RESEARCH COMMUNICATIONS

- Talwar, P. K. and Kacker, R. K., *Commercial Sea Fishes of India, Handbook* (ed. Director, Zoological Survey of India), Zoological Survey of India, Kolkata, 1984, p. 997.
- Froese, R. and Pauly, D. (eds), Fishbase world wide web electronic publication, 2015; <u>www.fishbase.org</u>
- Bhusari, B. V., Biology and fishery of *Pseudosciaena sina* at Ratnagiri South Maharashtra. *J. Bombay Nat. Hist. Soc.*, 1975, **72**(2), 357–367.
- Bagenal, T. B., Aspects of fish fecundity. In *Ecology of Freshwater Fish Production* (ed. Gerking, S. D.), Blackwell Scientific Publications, Oxford, UK, 1978, pp. 75–101.
- June, F. C., Spawning of Yellowfin tuna in Hawaiian waters. US Natl. Mar. Fish. Serv. Fish. Bul., 1953, 54, 47–64.
- Yuen, H. S. Y., Maturity and fecundity of big-eye tuna in the Pacific. US Fish Wildl. Serv. Spec. Sci. Rep., 1955, 150, 1–30.
- Bagenal, T. B., A short review of fish fecundity. In *The Biological Basis of Freshwater Fish Production* (ed. Gerking, S. D.), Blackwell Scientific Publications, Oxford, UK, 1967, pp. 89–111.
- Murty, V. S. and Ramalingam, P., Observations on some aspects of biology of *Johnius (Johnieops) vogleri* (Bleeker) and *Pennahia macrophthalmus* (Bleeker) in the Kakinada region. J. Mar. Biol. Assoc. India, 1986, 28(1 and 2), 57–62.
- 23. Muthiah, C., Study on the biology of *Johnieops vogleri* (Bleeker) of Bombay waters. *Indian J. Fish.*, 1982, **25**, 118–133.
- Nair, K. V. S., Food and feeding habits of *Otolithes ruber* (Schneider) at Calicut. *Indian J. Fish.*, 1979, 26(1 and 2), 133– 139.
- Goa, Daman and Diu Marine Fishing Regulation Rules, Notification 2-2-81–FSH. Government of Goa, Daman and Diu, Forest and Agriculture Department, Panaji, Goa, India, 1981, pp. 39–64.
- Dadzie, S., Vitellogenesis, oocyte maturation pattern, spawning rhythm and spawning frequency in *Otolithes ruber* (Schneider, 1801) (Sciaenidae) in the Kuwaiti waters of the Arabian Gulf. *Sci. Mar.*, 2007, **71**(2), 239–248.
- Conover, D. O., Field and laboratory assessment of patterns in fecundity of a multiple spawning fish: the Atlantic silverside, *Menidia menidia. Fish. Bull.*, 1985, 83, 331–341.
- Hunter, J. R., Feeding ecology and predation of marine fish larvae. In *Marine Fish Larvae: Morphology, Ecology and Relation to Fisheries* (ed. Lasker, R.), Washington Sea Grant Program, Washington, USA, 1981, pp. 33–71.
- Bailey, K. M. and Houde, E. D., Predation on eggs and larvae of marine fishes and the recruitment problem. *Adv. Mar. Biol.*, 1989, 25, 1–83.
- Sheaves, M., Baker, R., Nagelkerken, I. and Connolly, R. M., True value of estuarine and coastal nurseries for fish: incorporating complexity and dynamics. *Estuar. Coast.*, 2014, 38(2), 401– 414.
- Offem, B. O., Samsons, Y. A. and Omoniyi, I. T., Trophic ecology of commercially important fishes in the Cross River, Nigeria. J. Anim. Plant. Sci., 2009, 19(1), 37–44.
- 32. Murphy, H. M., Jenkins, G. P., Hamer, P. A. and Swearer, S. E., Interannual variation in larval survival of snapper (*Chrysophrys auratus*, Sparidae) is linked to diet breadth and prey availability. *Can. J. Fish. Aquat. Sci.*, 2012, **69**, 1340–1351.
- Fish trails, *Goan fish trails*, An overview of Department of Fisheries, Govt of Goa, Goa, 2015, 2, 53.
- Naqvi, S. W. A. *et al.*, Seasonal anoxia over the western Indian continental shelf. In *Indian Ocean Biogeochemical Processes and Ecological Variability* (eds Wiggert, J., Hood, R. and Naqvi, S. W. A.), American Geophysical Union, Washington DC, USA, 2009, pp. 335–345.
- 35. Fish trails, *Goan fish trails*, An overview of Department of Fisheries, Govt of Goa, Goa, 2014, **1**, 40.
- Breitburg, D. L., Hondorp, D. W., Davias, L. A. and Diaz, R. J., Hypoxia, nitrogen, and fisheries: integrating effects across local and global landscapes. *Ann. Rev. Mar. Sci.*, 2009, 1, 329–349.

- 37. Joseph, M. M. and Jayaprakash, A., Status of exploited marine fishery resources of India. CMFRI, Kochi, 2002, p. 22.
- Allen, L. G., Pondella, D. J. and Horn, M. H. (eds), *The Ecology* of Marine Fishes: California and Adjacent Waters, Berkeley and Los Angeles: University of California Press, 2006, pp. 595–610.
- 39. Wu, R. S. S., Chapter 3 Effects of hypoxia on fish reproduction and development. *Fish Physiol.*, 2009, **27**, 79–141.
- Vaquer-Sunyer, R. and Daurte, C. M., Thresholds of hypoxia for marine biodiversity. *Proc. Natl. Acad. Sci. USA*, 2008, 105(40), 15452–15457.

ACKNOWLEDGEMENT. The first author thanks the Council of Scientific and Industrial Research (CSIR), New Delhi for a fellowship supporting the Ph D programme.

Received 20 September 2017; revised accepted 9 April 2018

doi: 10.18520/cs/v115/i1/146-152

Fragilariopsis sp. bloom causes yellowish-brown waters off Alappuzha, south-central Kerala coast, India, during the mud bank-upwelling phase

R. Jyothibabu^{1,*}, N. Arunpandi¹, C. Karnan¹, L. Jagadeesan¹, T. M. Manojkumar¹, K. K. Balachandran¹ and S. W. A. Naqvi²

 ¹CSIR-National Institute of Oceanography, Regional Centre, Kochi 682 018, India
²CSIR-National Institute of Oceanography, Dona Paula, Goa 403 004, India

Mud banks (Chakara) of Kerala are calm coastal waters that form in several isolated stretches along the coast usually during the southwest monsoon (SWM) period (June-September). They are characterized by the damping of incident waves, generating localized calm sea environment conducive for fishing activities, while the high monsoon waves create hostile environment in the rest of the region. Here, we present the scientific basis of the yellowish-brown discoloration of water column that occurs off Alappuzha, Kerala annually during the peak and late SWM associated with coastal upwelling-mud bank event. The discoloured waters that occur off Alappuzha associated with these events are locally known as 'pola vellam', which is nothing but diatom blooms. In 2014, pola vellam was actually caused by the bloom of Fragilariopsis (= Fragilaria) sp.; hereafter Fragilariopsis, which was

^{*}For correspondence. (e-mail: rjyothibabu@nio.org)

widespread in the study region, even beyond the mud bank domain. This bloom feature is attributable to the nutrient enrichment associated with intense coastal upwelling that was dominant over a larger spatial extent in the study domain, including the mud bank. FlowCAM-based plankton data strongly support the above view, as the abundance, biovolume and biomass of *Fragilariopsis* had similar temporal trend both in the mud bank and non-mud bank regions. The general ecology and importance of *Fragilariopsis* bloom in the study domain, from the point of view of commercial fisheries, is also elaborated in this communication.

Keywords: Bloom, coastal upwelling, *Fragilariopsis* sp., mud bank, fisheries.

MUD bank (Chakara) of Kerala is a unique coastal oceanographic feature that occurs in several isolated parts along the Kerala coast, between Alappuzha (south) and Ponnani (north), usually during the southwest monsoon (SWM; June-September). In the mud bank, significant damping of the incident waves results in a calm sea environment conducive for fishing while the high monsoon waves create hostile environment in the rest of the region¹⁻⁶. Relatively high concentration of suspended sediments in the water column derived from a few metres thick, fluid, muddy layer close to the sea bottom is responsible for wave damping and calm sea conditions in the mud bank^{1,5,6}. Several hypotheses are in place to explain mud bank formation along the Kerala coast⁷; however, a foolproof, scientifically convincing explanation is yet to evolve. Among the several mud banks that form along the Kerala coast, the one that forms off Alappuzha has attracted special scientific and societal attention due to its consistent occurrence every year and also the rich fishery associated with it¹⁻⁴. Indeed, large stocks of planktivorous fishes (sardine, mackerel and anchovies) and shrimps are being landed on the adjacent shore every year associated with the mud bank off Alappuzha during SWM²⁻⁴.

Another significant and rather widespread oceanographic process that occurs along the southwest coast of India, having profound biophysical impact in the region, is the coastal upwelling operating during SWM⁸⁻¹³. The physico-chemical signatures of coastal upwelling include the surfacing of cool, nutrient-rich, low-oxygenated (hypoxic to anoxic) waters towards the coast; these eventually induce significantly high plankton biomass and production in the continental shelf region⁸⁻¹³. World over, the coastal upwelling areas possess immense socioeconomic significance as they represent regions of high fish availability^{14,15}. It is pertinent here to consider that the mud banks of the Kerala coast during SWM actually coexist with coastal upwelling². Therefore, it was not clear earlier whether the enhanced plankton production and high fish landings observed every year associated with the mud bank event was contributed by the mud bank itself or by coastal upwelling, which has a demon-

CURRENT SCIENCE, VOL. 115, NO. 1, 10 JULY 2018

strated biophysical impact over a large spatial extent². This enigma was investigated in an extensive fisheries study in the Alappuzha coastal upwelling-mud bank region spanning several years, which brought to light the fact that the entire area off Alappuzha, predominantly outside the mud bank region, harbours large quantities of fish^{3,4}. These studies deciphered the enigma and presented a lucid interpretation that the high fish catch landing that is apparently associated with Alappuzha mud bank is the biological manifestation of coastal upwelling^{3,4}. As proposed originally by Banse^{8,9}, the surfacing of lowoxygenated subsurface water associated with coastal upwelling tends to concentrate the fishes towards surface waters, and these high fish concentrations in the surface waters are being exploited efficiently by fishermen by operating their fishing vessels through the mud bank⁷. This implies that the high fishery landing in the adjacent shore of the mud bank is coastal upwelling-driven and not mud bank-generated³⁻⁵. In short, the mud bank, being a calm area with smooth sea surface, acts as launching/landing place and safe anchorage for fishing vessels to exploit the coastal upwelling-induced high fish stock available in the surface waters of the mud bank and its vicinity³⁻⁶. All the above-stated points strongly indicate that the high fish landing associated with Alappuzha mud bank, is no longer a puzzle. Nonetheless, it is a fact that high fish landing occurs on the shore concomitant with the Alappuzha mud bank event, even though its major role is to function as a launching/landing place and safe anchorage for fisher folks (Figure 1).

In 2014, CSIR-National Institute of Oceanography launched a multidisciplinary field programme off Alappuzha (AMPS – Alappuzha mud bank process studies), mainly to track the causes of mud bank formation and sustenance during SWM (Figure 2 a). Under AMPS, field sampling in three locations (M1–M3) was carried out from April (pre-monsoon) to September (late-SWM), which included 15 weekly samplings (22 April to 2 August) and three biweekly samplings (16 August to 20 September). Three locations off Alappuzha were sampled in



Figure 1. Typical view of mud bank as a launching/landing place and safe anchorage for fishing vessels.



Figure 2. *a*, Study area and sampling locations; *b*, *c*, trails of the boat between locations M1 and M2 that depict yellowish-brown discoloration of water column due to proliferation of *Fragilariopsis* sp., especially in the subsurface waters.

this programme that included one location in the region where mud bank forms during SWM (M2). Two other locations (M1 and M3) were considered as reference points of the mud bank location. M1 and M2 were located at 8 m depth contour, while M3 was at 13 m depth contour. The alongshore distance between M1 and M2 was 8 km, whereas the cross-shore distance between M2 and M3 was 3 km. Typically, the mud bank off Alappuzha forms during the onset of SWM (June), characterized by relatively high suspended sediments and damping of waves in a roughly semi-circular shape¹. During the present sampling period also, mud bank formed around location M2 by the early SWM (June), signified by visually discernible damping of incident waves and prevalence of calm sea environment in a semi-circular shape.

During the peak and late SWM (July and August), when mud bank was prevalent at location M2, a peculiar discolouration of water was noticed in the entire study area. This discolouration was not only found in all three sampling locations, but also the entire sailing route between the three locations (8 km alongshore between M1 and M2 and 3 km cross-shore between M2 and M3). Importantly, it was noticed that the discolouration was more prominent in the trail of the sampling boat, indicating its high concentrations in the subsurface waters (Figure 2b and c). Several sightings of such discoloured water during the course of our time-series sampling during the peak and late SWM period motivated us to examine the history of incidence of such discoloured waters in the study area. According to native fisher folks, such dis-

colouration of the water is a characteristic feature off Alappuzha during the peak and late SWM period, for which the local terminology in use is 'pola vellam/kadal kara'. Here we present the scientific basis of pola vellam in the near-shore waters off Alappuzha during the peak and late SWM of 2014, and describe the hydrographical setting that favoured such an enigmatic feature, with their possible linkages with the high fishery resources in the study domain.

The data collected using the following methods have been utilized for this study. Temporal variation in the vertical distribution of temperature and salinity in all three locations sampled was measured with a conductivity temperature depth (CTD) profiler (Seabird Electronics, USA). Surface (0.5 m) and subsurface (5 m in M1 and M2, and 8 m in M3) water samples collected using Niskin samplers were analysed for nitrate and dissolved oxygen concentrations¹⁶. Turbidity of the surface and subsurface water was analysed using a turbidity meter (Eutech TN 100). The temporal change in surface and subsurface chlorophyll a concentration was estimated using a Trilogy fluorometer (Turner designs, USA)¹⁷, with water samples collected using a Niskin sampler (Hydrobios, Germany). Water samples from the surface and subsurface were collected from all three locations and the plankton components present in the water column were qualified using a FlowCAM (Fluid Imaging Technologies, USA), which is an advanced semi-automated equipment that combines the principles of both microscopy and flow cytometry^{5,18–20}. FlowCAM is an efficient

RESEARCH COMMUNICATIONS



Figure 3. Photomicrographs of *Fragilariopsis* sp. Bloom. (*a* and *b*) light microscopy, (*c*) fluorescence microscopy and (*d*) scanning electron microscopy. (*e*, *f*) FlowCAM images of *Fragilariopsis* sp. during (*e*) pre-monsoon/early southwest monsoon and (*f*) peak/late-southwest monsoon periods. Scale bars of (*e*) and (*f*) are the same; 300 μ m FOV flow cell and 4X objective of FlowCAM were used to capture these images.

tool to estimate the biovolume of plankton components present in the water samples, in addition to the usual qualitative and numerical abundance parameters^{5,18-20}. Details of all the procedures adopted for the FlowCAM analyses under AMPS have been presented recently⁵. Unpreserved samples were brought to the laboratory in black polythene bottle in cool condition within ~2 h of collection and analysed immediately using FlowCAM. The volume of the samples analysed was chosen on the basis of the nature and abundance of the phytoplankton cells present in the samples during different seasons. The FlowCAM analyses was based on 1000 ml of water samples during the pre/early SWM, whereas only 200 ml of water samples was analysed during rest of the peak and late SWM. Prior to the analyses, water samples were prefiltered using a 300 µm bolting silk and then concentrated with a 20 µm bolting silk to 20 ml volume. This was done by concentrating the prefiltered samples by siphoning through a PVC tube till 20 ml volume remained inside the bottle. One end of the siphoning tube, which was immersed in the sample, was attached with a 20 µm bolting silk for retaining all the particles $>20 \mu m$ size at the bottom of the sample container. Analysis of each concentrated sample (20 ml) through FlowCAM took around 20 min to complete the imaging of all particles present in the sample. The quantification of phytoplankton using FlowCAM was based on a combination of 300 µm fieldof-view (FOV) flow cell and 4X objective. Area based diameter (ABD) algorithm of FlowCAM was used as it measures the biomass of complex biological particles with more accuracy than a traditional microscope²¹. In ABD algorithm of FlowCAM, diameter is measured by the number of grey-scale pixels of the binary image of particles, which is then automatically converted to a

CURRENT SCIENCE, VOL. 115, NO. 1, 10 JULY 2018

circle with the same number of pixels²¹. Subsequently, from the pixel volumes of the image, total biovolume of the individual gets generated, which is more accurate in cases of organisms with extended and protruded body parts²¹. The water samples during each sighting of the discoloured water were collected and analysed with a combination of light (Olympus IX 51), fluorescence (Olympus BX 53) and scanning electron (Neoscope, Nikon) microscopes for obtaining relevant morphological and physical characteristics of the plankton components. Every time during the sighting of the discoloured water, 5-10 Indian oil sardine (Sardinella longiceps) specimens were collected from the fish landings on the adjacent shore, which was irrespective of the sampling locations, and the gut contents were inspected qualitatively under an inverted microscope (Olympus IX 51) following standard procedure²². This was done to understand whether there is a direct linkage between high landings of oil sardine and discoloured water in the mud bank region during the peak and late SWM period.

Figure 3 presents the results of microscopic analyses of the discoloured water. It is clear that the causative organism of the discoloured water in the study area during peak and late SWM is *Fragilariopsis*, a long, chain-forming, ribbon-like diatom considered to be significant in sustaining high oil sardine fishery along the southwest coast of India²³. The oceanographic process that favoured the blooming of *Fragilariopsis* in the study domain is evident in the temporal evolution of water column temperature in all three study locations (Figure 4). The most noteworthy feature is the significant drop in temperature in the entire water column (cool waters) during the onset of SWM (June), which indicates intense upwelling occurring in the study region. This upwelling signature was well reflected

RESEARCH COMMUNICATIONS



Figure 4. Temporal variation in the vertical distribution of temperature. Intense coastal upwelling is reflected in all three locations studied, i.e. (a) M1, (b) M2 and (c) M3, as a significant drop in water column temperature from June onwards.

in the concentration of dissolved oxygen and nitrate in all three locations (Table 1). Dissolved oxygen showed a large drop, especially in the subsurface waters during SWM, whereas nitrate concentration noticeably increased in the entire water column during the period. High salinity (>30) prevailed in the area throughout the study period, with a relatively low salinity period during late SWM period associated with increased rainfall, land runoff and river influx. There was a general increase in the water column turbidity in all the locations by the onset of the SWM, which was more pronounced in the bottom waters. In the surface waters, relatively more turbidity was found at M2 during SWM due to the prevailing mud bank in the region. Based on the turbidity and its influence on the native plankton community, it can be concluded that the turbidity level in the mud bank is well below the critical level to inhibit plankton growth and sustenance⁶. Similarly, results of the size, composition, abundance and diversity aspects of plankton considered under AMPS, based on FlowCAM data, have recently been published⁵. Importantly, as evident in Table 1, there is a noticeable increase in phytoplankton biomass (chlorophyll *a*) during the three different phases of SWM compared to the premonsoon. A significant percentage of this high chlorophyll *a* stock especially during the peak and late SWM, was contributed by the large, ribbon-like chains of *Fragilariopsis* dominant during these periods.

Table 1 presents the temporal variations in abundance, biovolume and biomass of *Fragilariopsis* based on FlowCam measurements. These data reveal some

Table 1.	Physico-chemical	and	biological	parameters	in	three	locations	(M1-M3)	in	the	study	area.	Mud	bank	prevailed	in	location	M2
during the SWM period																		

Station	Parameters	Depth	Pre-monsoon (April–May)	Early-SWM (June)	Peak-SWM (July)	Late-SWM (August)
M1	Temperature (°C)	S	30.77 ± 1.21	27.45 ± 1.50	26.34 ± 1.30	26.46 ± 1.43
		SS	29.46 ± 0.75	26.82 ± 2.02	24.46 ± 0.65	25.75 ± 1.38
	Salinity	S	34.26 ± 0.64	34.71 ± 0.32	34.15 ± 0.43	32.80 ± 2.45
		SS	34.71 ± 0.42	35.01 ± 0.41	34.48 ± 0.28	34.40 ± 0.84
	Dissolved oxygen (µM)	S	190.48 ± 21.34	172.45 ± 68.90	178.65 ± 37.74	192.18 ± 20.90
		SS	153.64 ± 26.74	66.58 ± 51.53	32.12 ± 25.55	76.19 ± 58.83
	Turbidity (NTU)	S	1.66 ± 0.42	3.67 ± 0.62	5.76 ± 0.72	2.45 ± 1.06
		SS	2.42 ± 0.16	6.65 ± 2.24	11.59 ± 0.71	6.27 ± 0.86
	Nitrate (µM)	S	0.66 ± 0.94	1.18 ± 1.02	3.41 ± 2.13	1.39 ± 0.72
		SS	1.61 ± 1.76	3.65 ± 2.69	6.04 ± 3.27	3.49 ± 2.81
	Chlorophyll $a (mg m^{-3})$	S	1.77 ± 0.85	5.03 ± 2.62	9.3 ± 2.42	3.19 ± 1.84
		SS	1.76 ± 1.59	2.42 ± 1.04	5.20 ± 1.10	3.3 ± 0.49
	<i>Fragilariopsis</i> sp. abundance (ind.l ⁻¹)	S	66 ± 63	126 ± 79	966 ± 650	470 ± 156
	<i>Fragilariopsis</i> sp. biovolume (mm ³ l ⁻¹)	S	0.01 ± 0.01	0.03 ± 0.03	0.27 ± 0.12	0.23 ± 0.05
	Fragilariopsis sp. biomass (mgC l ⁻¹)	S	0.01 ± 0.01	0.02 ± 0.02	0.23 ± 0.12	0.20 ± 0.04
M2	Temperature (°C)	S	30.37 ± 1.11	27.58 ± 1.60	25.84 ± 1.43	27.06 ± 1.61
		SS	29.45 ± 0.58	26.88 ± 1.81	24.80 ± 0.95	25.89 ± 1.70
	Salinity	S	34.67 ± 0.23	33.85 ± 0.75	33.93 ± 0.89	32.93 ± 3.52
		SS	34.71 ± 0.53	34.63 ± 0.69	34.57 ± 0.30	34.10 ± 1.91
	Dissolved oxygen (µM)	S	175.06 ± 24.79	147.8 ± 0.52	199.7 ± 51.15	221.55 ± 34.39
		SS	128.34 ± 40.31	69.55 ± 7.54	34.41 ± 30.10	119.72 ± 76.25
	Turbidity (NTU)	S	1.72 ± 0.26	5.41 ± 0.76	6.74 ± 1.91	3.63 ± 1.96
		SS	3.15 ± 0.50	8.20 ± 2.60	12.85 ± 0.60	5.96 ± 1.72
	Nitrate (µM)	S	0.70 ± 0.69	4.21 ± 3.01	3.25 ± 3.19	1.15 ± 0.97
		SS	1.94 ± 1.02	5.60 ± 4.30	5.87 ± 4.40	3.91 ± 1.86
	Chlorophyll <i>a</i> (mg m ⁻³) <i>Fragilariopsis</i> sp. abundance (ind.l ⁻¹) <i>Fragilariopsis</i> sp. biovolume (mm ³ l ⁻¹)		2.66 ± 1.82	4.51 ± 2.92	10.72 ± 7.78	4.98 ± 4.51
			2.09 ± 2.19	1.66 ± 0.68	5.70 ± 1.27	3.86 ± 1.83
			84 ± 70	323 ± 120	1599 ± 930	280 ± 35
			0.02 ± 0.01	0.05 ± 0.02	0.39 ± 0.21	0.92 ± 1.2
	Fragilariopsis sp. biomass (mgC l ⁻¹)	S	0.01 ± 0.01	0.04 ± 0.02	0.35 ± 0.25	0.77 ± 1.3
M3	Temperature (°C)	S	29.98 ± 1.36	27.52 ± 0.65	26.92 ± 2.08	26.72 ± 1.57
	• • • •	SS	28.98 ± 0.76	24.33 ± 0.86	24.32 ± 0.79	25.26 ± 1.17
	Salinity	S	34.34 ± 0.65	34.29 ± 0.35	33.97 ± 0.49	32.91 ± 1.08
		SS	34.90 ± 0.40	35.02 ± 0.49	34.70 ± 0.11	35.30 ± 0.28
	Dissolved oxygen (µM)	S	181.22 ± 9.65	147 ± 12.30	225.53 ± 60.55	248.07 ± 73.72
		SS	146.80 ± 30.43	16 ± 11.13	29.94 ± 19.25	41.72 ± 25.23
	Turbidity (NTU)	S	0.61 ± 0.24	1.69 ± 0.72	3.72 ± 1.92	1.37 ± 1.31
		SS	1.71 ± 0.78	10.18 ± 0.30	9.28 ± 0.71	6.24 ± 0.74
	Nitrate (µM)	S	0.37 ± 0.47	0.54 ± 0.42	1.72 ± 1.53	1.32 ± 1.13
		SS	2.28 ± 1.41	11.26 ± 5.44	11.11 ± 2.07	11.35 ± 8.51
	Chlorophyll a (mg m ^{-3})	S	0.66 ± 0.20	7.38 ± 6.42	10.05 ± 7.63	3.97 ± 2.95
		SS	0.63 ± 0.36	2.13 ± 1.49	6.03 ± 3.01	2.82 ± 2.38
	<i>Fragilariopsis</i> sp. abundance (ind. l^{-1})	S	16 ± 20	98 ± 141	1245 ± 534	360 ± 65
	<i>Fragilariopsis</i> sp. biovolume $(mm^3 l^{-1})$	S	0.01 ± 0.01	0.04 ± 0.05	0.31 ± 0.17	0.26 ± 0.01
	Fragilariopsis sp. biomass (mgC l ⁻¹)	S	0.01 ± 0.01	0.03 ± 0.1	0.27 ± 0.15	0.25 ± 0.02

Mean \pm standard deviation values are presented. SWM, Southwest monsoon; S, Surface (0.5 m) and SS, Subsurface (5 m in M1 and M2 and 8 m in M3). ind. l^{-1} , Individuals per litre.

important ecological aspects of *Fragilariopsis* in the study domain during different hydrographical settings, as was the case during the pre-monsoon and different phases of SWM. During the pre-monsoon, when the water column was warmer and the nutrient concentration was relatively low, the abundance, biovolume, and biomass of *Fragilariopsis* was also low compared to different phases of SWM. The most striking ecological feature of *Fragilariopsis* observed in the present study is the increase in its biovolume and biomass during the peak and late

SWM, which was more pronounced than its increase in abundance during these periods. As mentioned above, the significantly high biovolume and biomass of *Fragilariopsis* observed during peak and late SWM was due to the high abundance of ribbon-like long chains, favoured by cool and nutrient-enriched upwelled waters. The FlowCam images representing the overall size of *Fragilariopsis* during the pre-monsoon/early SWM and peak/late SWM are presented in Figure 3 *e*, which depicts the above aspect. During the pre-monsoon/early SWM,

Fragilariopsis chains were composed of 2-5 fragments, while during the peak/late SWM, they appeared to be composed of tens of fragments. The long chain-forming character of diatoms under favourable growth conditions in cultured and natural populations has been noticed earlier as well, even though the factors determining the chain length or number of cells per chain in a given species have not been clearly explained²⁴. Also, from the preliminary experimental data we have generated along with AMPS sampling, the optimum solar radiation requirements of Fragilariopsis blooms along the southwest coast of India is quite low $(30-70 \ \mu E \ m^{-2} \ s^{-1})$. This could be the reason why in the present study blooms were found in less-illuminated subsurface waters⁵. Further experimental studies are, however, necessary on the above aspects.

A synthesis of the historical records of Fragilariopsis bloom along the southwest coast of India is presented below in relation to the present observations. Nair and Subrahmanyan²³ reported extensive blooms of Fragilariopsis off Calicut (Kozhikode) and presented the view that the success of the Indian oil sardine fishery is directly linked with such blooms that form every year. Subrahmanyan²⁵ has discussed the importance of Fragilariopsis in connection with the small pelagic fishes along the southwest coast of India. Devassy²⁶ also recorded blooms of Fragilariopsis along the southwest coast of India and related them with successful oil sardine fishery. Even though the above historical studies described the bloomed diatom, here we do not consider the species-level identity of the bloomed diatom in the study region. This seems to be essential considering the standard phytoplankton identification manual of Tomas²⁷, which describes F. oceanica as a cold-water species. Therefore, more careful and focused studies are required regarding the bloom-forming diatom Fragilariopsis, its



Figure 5. *a*, Indian oil sardine (*Sardinella longiceps*) specimen from the mud bank region; *b*, *c*, *Fragilariopsis* sp. in their gut content.

species-level identity and linkage with monsoon fishery along the southwest coast of India. Nonetheless, the present fish gut content data also underline that Fragilariopsis forms a major diet content of Indian oil sardine in the study domain (Figure 5). However, due to the limited data, it is currently premature to draw general conclusions regarding the linkage of Fragilariopsis stock and oil sardine fishery success along the southwest coast of India. We suggest that future research on this aspect should consider the fact that *Fragilariopsis* bloom usually occurs close to the southwest Indian coastline, as in the case of the present study and also in historical times^{23,26}. Therefore, any future research in this line should focus on near-shore waters along the southwest coast of India. A focused multidisciplinary study considering the above aspects could be useful in resolving the current uncertainty associated with the occurrence of Fragilariopsis blooms along the southwest coast of India, and its proposed linkage with the oil sardine fishery in the region.

- 1. Damodaran, R., Meio-benthos of the mud banks of Kerala coast. *Proc. Indian Acad. Sci., Sect. B*, 1972, **38**, 288–297.
- Silas, E. G., Mud banks of Kerala–Karnataka: need for integrated study. *CMFRI Bull.*, 1984, 31, 2–7.
- Regunathan, A., Mathew, K. J., Kurup, N. S. and Murty, A. V. S., Monsoon fishery and mud banks of Kerala coast. *CMFRI Bull.*, 1981, 30, 37–41.
- Regunathan, A., Mathew, K. J., Rao, D. S., Gopinathan, C. P., Kurian, N. P. and Murty, A. V. S., Fish and fisheries of the mudbanks. *CMFRI Bull.*, 1984, 31, 60–71.
- Karnan, C., Jyothibabu, R., Arunpandi, N., Jagadeesan, L., Prathihari, A., Balachandran, K. K. and Naqvi, S. W. A., Discriminating the role of coastal upwelling and mud bank in structuring the plankton size and lower level food web along the southwest coast of India. J. Mar. Syst., 2017, 172, 24–42.
- Jyothibabu, R., Balachandran, K. K., Jagadeesan, L., Karnan, C., Arunpandi, N., Naqvi, S. W. A. and Pandiyarajan, R. S., Mud bank along the southwest coast of India are not too muddy for plankton. *Sci. Rep.*, 2018, **8**, 2544.
- Murty, A. V. S., Rao, D. S., Reghunathan, A., Gopinathan, C. P. and Mathew, K. J., Ecology of mud banks-hypothesis on mud banks. *CMFRI Bull.*, 1984, **31**, 8–20.
- Banse, K., On the upwelling and bottom trawling off the southwest coast of India. J. Mar. Biol. Assoc. India, 1959, 1, 33–49.
- 9. Banse, K., Hydrography of the Arabian Sea shelf of India and Pakistan and effects on demersal fishes. *Deep-Sea Res. 1*, 1968, **15**, 45–79.
- Nair, S. R. S., Devassy, V. P. and Madhupratap, M., Blooms of phytoplankton along the west coast of India associated with nutrient enrichment and the response of zooplankton. In *Marine Coastal Eutrophication. The Response of Marine Transitional Systems to Human Impact: Problems and Perspectives for Restoration* (eds Vollenweider, R. A., Marchetti, R., Viviani, R.), Elsevier, Amsterdam, The Netherlands, 1992, pp. 819–828.
- Madhupratap, M., Shetye, S. R., Nair, K. N. V. and Nair, S. R. S., Oil sardine and Indian mackerel: their fishery, problems and coastal oceanography. *Curr. Sci.*, 1994, 66, 340–348.
- Jyothibabu, R. *et al.*, The response of microzooplankton (20– 200 μm) to coastal upwelling and summer stratification in the southeastern Arabian Sea. *Cont. Shelf Res.*, 2008, 28, 653–671.
- 13. Jyothibabu, R., Madhu, N. V., Habeebrehman, H., Jayalakshmy, K. V., Nair, K. K. C. and Achuthankutty, C. T., Re-evaluation of

'paradox of mesozooplankton' in the eastern Arabian Sea based on ship and satellite observations. *J. Mar. Syst.*, 2010, **81**, 235– 251.

- Mann, K. H., Physical oceanography, food chains, and fish stocks: a review. *ICES J. Mar. Sci.*, 1993, 50, 105–119.
- Cury, P., Bakun, A., Crawford, R. J. M., Jarre, A., Quinones, R. A., Shannon, L. J. and Verheye, H. M., Small pelagic in upwelling systems: patterns of interaction and structural changes in 'waspwaist' ecosystems. *ICES J. Mar. Sci.*, 2000, **57**, 603–618.
- Grasshoff, K., In *Methods of Seawater Analysis* (eds Grasshoff, K., Ehrhardt, M. and Kremling, K.), Verlag Chemie, Weinheim, 1983, p. 419.
- United Nations Educational, Scientific and Cultural Organization (UNESCO), Protocols for the Joint Global Ocean Flux Study (JGOFS). Core measurements. UNESCO, Paris, 29, 1994, p. 170.
- Sieracki, C. K., Sieracki, M. E. and Yentsch, C. S., An imagingin-flow system for automated analysis of marine microplankton. *Mar. Ecol. Prog. Ser.*, 1998, 168, 285–296.
- Le Bourg, B., Cornet-Barthaux, V. R., Pagano, M. and Blanchot, J., FlowCAM as a tool for studying small (80–1000 μm) metazooplankton communities. *J. Plankton Res.*, 2015, **37**, 666–670.
- Mauchline, J., Blaxter, J. H. S. and Tyler, P. A., The biology of calanoid copepods. In *Advances in Marine Biology*, Academic Press, San Diego, CA. USA, 1998, vol. 33, p. 710.
- Karnan, C., Jyothibabu, R., Manoj Kumar, T. M., Jagadeesan, L. and Arunpandi, N., On the accuracy of assessing copepod size and biovolume using FlowCAM and traditional microscopy. *Indian J. Geo-Mar. Sci.*, 2017, 46, 1261–1264.
- Zachariah, P. U. and Abdurahiman, K. P., Methods of stomach content analyses of fishes – building mass balance trophic and simulation models. Technical Notes, Central Marine Fisheries Research Institute (CMFRI), Kochi, 2004, p. 200.
- 23. Nair, R. V. and Subrahmanyan, R., The diatom, *Fragilaria oceanica* Cleve, an indicator of abundance of the Indian oil sardine, *Sardinella longiceps* Cuv. and Val. *Curr. Sci.*, 1955, **24**, 41–42.
- Takabayashi, M., Lew, K., Johnson, A., Marchi, A. L., Dugdale, R. and Wilkerson, F. P., The effect of nutrient availability and temperature on chain length of the diatom, *Skeletonema coastatum. J. Plankton Res.*, 2006, 28, 831–840.
- 25. Subrahmanyan, R., Studies on the phytoplankton of the west coast of India. *Proc. Indian Acad. Sci.*, *Sect. B*, 1959, **4**, 189–252.
- 26. Devassy, V. P., Observations on the bloom of a diatom *Fragilaria oceanica* Cleve. *Mahasagar*, 1974, **7**, 101–105.
- 27. Tomas, C. R., *Identifying Marine Phytoplankton*, Academic Press/ Harcourt Brace, San Diego, CA, USA, 1997, p. 858.

ACKNOWLEDGEMENTS. We thank the Directors of CSIR-National Institute of Oceanography and ICAR-Central Marine Fisheries Research Institute India for providing the facilities. We also thank all our colleagues who participated in the field work under hostile sea conditions to study the Alappuzha mud bank. This is NIO contribution 6185.

Received 3 August 2016; revised accepted 29 January 2018

doi: 10.18520/cs/v115/i1/152-159

AdaBoost-based long short-term memory ensemble learning approach for financial time series forecasting

Yungao Wu^{1,2,*} and Jianwei Gao¹

 ¹School of Economics and Management, North China Electric Power University, Beijing, 102206, China
²Department of Mathematical Sciences, Ordos Institution of Applied Technology, Ordos, 017000, China

A hybrid ensemble learning approach is proposed for financial time series forecasting combining AdaBoost algorithm and long short-term memory (LSTM) network. First, LSTM predictor is trained using the training samples obtained by AdaBoost algorithm. Then, AdaBoost algorithm is applied to obtain the ensemble weights of each LSTM predictor. The forecasting results of all the LSTM predictors are combined using ensemble weights to generate our final results. Four major daily exchange rate datasets and two stock market index datasets are selected for model evaluation and model comparison. The empirical study demonstrates that the proposed AdaBoost-LSTM ensemble learning approach outperform other single forecasting models and other ensemble learning approach in terms of both level forecasting accuracy and directional forecasting accuracy. This suggests that the AdaBoost-LSTM ensemble learning approach is a highly promising for financial time rates forecasting.

Keywords: AdaBoost algorithm, ensemble learning, financial time series forecasting, long short-term memory network.

GLOBAL financial markets function in a complex and dynamic manner as high noisy data volatility is routine. Many factors impact the financial market, such as economic conditions, political events, and even traders' expectations. Hence, financial time series forecasting is usually regarded as one of the most challenging tasks among time series forecasting due to the high degrees of nonlinearity and irregularity. How to accurately forecast stock and exchange rate movement is still an open question with respect to the economic and social organization of modern society.

Many common econometric and statistical models have been applied to financial time series forecasting, such as linear regression models, autoregressive integrated moving average (ARIMA) models^{1,2}, co-integration models^{3,4}, generalized autoregressive conditional heteroscedasticity (GARCH) models^{1,5}, vector auto-regression (VAR) models^{6,7} and error correction models (ECM)⁴.

^{*}For correspondence. (e-mail: wuyungao_2007@126.com)