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Biocompatibility of synthetic and bio-material fusion

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This communication proposes methods to improve the biocompatibility performance of synthetic materials for biological and biological material for synthetic applications. π -cloud extension by suitable ligand–ligand/metal–ligand interactions can make the synthetic–biological fusion suitable for such applications. The judicious use of ligands for π -cloud extension can be applied to carbon transformations and target-oriented drug delivery systems. Embedded metal-centre catalysts for synthetic–biological fusion include: (i) axial coordination via bridging ligands; (ii) ligands with weak to intermediate field strength and multidenticities; (iii) design of inert complexes.

Keywords: Biocompatibility, carbon transformation, drug delivery, sequestration, synthetic–natural fusion.

SYNTHETIC and biological materials can be used for many complex transformations in carbon management and target-oriented drug delivery systems. Carboxylation and reduction are two important reactions responsible for carbon management in nature. Eight biological pathways are known for converting inorganic carbon to organic material in cell biomass: (i) reductive pentose phosphate; (ii) Hatch–Slack cycle; (iii) Crassulacean acid metabolism; (iv) reductive citric acid; (v) 3-hydoxypropionate; (vi) dicarboxylate/4-hydroxybutyrate; (vii) 3-hydroxypropionate/4-hydroxybutyrate pathway and, (viii) reductive acetyl-CoA cycle. The first three are present in plant and some prokaryotes, 4th and 5th in bacteria, 6th and 7th in archaea and 8th in bacteria and archaea.

The choice of a carrier molecule is important in targetted drug delivery because it significantly affects pharmacodynamics and pharmacokinetics of drugs. Materials like lipids, natural and synthetic polymers, carbohydrates, surfactants and dendrimers are used as drug carriers^{1–3}. The drug conjugate can be designed for improving its potential for complex π - π interactions towards the target moiety and drug^{4–7}.

Biological materials are eco-friendly, but they have limitation with regard to the proposed application as they are less durable in terms of mechanical strength and resistance to corrosion. Synthetic materials, on the other hand, have issues related to environment and biocompatibility in complex transformations. Metal complexes can serve

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Figure 1. Synthetic-natural fusion.

Pathways	Organisms	CO ₂ -fixing enzyme	Proposed complexes for chemical modification
Calvin–Benson Bassham cycle (reductive pentose phosphate cycle) ^{27,28}	Oryza sativa, Nicotiana sylvestris, Nicotiana tabacum, Gossypium hirsutum, Solanum tuberosum	Ribulose-1,5-bisphosphate carboxylase/oxygenase	Carboxylation: N-heterocyclic carbene (NHC) copper complexes; Pd(OAc) ₂ with Cs ₂ CO ₃
	Saccharomyces cerevisiae Saccharomyces cerevisiae	Transketolase Phosphoribulokinase	
Hatch–Slack cycle (dicarboxylic acid pathway) ²⁹	Flaveria trinervia	Phosphoenol pyruvate carboxylase	Reduction
Crassulacean-acid metabolism ³⁰	Saccharum officinarum Bryophyllum tubiflorum, Sedum praealtum Aptenia cordifolia	Malic anhydrase Phosphoenol pyruvate carboxylase Malic enzyme	Electrocatalytic: Iridium dihydride pincer complexes and carbene-supported copper(I)boryl
			complex; CODH-modified TiO ₂ nanoparticles
Arnon–Buchanan cycle (reductive citric acid cycle) ³¹	Chloroflexus aurantiacus Aquifex pyrophilus, Thermoproteus neutrophilus, Hydrogenobacter thermophiles	2-Oxoglutarate synthase ATP-citrate lyase	Photocatalytic: Ru(bpy) ³⁺ /triethanol amine, Ru(bpy) ²⁺ /methylviologen/ triethanolamine, Ru(bpy) ²⁺ /
	Advenella mimigardefordensis Pyrococcus sp. Clostridium nerfringens	Succinyl-CoA synthetase Fd-dependent pyruvate synthase/PFOR PFP carboxylase	nicyclam ²⁺ /ascorbic acid, FeTTP/triethylamine
Wood–Ljungdahl pathway (reductive acetyl-CoA pathway) ³²	Moorella thermoacetica Clostridium thermoaceticum, Methanobacterium formicium	Acetyl-CoA synthase Formate dehydrogenase	Bpy-2,2'-bipyridine, Cyclam-1,4,8,11- tetraazacyclotetradecane
	Clostridium thermoaceticum, Clostridium formicoacetium, Eubacterium acidaminophilum,	5,10-Methylene-H ₄ folate dehydrogenate	TTP-5,10,15,20- tetraphenylporphinato
	Acetobacterium woodi Clostridium thermoaceticum Carboxydothermus hydrogenoformans, Rhodosprillium rubum	Corrinoid CODH	
3-Hydroxypropionate ³³	Chloroflexus aurantiucus, Metallosphaera sedula	Propionyl-CoA synthase	
	Metallosphaera sedula, Sulfolobus spp.	Malonyl-CoA reductase Malyl-CoA lyase	
3-Hydroxypropionate-4 hydroxybutyrate cycle ³⁴	Metallosphaera sedula	Acetyl-CoA-propionyl CoA carboxylase	
Dicarboxylate-4-hydrobutyrate cycle ³⁵	Propionibacterium shermanii Clostridium aminobutyricum, Ignicoccus hospitalis	Methylmalonyl-CoA mutase 4-Hydroxybutyryl-CoA dehydratase	

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	Table 2.	Ligands for targeted drug delivery			
Ligands	Ligand structure	Receptor	Target	Disease	Reference
Liposomes	Ó	Doxorubicin	Squamous cell lung	Carcinoma	36
Folate	$\underset{\substack{H_{H} \leftarrow C \rightarrow 0}{H_{H}}}{\overset{Q}{\underset{\substack{H_{H} \leftarrow C \rightarrow 0}{H_{H}}}}}, \underset{\substack{H_{H} \leftarrow C_{H}}{\overset{H_{H}}{\underset{\substack{H_{H} \leftarrow 0}{H_{H}}}}, \underset{\substack{H_{H} \rightarrow 0}{\overset{H_{H}}{\underset{\substack{H_{H} \rightarrow 0}{H_{H}}}}, \underset{\substack{H_{H} \rightarrow 0}{\overset{H_{H}}{\underset{\substack{H_{H} \rightarrow 0}{H_{H}}}}, \underset{\substack{H_{H} \rightarrow 0}{\overset{H_{H}}{\underset{\substack{H_{H} \rightarrow 0}{H_{H}}}}, \underset{\substack{H_{H} \rightarrow 0}{\overset{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}{\overset{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}}, \underset{\substack{H_{H} \rightarrow 0}, \underset{\substack{H_{H} \rightarrow$	Doxorubicin	Cancer cells	Epithelial cancer, myeloid leukaemia	4
Alkylglycoside	Ho H	Sugars, alkyl and peptide moieties	Kidney membrane	Renal disorders	37
Chitosan	HO TO OT	Doxorubicin conjugates with cis-aconityl linkage, paditaxel conjugates with succinate linkage	Colon	TNBS-induced colitis	Ω.
Long-circulating (stealth) liposomes	PEG Midrophile Midrophile Midrophile	Polyethylene glycol (PEG)	Tumour cells	Cancer	38
Angiotensin-converting enzyme mAb 989		Streptavidin (SA)- biotin bridge	Luminal surface of pulmonary vasculature	Cardiovascular and pulmonary diseases	39
Fatty acids (³ H-benzoyl adduct of lauric acid)	Снсоон	Albumin or galactosylated albumin/asialoglycoprotein receptor(ASGPR)	Hepatocytes	Liver disease	40
Saccharide-poly(L-lysine) galactose, lactose, N-acetylgalactosamine	HHEORA MACOCH201043,20043 HHEOROA HHEOROA HEOROA	Hepatic receptors	Liver	Liver disease	و
Saccharide-poly(L-lysine) mannosyl and fucosyl	WHEORA MACONA MA	Reticuloendothelial system	Liver, spleen and bone marrow	Liver and spleen diseases	ى
					(Contd)

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Table 2. (Contd)					
Ligands	Ligand structure	Receptor	Target	Disease	Reference
Saccharide-poly(L-lysine) xylosyl	HHCO.R COOLOGIALSON, HCOOLUL	Hepatic receptors	Liver and lung	Liver and lung diseases	ې
Galactose and mannose	or of the second	Glycosylated carboxymethyl (CMD)-dextran	Liver, parenchymal or nonparenchymal cells	Fibrosis and cirrhosis	41
Lectin		Sugar groups, salivery pellicle	Oral cavity/salivary mucins	Gingivitis, oral candidosis, oral lesion, xerostoma	42
Nucleic acid (Aptomers)		Doxorubicin	T-cell acute lymphoblastic leukaemia, T-cell ALL cells/PSMA protein on surface of prostate cancer cells	Acute lymphoblastic, myeloblastic leukaemias, malignant lyphomes, bronchogenic	26, 43
Collagen		Antibiotics (gentamicine)	Adjancillary therapy, on-site delivery of antibacterial drugs	Treatment of prophylaxis of bone, wound healing, ophthalmic and periodontal treatment	43
Polyamine (PAMAM) folate dendrimer		Folate receptor	High-affinity folate receptor (hFR)	Ovarian tumours	44
Dendrimer	He for the second	Anti-HER2mAb	Human epidermal growth receptor-2 (HER2)	Immunotherapy	45
Heparin folate paclitaxel (HFT)	CH4,080, RO CH4,080, CO HNSO, COOR	Paclitaxel	Folate receptor (FR) expressing tumours (FR-positive human neck and head cancer cell line KB-3-1)	Human neck and head cancer	7

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as highly efficient and commonly utilized catalysts or precursors for organic and biological transformations using the principle of bioorthogonal chemistry^{8,9}. In nature, many bionuclear complexes with hydrophobic binding pockets show enhanced chemical reactivity towards the activation and transformation of small molecules such as CO_2 (ref. 10), and effect photosynthesis within a certain concentration¹¹. It is therefore an appealing strategy to make use of embedded metal centres as modified biological and synthetic catalysts or precursors. This communication proposes methods to fuse synthetic and biological complexes to design efficient and biocompatible materials. The proposed fusion has the advantages in the upcoming research in carbon management and target-oriented drug delivery using the benefits of both biological and synthetic materials (Figure 1). The methods/protocols used to design embedded metal-centre catalysts for synthetic-biological fusion include the following.

(i) Axial coordination via bridging ligands can be an important strategy to connect synthetic and biological components. Axial coordination to Ni^{II} and Zn^{II} for *trans*-III cyclams is favoured in protein complexes via bridging ligands such as phthalate and is responsible for their biological activity^{12–15}. Metal complexes are readily available for combinatorial synthesis of metal centres and ligand exchange. There are several examples of this fusion, such as insertion of symmetric metal complexes into the active site of apomyoglobin by binding to His93, which enables these new semisynthetic metalloenzymes to catalyse enantioselective sulphoxidation using the chiral protein cavity¹⁶.

(ii) Ligands with weak to intermediate field strength and multidenticities may be suitable to design pharmaceutically acceptable metal carriers with embedded metal centres. To make hexamine cobalt $[Co(NH_3)_6]^+$ biocompatible, the four NH₃ ligands are replaced by ligands such as N₂O₄ or its isoelectronic moieties¹⁷. Furthermore, ligands with chiral centres can optimally tune the activity of metal centres towards their catalytic application in a living system¹⁸; and with some exceptions, aromaticity reduces cytotoxicity¹⁹.

(iii) Metals exert structural roles, and inert complexes can be generally biocompatible. Free ion activity controls the bioavailability of metals, and complexation restricts metal activity. Similar biological activities are found in isostructural ruthenium and osmium complexes, and confirm the structural role of metals²⁰. Labile metal complexes are more bioavailable and cytotoxic compared to inert complexes. Higher metal bioavailability makes the corresponding complex less biocompatible and vice versa⁸. Complexes with heavy metals, metals from the middle of the transition series or the third row down, or rare earth elements are preferred as a synthetic complex choice.

(v) Development of multi-nuclear complexes capable of forming an adduct with biomolecules. Improved bio-

activity is found in tetranuclear ruthenium complexes compared to dinuclear complexes²¹.

Ligand choices for this purpose are planar aromatic amines, alkyl amines, iminoethers, chiral dienes, amino acids, carbohydrates, steroids, alkaloids, small peptides and their isoelectronic entities. The above strategies can provide the optimal ligand for making biocompatible catalysts in order to optimize carbon management (Table 1). Synthetic-biological fusion can solve many problems in carbon sequestration by increasing the catalytic rate and/or oxygenase activity, improving plant photosynthesis, and making synthetic carbon transformations more efficient and eco-friendly.

Targeted drug delivery increases patient compliance efficiency of pharmaceutical agents through improved biodistribution and pharmacokinetics²²⁻²⁴. Syntheticbiological fusion can be used to design target-oriented drug delivery systems⁴⁻⁷ (Table 2) by selecting appropriate π -cloud extension, such as estrogen moieties for targeting the breast, or xylylbicyclam, a potent anti-HIV agent that mobilizes stem cells (AMD3100, 'Mozobil') via targeting the 7-helix membrane receptor, CXCR4. Specific metallomacrocycle configurations can be recognized by proteins via metal coordination to specific amino acid side chains, H-bonding and hydrophobic interactions, allowing drug design optimization²⁵. Nucleic acid (A10 RNA) ligands, aptamers and Dox which bind to the surface of prostate cancer cells have been used for targeted drug delivery²⁶.

The proposed fusion has the potential for scientific merger of different therapies (like Ayurvedic and allopathic medications) in cases where there is involvement of a metal either in the target or in the drug to develop effective medications with less side effects.

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