th ed, Pennsylvania, USA: NCCLS; 2000.
 16. Padro L, Machado V, Mollerach M, Mota MI, Tuchscher LP, Gaeda P. Characteristics of community-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA) strains isolated from skin and soft-tissue infections in Uruguay. Int J Microbiology. 2009; 472126.
 17. Ghazal S, Hakawi A, Syam C. King Fahad Medical City (KFMC) MRSA prevention and control program. In: KFMC Clinical Research Symposium; 2010; Riyadh.
 18. Draghi DC, Sheehan DF, Hogan P, Sahm DF. Current antimicrobial resistance profiles among methicillin-resistant *Staphylococcus aureus* encountered in the outpatient setting. Diagn Microbiol Infect Dis. 2006; 55:129-33.
 19. Neela V, Sasikumar M, Ghaznavi GR, Zambari S, Mariana S. In vitro activities of 28 antimicrobial agents against methicillin-resistant *Staphylococcus aureus* (MRSA) from a clinical setting in Malaysia. Southeast Asian J Trop Med Public Health. 2008; 39:885-92.
 20. Madani TA, Al-Abdullah NA, Al-Sanousi AA, Ghabrah TM, Afandi SZ, Bajunid HA. Methicillin-resistant *Staphylococcus aureus* in two tertiary care centers in Jeddah, Saudi Arabia. Infect Control Hosp Epidemiol. 2001; 22:211-216.
 21. Nagwa M, El Amin, Hani S, Faidah. Methicillin-resistant *Staphylococcus aureus* in western region of Saudi Arabia: prevalence and antibiotic susceptibility pattern. Ann Saudi Med. 2012; 32(5):513-6.
 22. Balode A, Punda-Polić V, Dowzicky MJ. Antimicrobial susceptibility of Gram-negative and Gram-positive bacteria collected from countries in Eastern Europe: results from the Tigecycline Evaluation and Surveillance Trial (T.E.S.T.) 2004-2010. Int J Antimicrob Agents. 2013; 41:527-35.
 23. Yao JDC, Moellering RC Jr. Antibacterial agents. In: Murray PR, Baron EJ, Jorgensen JH, Landry ML, Pfaller MA, editors. Manual of Clinical Microbiology. 9th ed. Ankara: Atlas Kitapçılık Tic. Ltd. Sti; 2009. p.1077-1113.
 24. Kimberly AN, Heather JA, Diane LR, George RG, Lagacé-Wiens PRS. Changing epidemiology of methicillin-resistant *Staphylococcus aureus* in Canada. J Antimicrob Chemother. 2013; 68(1):47-55.