

Control and management of cyanophycean (*Spirulina platensis*) bloom in Padmatheertham, Thiruvananthapuram, India

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The blue-green algal (BGA) bloom that appeared in Padmatheertham, the sacred pond in Sree Padmanabha Swamy Temple, Thiruvananthapuram, Kerala, India is associated with *Spirulina platensis*, a cyanobacterium rich in proteins, considered as a safe, functional food. Considering the unaesthetic appearance of the BGA bloom and its foul odour on open decomposition, various non-chemical methods were employed for its control. Several methods for nutrient remediation in the pond system were also explored. The efficacy of using decomposing rice straw to inhibit algal growth was studied. The possibility of control of BGA by stocking tilapia and filter-feeding bivalve, *Villorita cyprinoides* capable of ingesting and digesting the algae was analysed. Experimental assays carried out on *V. cyprinoides* revealed that it helped in the rapid utilization of BGA. The present study reinforces our understanding of the fundamental ecosystem services that filter-feeder communities provide to counter the invasive effects of eutrophication through consumption and assimilation.

Keywords: Bioremediation, blue-green algae, eutrophication, hypoxic, *Spirulina platensis*.

BLUE-GREEN Algae (BGA) associated harmful algal blooms (HABs) and their toxic effects are a growing concern worldwide¹⁻³. There is a wealth of research on this topic⁴⁻⁶. A report of the United Nations Intergovernmental Panel on Climate Change (IPCC) directly links HABs to climate change⁷. BGA can grow in excess, causing BGA blooms when the water is warm and if there are sufficient nutrients, viz. phosphorus and nitrogen in the water body. Run-off from fertilized agricultural areas and sewage effluents are the major external sources of phosphorus and nitrogen in the waterways. Large-scale death and decomposition of blooming algae can deplete oxygen and promote the internal origin of nutrients from the lake/reservoir sediments.

The massive blooming of an unknown alga with an unpleasant odour was seen in Padmatheertham, the pond in Sree Padmanabhaswamy temple, Thiruvananthapuram, Kerala, India, in August 2019, after the renovation work of the

pond by the Archaeology Department. Due to an unaesthetic appearance of the algal bloom that became intense during the summer months and its foul odour on open decomposition, the temple authorities sought nonchemical remedial measures to contain the menace before the 'murajapam' a 56-day traditional ceremony held every sixth year, when Jalajapam, chanting of the Vedas is performed by the pundits in the water body at the Padmatheertham pond.

Materials and methods

This study was carried out at the Padmatheertham (13,500 sq. m), one of the oldest ponds in Thiruvananthapuram situated in front of the East Nada of the Sree Padmanabhaswamy Temple. Water quality and algal abundance were monitored from August to November 2019, when the pond witnessed a rapid bloom of algae immediately after the reconstruction process (Figure 1).

Surface-water samples were collected using a Van Dorn sampler and analysed for physico-chemical parameters, viz. pH, salinity, alkalinity, hardness, free carbon dioxide (CO₂), dissolved oxygen (DO), biochemical oxygen demand (BOD), phosphate and nitrate in the waterbody. Sediment organic carbon, qualitative and quantitative assessment of algal count and chlorophyll was analysed after American Public Health Association (APHA)⁸. Transparency was measured with a Secchi disc and the pH of water was measured electrometrically using a pH meter. Salinity was measured with a salinity meter (Oakton SALT 6+) and the same was also confirmed titrimetrically according to the Mohr-Knudsen method. DO was determined by the modified Winkler method. The alkalinity of the sample and free CO₂ were estimated titrimetrically. Phosphate was estimated after Fonselius and Carlberg⁹, and nitrites and nitrates were estimated as per methods described by Mullin and Riley¹⁰ after APHA⁸. Total soil organic carbon was estimated using the Walkley and Black method¹¹, as described by Buchanan¹². The light and dark bottle method was used for measuring the primary productivity of the water body¹³.

Zooplanktons were collected using plankton net of bolting silk having a mesh aperture of 100 µm. The total count and numerical abundance of individual plankter species were

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determined. Algal counts were monitored according to the methods described by Saraceni and Ruggiu¹⁴. Cell counts were enumerated using a modified Sedgwick–Rafter cell, as recommended by Lund *et al.*¹⁵ and Frontier¹⁶. Chlorophyll analysis was done by acetone extraction method⁸.

As the temple authorities were keen on tackling the menace as soon as possible, before the traditional ceremony and wanted to adopt only non-chemical control methods, a combination of bloom control methods (BCMs) were employed for integrated control and management of the algal bloom¹⁷. Biological option of stocking fishes, viz. *Oreochromis niloticus*, GIFT breed and bivalve *Villorita cyprinoides*, capable of ingesting BGA, were also employed. GIFT breed tilapia was procured from the Marine Product Export Development Authority (MPEDA) hatchery at Kochi, Kerala and was stocked @ 5180/ha in the pond.

As part of the algal control plan, decomposing rice straw known to release algicidal exudates was utilized according to the procedure described by Islami and Filizadeh¹⁸. For this, 42 bales of paddy straw weighing 1050 kg were chopped into small pieces and were placed loosely in onion net bags in the pond at 20 cm below the surface at 10 g straw per sq. m (ref. 19). The straw bales were, safely anchored on to the side steps of the pond. Straw bundles were allowed

to rot for the entire study period so that the exudates released during the process could be utilized to inhibit the growth of algae. In order to compensate for the reduction of oxygen and limit the release of sediment-bound phosphorus that promotes sustained eutrophication, violent aeration of water was ensured by employing two 5 HP pumps fitted with a sprinkler system.

Filter-feeding black clams, *V. cyprinoides* of size 2.3 cm (7.4 g) capable of ingesting algae were stocked according to the methodology described by Smith *et al.*²⁰. For this, baby clams collected from the clam beds in Thottappally backwaters, Alappuzha, Kerala, were carefully transported live and stocked in the pond. They were deposited at 9000/ha, along the pond's submerged steps and sandy bottom slopes.

In order to assess the rate of utilization of algae and their zooremediation by the bivalve, *V. cyprinoides* feeding assays were undertaken through laboratory trials by stocking black clams in 40-litre capacity experimental tanks filled with filtered sand bed up to 20 cm depth (Figure 2). The tanks were filled up to 40 cm depth with algal-rich water collected from the temple pond with stocking density at 6 nos/l. Moderate aeration was provided in the experimental tanks and the biocontrol efficacy of the clams to consume algae by biofiltration was estimated by continually monitoring algal counts in the tanks. Replicated trials were carried out and the results were compared with control tanks without animals, both under continuous lighting and aeration. The water quality parameters and algal counts were evaluated.

Results

The algal bloom that appeared in the temple pond in August 2019 was identified as *Spirulina platensis*. With the



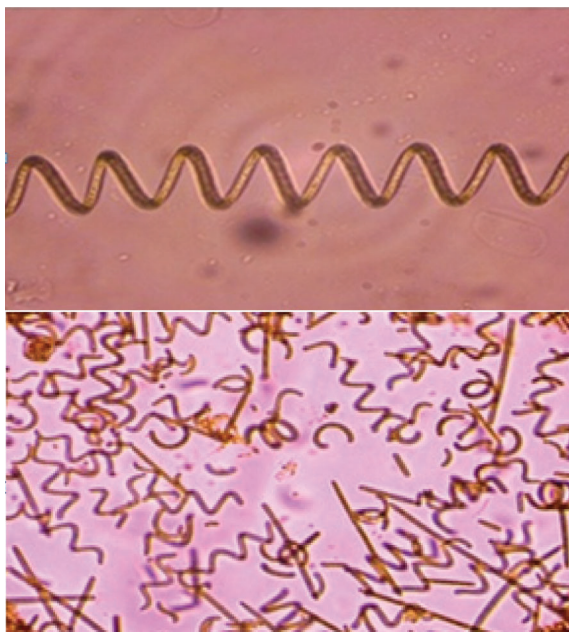
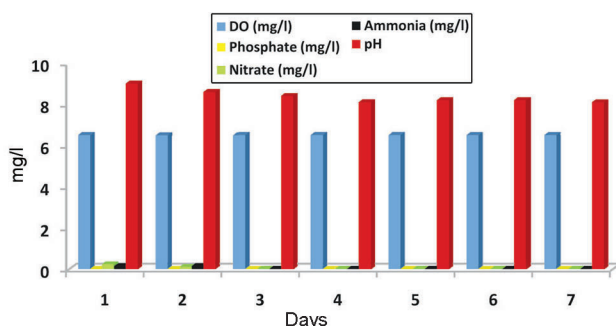
Figure 1. a, Padmatheertham pond reconstruction work. b, Blue–green algal bloom. c, Pond after removal of algal bloom.



Figure 2. Results after treatment with clam.

Table 1. Environmental parameters in Padmatheertham pond during the algal bloom period

Month (2019)	August	September	October	November
Temperature (°C)	27.5	30	29.5	31
pH	9	8.9	9	8.9
Free CO ₂ (mg/l)	–	–	–	–
Dissolved oxygen (Bottom; mg/l)	5.5 (1.2)	6.5 (1.2)	4.2 (1.0)	4.0 (0.0)
Biochemical oxygen demand (mg/l)	1.8	0	0	0
Productivity (mg C/m ³ /h ⁻¹)	468.5	–	–	–
Alkalinity (mg/l)	307.3	200	240	200
PO ₄ -P (mg/l)	1.65	0	0	0
NO ₃ -N (mg/l)	0.15	0.05	0	0
Ammonia (mg/l)	0.86	0.05	0.5	0
Transparency (cm)	5	20	2	–
Organic carbon (%)	0.47	0.28	0.26	–
<i>Spirulina platensis</i> (10 ¹⁰ /ml)	18.56	0.01	1.08	0.13

**Figure 3.** *Spirulina platensis*.**Figure 4.** Water quality changes *in vivo* tank experiments using *Villorita cyprinoides*.

overgrowth of algae, the pond surface was covered with a scum mat having an unpleasant odour (Figure 3). Thick blooms block light from reaching the ponds bottom and dead cells decompose and consume oxygen, resulting in hypoxic 'dead zones' in the pond.

The moderately warm temperature (26.25°–31°C) and the highly alkaline pond water conditions at pH 9 and high nutrient concentration favoured the growth and proliferation of *S. platensis*. Table 1 shows the water quality parameters in the pond before and during the treatment period. The air temperature varied from 26.25°C to 31°C and water temperature ranged from 27.5°C to 31°C. The water was almost turbid throughout, with low transparency ranging from 2 to 20 cm, while the pH ranged from 8.9 to 9. Free CO₂ was not detected at any time. DO in the surface water was observed to vary from 4.2 to 6.5 mg/l, while the bottom water was hypoxic with DO ranging from 1.00 to 1.2 mg/l. The BOD of surface water during the peak of bloom was 1.80 mg/l (Figure 4). The total alkalinity of the pond water varied from 200 to 307.30 mg/l (Figure 5). The gross primary productivity was very high, i.e. 468.5 mg/cm³/h⁻¹. The phosphate and nitrate concentrations were very low, ranging from nil to 1.65 and 0.05 to 0.15 mg/l respectively. Free ammonia in the pond varied from 0.05 to 0.86 mg/l, with the highest value during August when the bloom was almost at its peak. The organic carbon percentage in the pond sediments ranged from 0.26% to 0.47% by employing control measures, there was a marginal reduction in pH to 8.9 by September and the algal concentration was subsequently reduced to 0.13 × 10¹⁰ cells/ml.

The algal count of *S. platensis* during the peak bloom period was 18.56 × 10¹⁰ cells/ml, which could be gradually brought down to 0.13 × 10¹⁰ cells/ml by November 2019, with the adoption of different BCMS. The BGA was represented predominantly by *Spirulina* and occasionally by *Oscillatoria* sp. With low pH, *Oscillatoria* sp. was encountered in higher numbers. In the initial phase, no algal elements other than *Spirulina* could be observed till the subsidence of the bloom in November 2019.

The internal generation of nutrients at anaerobic conditions was limited through aeration and oxygenation and by blocking all below-ground seepage connections to the pond. As part of the integrated approaches to control the bloom, paddy straw bales were deposited, which underwent active decomposition from around 1–2 weeks. At water temperatures from 27.5°C to 30°C, the decomposition of straw was

almost complete in three months. With a gradual lowering of pH, the abundance of *S. platensis* had also drastically reduced. The biological control facilitated by stocking of fish, especially *Oreochromis mossambicus* and filter-feeding bivalve *V. cyprinoides*, also helped control the algal elements, as the stocked fish and shell-fish voraciously consumed the algal mats. The integrated approaches adopted helped control the BGA menace from the third month with a rapid reduction in algal cell concentration.

The phosphate and nitrate concentration in the pond water was found to be low, although the bloom was seemingly sustained by internal regeneration of nutrients with the decomposition of algal biomass and near hypoxic conditions at the bottom sediments. DO was very low during the entire bloom period and hypoxic concentrations (1.0–1.2 mg/l) persisted throughout the bottom waters.

In order to assess the biocontrol properties of black clam, *V. cyprinoide*, on blue-green algae *S. platensis*, laboratory based tank trial assays were carried out concurrently. These studies indicated a rapid reduction of the algal cell count from 0.12×10^{10} cells/ml to nil in seven days in the first tank trial and from 1.08×10^{10} cells/ml to nil in 4 day after stocking (DAS) in subsequent trials (Figure 6). In the control system, without clams but with aeration, there was a marginal reduction of algal density. High DO was maintained close to 6.5 mg through aeration in the laboratory assay. The algal feeding capacity of bivalve clam was perceptible in all the replicated trials, as evident from reduced algal counts.

With the adoption of integrated measures, including deposition of paddy straw and stocking of tilapia and *V. cyprinoides*, and aeration, water transparency gradually improved and the bloom almost subsided by November. The pond could be restored to a near-normal situation prior to the conduct of the traditional ceremony.

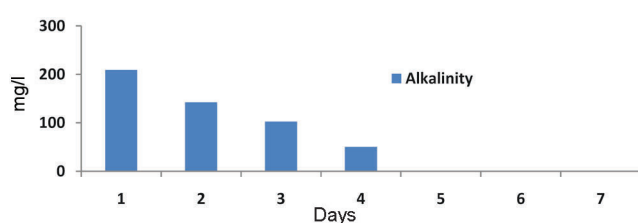


Figure 5. Alkalinity changes in temple pond.

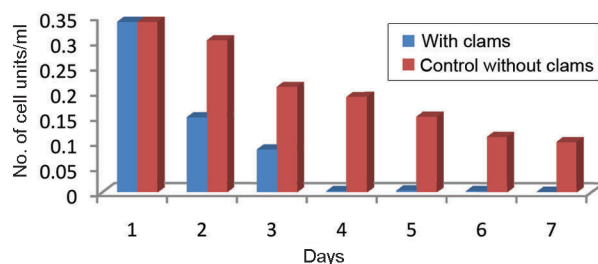


Figure 6. Bivalve feeding assay using *V. cyprinoides*.

Discussion

Chemical remediation of algal blooms is generally performed using different algaecides²¹. However, a biological option was preferred by the temple authorities in the sacred pond. The *S. platensis* bloom in the pond was apparently an outcome of elevated pH and alkalinity of water due to the use of highly alkaline lime mortar mix used for the pond reconstruction work following the Ancient and Historical Monuments and Archaeologic Conservation Procedure (AHMACP).

As observed in this study, during the peak bloom, bicarbonate ion (HCO_3^-) and carbonate ion (CO_3^{2-}) increased the pH of surface water to around pH 9–10. Such a highly alkaline environment is inimical to most other algae that rely on dissolved CO_2 compared to BGA that consume HCO_3^- . *S. platensis* thrives well in highly alkaline waters where no other microorganisms can grow²². The highest biomass and algal abundance were also noticed at pH 9, which is considered as the optimum value for survival of *S. platensis*^{23,24}.

The bloom was also contributed by high nutrient conditions due to the influence of sewage influx and urban run-off coupled with stagnant water conditions consequent to blockage of the traditional flushing system in the temple pond. The pond reconstruction works for restoration by desilting served little on the levels of reactive phosphorus, as during the renovation work, the bottom soils that were excavated and bailed out were not removed but were utilized to rebuild the pond dykes²⁵.

The breakdown of the century-old, self-flushing mechanism that brought in water from the Kochar, a tributary of the Killi River, constructed during the Travancore period also contributed to the algal bloom. However, the low concentrations of phosphate ion ($\text{PO}_4\text{-P}$) and inorganic nitrogen during the bloom are attributed to the rapid consumption of nutrients by the algae.

Evidently, with the hypoxic condition of the pond, during the algal bloom and its eventual decay, the release of sediment-bound phosphates sustained the algal growth. Aeration facilitated as part of the integrated management helped check the release of sediment-bound phosphorus to the water column²⁶. The artificial destratification due to violent aeration also promoted water circulation between the shallower and deeper layers of the water body^{27,28}. This not only helped enhance DO in the water body but also limited the availability of reactive phosphorus.

Diverse options were used to control the algal bloom in the pond. Stocking filter-feeding fish, *O. niloticus*, helped ingest and digest BGA in the experimental aquaculture system²⁵. This is a highly evolved planktonic herbivore fish with an unusually low stomach acid pH of 1.4 that help lyse BGA cells and digest them²⁹. Bivalves are a natural solution to remove particles from the water column and remediate nitrogen loads³⁰. *O. niloticus* and bivalve, *V. cyprinoides* helped control the algal bloom. Positive effects of combining two native filter-feeders, bighead carp (*Aristichthys nobilis*) and Asian clam (*Corbicula fluminea*), to control cyanobacterial

bloom have been reported earlier³¹. This is a typical case of aquatic zooremediation, wherein biotic communities are deployed to remediate eutrophied environments, similar to that reported for some species of green mussels *Perna viridis*^{32,33}.

The experimental assays on feeding responses by black clams indicate that bloom-forming algal species are foraged by *V. cyprinoides*. This indicates that healthy populations of this native filter-feeding species aid in the mitigation and prevention of HABs in estuarine systems. The present study calls for a more detailed examination on the role of indigenous epifaunal and benthic filtering communities that silently perform significant ecosystem functions^{34,35}. *V. cyprinoides* with high filtration capacity, apparently aids in the bioremediation of algae.

Use of decomposing straw, especially barley straw, for the control of algae and cyanobacteria has been a subject of research since early 1990 (refs 36–38). In the present study, we used readily available paddy straw in place of barley straw. Paddy straw is also effective in inhibiting algal growth^{39,40}. Thus, the agricultural straw application is an effective and eco-friendly method for the inhibition of algal blooms in eutrophic waters^{41–43}.

According to some researchers, as the straw decomposes and ‘rots’ under aerobic conditions, phenolic compounds such as lignin, especially oxidized phenolics are released and the inhibiting effects of straw extract are attributed to release of chemical compounds such as hydrogen peroxide⁴⁴. Some authors, however, suggest that the decomposing straw provides a carbon source for microbial growth in the carbon-limited environment and this microbial community ‘hinders’ phosphorus uptake by the algae and limits algal growth. The straw application is a preventive measure, rather than a treatment for the existing blooms as it helped facilitate a form of ‘nutrient stripping’, which reduces conditions favourable for the growth of harmful algae. The identification of rice straw as an effective material for the inhibition of BGA implies that it may be used as an environment-friendly biomaterial for controlling algal blooms in eutrophic waters.

The observation supports the view that indigenous clam *V. cyprinoides*, have a carbon sequestration role as carbon from its environment is utilized for shell formation during their various growth stages⁴⁵. The present study reveals that native filter-feeding fish and bivalve species are adapted for mitigation and prevention of harmful algal menace in estuarine systems.

Conclusion

The study indicates that *V. cyprinoides* with high filtration capacity can be employed as an agent for the bioremediation of cyanophycean algae. The positive effects of combining omnivorous fish tilapia and a filter-feeding bivalve species to control cyanobacterial bloom have been demonstrated. The identification that common paddy straw is an effective

bio material for inhibition of BGA blooms implies that it can be used as an environment-friendly biomaterial for controlling algal menace in eutrophic waters. The present study reinforces our understanding of the fundamental ecosystem services that filter-feeder communities provide by countering the adverse effects of algal eutrophication.

1. Anderson, D. M., Cembella, A. D. and Hallegraeff, G. M., Progress in understanding harmful algal blooms: paradigm shifts and new technologies for research, monitoring, and management. *Annu. Rev. Mar. Sci.*, 2012, **4**, 143–176.
2. Glibert, P. M., Harmful algal blooms in Asia: an insidious and escalating water pollution phenomenon with effects on ecological and human health. *ASIA Network Exchange, Spring 2013*, 2014, vol. 21, 1.17.
3. Gobler, C. J., Doherty, O. M., Griffith, A. W., Hattenrath, T. K., Lehmann, Y. Kang and Litaker, W., Ocean warming since 1982 has expanded the niche of toxic algal blooms in the North Atlantic and North Pacific Oceans. *Proc. Natl. Acad. Sci. USA*, 2017, **114**, 4975–4980.
4. Paerl, H. W. and Huisman, J., Blooms like it hot. *Science*, 2008, **320**(4) 57.
5. Paerl, H. W. and Huisman, J., Climate change: a catalyst for global expansion of harmful cyanobacterial blooms. *Environ. Microbiol. Rep.*, 2009, **1**, 27–37.
6. Padmakumar, K. B., Menon, N. R. and Sanjeevan, V. N., Is occurrence of harmful algal blooms in the exclusive economic zone of India on the rise? *Int. J. Oceanogr.*, 2012, **7**.
7. Pörtner, D. C. et al. (eds), *IPCC Special Report on the Ocean and Cryosphere in a Changing Climate*, Intergovernmental Panel on Climate Change, Geneva, Switzerland, 2019.
8. APHA, *Standard Methods for the Examination of Water and Waste Water*, American Public Health Association, Washington, DC, USA, 2005, p. 167.
9. Fonselius, S. H. and Carlberg, S., Determination of dissolved inorganic phosphate. In Cooperative Research Report, Series, A, No. 29, New Baltic Manual (ed. Carlberg, R.), International Council for the Exploration of the Sea, Denmark, 1972, pp. 37–43.
10. Mullin, J. D. and Riley, J. P., The spectrophotometric determination of nitrate in natural waters, with particular reference to sea water. *Anal. Chem. Acta*, 1955, **12**, 464–480.
11. Walkley, A. and Black, I. A., An examination of Degtjareff method for determining soil organic matter and a proposed modification of the chronic acid titration method. *Soil Sci.*, 1934, **37**, 29–37.
12. Buchanan, J. B., Sediments. In *Methods for the Study of Marine Benthos*, IBP Handbook No. 16 (eds Holme, N. A. and McIntyre, A. D.), Blackwell Scientific Publication, Oxford, UK, 1971, p. 3052.
13. Trivedi, R. K. and Goel, P. K., *Chemical and Biological Method for Water Pollution Studies*, Environmental Publications, Karad, 1986, p. 248.
14. Saraceni, C. and Riggio, D., Techniques for sampling water and phytoplankton. In *A Manual on Methods for Measuring Primary Production in Aquatic Environments*. IBP, Handbook No. 12 (eds Vollenweider, R. A., Talling, J. F. and Westlake, D. F.), Blackwell Scientific Publication, Oxford, UK, 1969.
15. Lund, J. W. G., Kliplin, C. and Le Cren, E. D., The inverted microscope method of estimating algal numbers and the statistical basis of estimation by counting. *Hydrobiologia*, 1958, **11**, 143–170.
16. Frontier, S., Calcul del’ erreur Sur un comptage de zooplancton. *J. Exp. Mar. Biol. Ecol.*, 1972, **8**, 121–132.
17. Newman, J., Control of Algae with Barley Straw, NERC/Centre for Ecology and Hydrology, Wallingford, UK, 2012, p. 13.
18. Islami, H. R. and Filizadeh, Y., Use of barley straw to control nuisance freshwater algae. *Am. Water Works Assoc. J.*, 2011, **103**, 111–118; doi:10.1002/j.1551-8833.2011.tb11458.

19. Newman, J. R., Report on the control of growth of *Microcystis aeruginosa* by decomposing barley straw and the development of a bankside straw digester. Long-Ashton Crop Research–Aquatic Weeds Research Unit, Reading, UK, 1994.
20. Smith, M. J., Shaffer, J. J., Koupal, K. D. and Wyatt Hoback, W., Laboratory measures of filtration by freshwater mussels: an activity to introduce biology students to an increasingly threatened group of organisms. *Bioscience*, 2012, **38**(2), 10–15.
21. Ling, Li Xian, Song, Hai-Liang, Li, Wei, Lu, Xi-Wu and Nishimura, O., An integrated ecological floating-bed employing plant, freshwater clam and biofilm carrier for purification of eutrophic water. *Ecol. Eng.*, 2010, **36**(4), 382–390.
22. Pendersen, M. F. and Hensen, P. J., Effects of high pH on the growth and survival of six marine heterotrophic protists. *Mar. Ecol. Prog. Ser.*, 2003, **260**, 33–41.
23. Fox, R. D., *Algoculture: Spirulina, Hope for the World of Hunger*, Edisud, France, 1986, p. 319.
24. Ndjouondo, G. P., Fotsop, S. D. D., Wamba, O. and Taffou, V. D., Growth, productivity and some physico-chemical factors of *Spirulina platensis* cultivation as influenced by nutrients change. *Int. J. Bot.*, 2017, **13**, 67–74.
25. Southern Regional Aquaculture Centre, Stoneville, Control of blue-green algae in aquaculture ponds. In Fifteenth Annual Progress Report, December, 2002, pp. 20–49.
26. New England Interstate Water Pollution Control Commission, *Harmful Algal Control Methods Synopses*, NEIWPCC HAB Workgroup's Control Methods, 2015, p. 28.
27. Reynolds, C. S., Wiseman, S. W., Godfrey, B. M. and Butterwick, C., Some effects of artificial mixing on the dynamics of phytoplankton populations in large limnetic enclosures. *J. Plankton. Res.*, 1983, **5**, 203–234.
28. Becker, A., Herschel, A. and Wilhelm, C., Biological effects of incomplete destratification of hypertrophic freshwater reservoir. *Hydrobiologia*, 2006, **559**, 85–100.
29. Moriarty, D. J. W., The physiology of digestion of blue-green algae in the cichlid fish. *Tilapia nilotica*. *J. Zool.*, 1973, **171**(1), 25–39.
30. Gifford, S., Dunstan, R. H., O'Connor, W., Koller, C. E. and MacFarlane, G. R., Aquatic zooremediation: deploying animals to remediate contaminated aquatic environments. *Trends Biotechnol.*, 2007, **25**(2), 60–65.
31. Shen Ruijie, Gu Xiaohong, Chen Huihui, Mao Zhigang, Qinfei, Z. and Erik, J., Combining bivalve (*Corbicula fluminea*) and filter-feeding fish (*Aristichthys nobilis*) enhances the bioremediation effect of algae: an outdoor mesocosm study. *Sci. Total Environ.*, 2020, **727**, 138692.
32. DeFranco, E., *Best Practices for Healthy Beaches and Watersheds in Maine: Potential Bioremediation Strategies for Improving Water Quality*, Marine Sea Grant Publications, USA, 2017, p. 133; https://digitalcommons.library.umaine.edu/seagrant_pub/133.
33. Rajesh, K. V., Mohamaed, K. S. and Kripa, V., Influence of algal cell concentration, salinity and body size on filtration and ingestion rate of cultivable Indian bivalves. *Indian J. Mar. Sci.*, 2001, **30**, 87–92.
34. Widdows, J., Fieth, P. and Worrall, C. M., Relationships between seston, available food and feeding activity in the common mussel *Mytilus edulis*. *Mar. Biol.*, 1979, **50**(3), 195–207.
35. Shumway, S. E., Cucci, T. L., Newell, R. C. and Yentsch, C. M., Particle selection, ingestion, and absorption in filter-feeding bivalves. *J. Exp. Mar. Biol. Ecol.*, 1985, **91**(1–2), 77–92.
36. Welch, I. M., Barrett, P. R. F., Gibson, M. T. and Ridge, I., Barley straw as an inhibitor of algal growth I: studies in the Chesterfield Canal. *J. Appl. Phycol.*, 1990, **2**(3), 231–239; doi:10.1007/BF0217-9780.
37. Barrett, P. R. F., Curnow, J. C. and Little John, J. W., The control of diatom and cyanobacterial blooms in reservoirs using barley straw. *Hydrobiologia*, 1996, **340**, 307–311.
38. Geiger, S., Henry, E., Hayes, P. and Haggard, K., Barley straw–algae control literature analysis. South Dakota State University, USA, 2005; <http://www.sdstate.edu/nrm/outreach/pond/upload/barleyalgae-control.pdf> (accessed on 27 March 2012).
39. Park, M. H., Chung, I. M., Ahmad, A., Kim, B. H. and Hwang, S. J., Growth inhibition of unicellular and colonial *Microcystis* strains (Cyanophyceae) by compounds isolated from rice (*Oryza sativa*) hulls. *Aquat. Bot.*, 2009, **90**(4), 309–314.
40. Jacob, M., Evaluating rice straw as a substitute for barley straw in inhibiting algal growth in farm ponds. Crop, Soil and Environmental Sciences Undergraduate Honors Thesis, University of Arkansas, 2019, p. 19; <https://scholarworks.uark.edu/csesuht/19>.
41. Su, W. A. H., Jia, Y., Lu, Y. and Kong, F., Effects of rice straw on the cell viability, photosynthesis, and growth of *Microcystis aeruginosa*. *Chin. J. Oceanol. Limnol.*, 2014, **32**(1), 120–129; doi:10.1007/s00343-014-3063-0.
42. Kang, P. G., Kim, B. and Mitchell, M. J., Effects of rice and rye straw extracts on the growth of a cyanobacterium, *Microcystis aeruginosa*. *Paddy Water Environ.*, 2017, **15**(3), 617–623; doi:10.1007/s10333-017-0580-4.
43. Hua, Q. *et al.*, Allelopathic effect of the rice straw aqueous extract on the growth of *Microcystis aeruginosa*. *Ecotoxicol. Environ. Saf.*, 2018, **148**, 953–959; doi:10.1016/j.ecoenv.2017.11.049.
44. Zhou, J., Inhibitory effect of decomposing barley on algal growth in water and waste water. ITSC, Reports RR-118, Illinois Sustainable Technology Centre, USA, 2010, pp. 1–30.
45. Paul, T. T., Shyam, S. S., Manoharan, V. S. and Usha, U., Identification and evaluation of ecosystem services provided by clam (*Villorita cyprinoides*) fisheries in wetland. *Indian J. Trop. Biodiver.*, 2015, **23**(1), 21–29.

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