

Diversity and role of flower visitors in onion seed production

M. Pushpalatha¹, C. S. Patil¹ and Dnyaneshwar M. Firake^{2,*}

¹Department of Entomology, Mahatma Phule Krishi Vidyapeeth, Rahuri 413 722, India

²Agricultural Entomology, ICAR-Directorate of Floricultural Research, Pune 411 036, India

Onion flowers attract a wide range of visitors, including pollinators. Many factors influence the abundance of floral visitors in onion, such as flower size, shape, colour, weather parameters and the availability of floral rewards. Quality seed production in onion (*Allium cepa* L.) is mainly driven by insect pollinators because of its protandrous nature and cross-pollination. The type of flower visitors, their numbers and the time of their visitation determine the pollination efficiency. Bees are the major pollinating agents in onions under natural conditions. However, using chemical pesticides against pests and diseases of onions during bloom decreases the diversity in and around crop fields. Anthropogenic factors like urbanization could have negative impacts on insect pollination services. Therefore, it is critical to adopt a sustainable approach to retain and conserve the existing insect pollinators and their diversity in the onion ecosystem. In this study, we review the research work done so far on the diversity and role of flower visitors and pollinators in onion seed production and the various factors affecting their survival and services in the onion ecosystem.

Keywords: Floral biology, foraging ecology, onion, pollinators, species diversity.

ONION, *Allium cepa* L. is one of the popular bulbous vegetables of Central Asia, belonging to the Liliaceae family¹. It has been India's most important commercial vegetable and condiment crop for more than 5000 years. In onion bulb production, India is next to China, with 311.29 lakh tonnes of production and 16.26 MT/ha of productivity². Maharashtra, Karnataka, Madhya Pradesh, Andhra Pradesh, Rajasthan, Tamil Nadu and Haryana are the major onion-producing regions in India. Even though the production is high, the average productivity of onion in India is much less than the global average. The main reason for this is the non-availability of quality seed material. Annually, India requires 9400 tonnes of onion seeds to cover 11.73 lakh ha area under cultivation. Nearly 40% of the country's onion seed requirement is contributed by the organized sector, while the remaining requirement is met by the farmer's own seeds³. Quality seed material in onions is interrelated with pollination, which is attributed to the number and diversity of insect visitors. Therefore, it is necessary to conserve the insect visitors for good quantity and quality of onion seeds.

Here, we review the research work done so far on the floral biology of onions, the diversity of flower visitors, the foraging ecology of pollinators and their role in onion seed production. We also discuss the effects of different factors on flower visitors in the onion ecosystem.

Floral biology of onion

Onion is a highly cross-pollinated crop due to its protandrous nature⁴. Self-pollination rarely occurs and the plant relies on insects for cross-pollination⁵. The umbelliferous inflorescence is visible at the top of the hollow green plant⁴. The umbel consists of a couple of white-coloured membranous sheaths that surround and protect the flowers. The middle of the umbel is slightly swollen. Each spathe bears an umbel, which contains 50–2000 flowers (normally 200–600) and the number of flower stalks produced per plant is also variable^{4,6}. Each flower has six stamens, six perianth lobes, three carpels and one pistil. Each carpel contains two ovules. Flowering is normally signalled by the appearance of the scape (flower stalk). The flower opening in an umbel is highly irregular and lasts about two weeks. Strongly insolated parts of the umbel usually produce open florets first¹. Thus, plants with many inflorescences can have open flowers for a month or more⁷. It may take 30 days or more for all stalks to flower⁸.

Flower pollination may occur as late as six days after opening in onions. Majority of the pollen is shed within three days after flowering begins and is completed before the stigma becomes receptive (i.e. days four and five)⁹. Anther dehiscence occurs 3–4 days before the style reaches full length and the stigma becomes receptive⁴. Anthesis to the withering of petals and anthers takes about 10 days at 18°C and about 5 days at 30°C (ref. 1). Anthers of the three inner-whorl stamens open one by one and shed their pollen before the anthers of the outer whorls open¹⁰. The nectarines at the base of the stamens produce 30–60% of the total sugar nectar^{10–12}, which attracts insects for pollination⁸. The pollen from the flower that blooms at the end of the blooming period is less viable¹³.

Diversity of flower visitors and pollinators in the onion crop ecosystem

Several flower visitors have been reported in onion flowers (Figure 1). Many factors influence the abundance of flower

*For correspondence. (e-mail: Dnyaneshwar.firake@icar.gov.in)

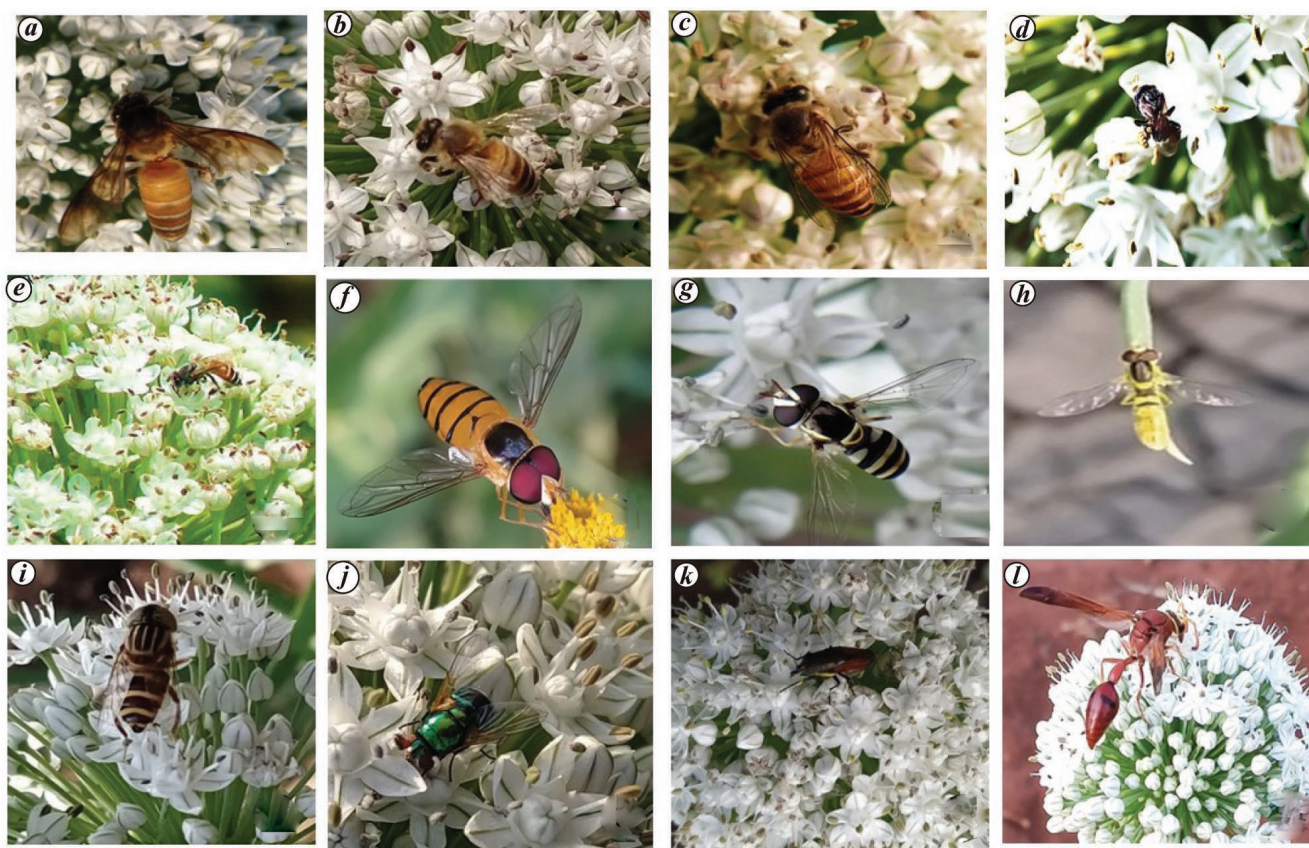


Figure 1. List of common flower visitors on onion blooms. *a*, *Apis dorsata*; *b*, *Apis mellifera*; *c*, *Apis cerana indica*; *d*, *Tetragonula* spp.; *e*, *Apis florea*; *f*, *Asarkina* spp.; *g*, *Ischidion* spp.; *h*, *Sphaerophoria* spp.; *i*, *Eristalinus aeneus*; *j*, *Lucilia serricata*; *k*, *Stomorphina* sp.; *l*, *Delta conoideum*.

visitors in onion, such as flower size, shape, colour, weather parameters and the availability of floral rewards. Pollen and nectar sources of onion blossoms attract insect visitors such as honey bees, syrphid flies, drone flies, halictid bees, butterflies, bumblebees, etc.¹⁴. Nearly 86.61% of flower visitors belong to the order Hymenoptera, followed by Diptera^{15,16}, Coleoptera and Lepidoptera (Tables 1–4). Though potassium content in the nectar deters the bees, different species of honey bees are among the major flower visitors in onion^{16–22}.

The visitation rate of Italian bee *Apis mellifera* has been reported as 5.48 bees/m²/min (ref. 23) and 12 visits/min (ref. 19) in onion. Rock bee, *Apis dorsata* is the major visitor in mountainous areas of the Indian subcontinent^{14,24}. Indian bee species *Apis cerana* and *Apis florea*, with visitation rates of 4.70 and 3.54 bees/m²/min respectively, and *Trigona irridipennis*^{25,26} are also important flower visitors in the onion ecosystem^{23,27}.

Syrphids like *Erisyrphus balteatus*, *Syrphus* sp.¹⁸ and dipterans like *Calliphora vomitaria*, *Musca domestica*²², *Ameigilla calens*, *Lipotrichus collaris*¹⁶, *Lipotrichus illustris*¹⁸, *Lasioglossum malachusum*, *Lasioglossum pauxillum*¹⁹ and horse flies²⁸ tend to visit onion flowers most frequently. The visitors were low in number during the initial flower-

ing days and significantly increased during full bloom²³. The flower visitation rate of *E. balteatus* has been recorded as 14 ± 4.61 flies/m²/min (ref. 14). It is known that flower visitors have an ecological threshold level below which no activity occurs, which varies between and within species depending on the level of adaptation of a species to a particular habitat²⁹. The flight activity of a pollinator may vary depending on the time of day and other weather factors like temperature, sunshine, wind velocity, rainfall and humidity^{18,30}.

From 6.00 am till 6.00 pm, onion flower visitors begin collecting floral rewards, with peak visitation from 10.00 am to 4.00 pm (refs 31, 32). The ideal temperature for optimum foraging activity is between 25°C and 30°C. An increase in temperature above the threshold and a decrease in relative humidity tend to have a negative impact on the foraging activity of flower visitors¹⁴. *A. mellifera* and syrphid flies have their peak foraging time in the morning (8.00–10.00 h) (ref. 24), whereas the peak foraging activity of *A. cerana*, *A. dorsata* and *A. florea* occurs between 10.00 and 14.00 h (ref. 14). The optimal conditions for bee flight and foraging are wind speed of less than 25 km/h, temperature of more than 21°C (70°F), humidity less than 75% and few or no clouds³³. In particular,

Table 1. List of major hymenopteran flower visitors in onion

Scientific name	Family	Geographical location
<i>Apis mellifera</i> Linnaeus	Apidae	India (Karnataka, West Bengal and Maharashtra) ^{15,18,45} , New Zealand ¹⁷ , Pakistan ^{14,31} , Cameroon ¹⁶ , Egypt ¹⁸ , Algeria ¹⁹
<i>Apis dorsata</i> Fabricius	Apidae	India (Karnataka, West Bengal, Himachal Pradesh and Maharashtra) ^{15,23,24,35,44} , Pakistan ^{14,31,34}
<i>Apis cerana</i> Fabricius	Apidae	India (Karnataka, West Bengal and Maharashtra) ^{23,34,65-67} , Australia ⁶⁸
<i>Apis florea</i> Fabricius	Apidae	India (Karnataka, West Bengal, Himachal Pradesh and Maharashtra) ^{15,23,35,36,44} , Pakistan ^{14,34} , Australia ⁶⁵
<i>Trigona iridipennis</i> Smith	Apidae	India (Karnataka and Maharashtra) ^{23,35,36} , Australia ⁶⁵
<i>Amegilla calens</i> (Lepelletier)	Apidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Xylocopa olivacea</i> Fabricius	Apidae	Pakistan ¹⁴ , Cameroon ¹⁶ , India (West Bengal) ⁴⁴
<i>Xylocopa inconstans</i> Smith	Apidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Xylocopa nigrita</i> (Fabricius)	Apidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Xylocopa fenestrae</i> (Fabricius)	Apidae	India (Karnataka) ^{23,36}
<i>Xylocopa pubescens</i> Spinola	Apidae	Egypt ¹⁸
<i>Bombus</i> spp.	Apidae	New Zealand ¹⁷ , Algeria ¹⁹
<i>Lipotriches collaris</i> (Vachal)	Halictidae	Pakistan ¹⁴ , New Zealand ¹⁶ , Cameroon ¹⁷
<i>Lipotriches azarensis</i> (Cockerell)	Halictidae	Pakistan ¹⁴ , New Zealand ¹⁶ , Cameroon ¹⁷
<i>Lasioglossum atricrum</i> (Vachal)	Halictidae	Pakistan ¹⁴ , New Zealand ¹⁶ , Cameroon ¹⁷
<i>Lasioglossum pauxillum</i> (Schenck)	Halictidae	Algeria ¹⁹
<i>Sphecodes</i> spp.	Halictidae	Algeria ¹⁹
<i>Lasioglossum saegeri</i> Pauly	Halictidae	Pakistan ¹⁴ , New Zealand ¹⁶ , Cameroon ¹⁷
<i>Halictus farinosus</i> Smith	Halictidae	Utah ¹⁹
<i>Belanogaster juncea</i> (Fabricius)	Vespidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Vespula vulgaris</i> Linnaeus	Vespidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Vespa cincta</i> Fabricius	Vespidae	India (West Bengal) ⁴⁴
<i>Eumenus</i> spp.	Vespidae	India (Karnataka) ³⁶
<i>Componatus flavimarginatus</i> (Mayr)	Formicidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Componatus compressus</i> (Fabricius)	Formicidae	India (Karnataka) ^{23,36}
<i>Oecophylla smaragdina</i> Fabricius	Formicidae	India (Karnataka) ^{23,36}
<i>Megachile bituberulata</i> Ritsema	Meghachilidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Chalicodoma rufipes</i> (Fabricius)	Meghachilidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Osmia rufa</i> (Linnaeus)	Meghachilidae	Wellebourne ³⁰ , USA (Utah) ^{41,42,69} , England ⁴³
<i>Ammophila</i> spp.	Sphecidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Philanthus triangulum</i> (Fabricius)	Sphecidae	Pakistan ¹⁴ , Cameroon ¹⁶

Table 2. List of major dipteran flower visitors in onion

Scientific name	Family	Geographical location
<i>Erisyrphus balteatus</i> (De Geer)	Syrphidae	Pakistan ^{14,34} , Cameroon ¹⁶ , India (West Bengal and Himachal Pradesh) ^{42,44}
<i>Metasyrphus conferator</i> (Wiedemann)	Syrphidae	India (Himachal Pradesh) ⁴²
<i>Sphaerophoria scripta</i> (Linnaeus)	Syrphidae	India (West Bengal) ⁴⁴ , Pakistan ^{14,34}
<i>Eupodes corolla</i> (Fabricius)	Syrphidae	Pakistan ¹⁴
<i>Mesembrius bengalensis</i> (Wiedemann)	Syrphidae	Pakistan ¹⁴
<i>Eristalinus aeneus</i> (Scopoli)	Syrphidae	Pakistan ¹⁴
<i>Chrysotoxum intermedium</i> Meigen	Syrphidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Paragus borbonicus</i> Macquart	Syrphidae	Pakistan ¹⁴ , Cameroon ¹⁶ , India (Karnataka) ^{23,36}
<i>Eristalis</i> spp.	Syrphidae	Wellebourne ³⁰ , India (Maharashtra) ³⁵ , USA (Utah) ^{41,42,69} , England ⁴³
<i>Musca domestica</i> Linnaeus	Muscidae	India (Karnataka, West Bengal and Maharashtra) ^{15,23,35,36,44} , Pakistan ^{14,34} , Cameroon ¹⁶ , Egypt ¹⁸ , Australia ¹⁷ , Israel ²⁹ , Washington ⁶⁷
<i>Chrysomya chloropyga</i> (Wiedemann)	Calliphoridae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Calliphora</i> spp.	Calliphoridae	Pakistan ^{14,34} , Egypt ¹⁸ , Wellebourne ³⁰ , USA (Utah) ^{41,42,69} , England ⁴³
<i>Lucilia</i> spp.	Calliphoridae	New Zealand ¹⁵ , Egypt ¹⁸ , Wellebourne ³⁰ , USA (Utah) ^{41,42,69} , England ⁴³ , Israel ⁶⁷ , The Netherlands ⁶⁶ , Australia ⁶⁸
<i>Chrysomya bezziana</i> (Villeneuve)	Calliphoridae	India (West Bengal) ⁴⁴
<i>Sarcophaga</i> sp.	Sarcophagidae	Pakistan ^{14,34} , India (West Bengal) ⁴⁴ , Egypt ¹⁸
<i>Anthomyia punctipennis</i> Wiedemann	Anthomyidae	Australia ^{17,68}
<i>Anthomyia punctipennis</i> Wiedemann	Anthomyidae	Australia ^{17,68}

A. dorsata requires a temperature of 16°C, relative humidity of 74%, light intensity of 600 lx and solar radiation of 10 mW/cm²; *A. mellifera* requires 16°C, 75% relative hu-

midity, 800 lx light intensity and 10 mW/cm² solar radiation; *A. cerana* needs 15.5°C, 76% relative humidity, 600 lx light intensity and 9 mW/cm² solar radiation, and *A. florea*

Table 3. List of major coleopteran flower visitors in onion

Scientific name	Family	Geographical location
<i>Aulucophora faevicollis</i> (Lucas)	Chrysomelidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Aulucophora abdomanalis</i> (Fabricius)	Chrysomelidae	India (Karnataka) ^{23,36}
<i>Monolepta signata</i> (Oliver)	Chrysomelidae	India (Karnataka) ^{23,36}
<i>Eurema</i> spp.	Chrysomelidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Illeus cinctus</i> Fabricius	Coccinellidae	India (Karnataka) ^{23,36}
<i>Coccinella</i> spp.	Coccinellidae	India (Karnataka) ¹⁵
<i>Coryna</i> spp.	Meloidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Mylabris</i> spp.	Meloidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Pachnoda interrupta</i> (Olivier)	Cetonidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Chrysoperla cornea</i> Stephens	Chrysopidae	Egypt ¹⁸ , India (Karnataka) ^{23,36}
<i>Anoplocnemis curvipes</i> (Fabricius)	Pentatomidae	Cameroon ¹⁶
<i>Nezara</i> spp.	Miridae	India (Karnataka) ¹⁵ , Egypt ¹⁸

Table 4. List of major lepidopteran flower visitors in onion

Scientific name	Family	Geographical location
<i>Peiris</i> spp. Schrank	Pieridae	India (Karnataka and Maharashtra) ^{15,35}
<i>Venessa cardui</i> (Linnaeus)	Pieridae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Catopsilia florelia</i> (Fabricius)	Peiridae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Danaus chrysippus</i> (Linnaeus)	Nymphalidae	India (Karnataka and Maharashtra) ^{15,35} , Pakistan ¹⁴ , Cameroon ¹⁶
<i>Danaus plexippus</i> (Linnaeus)	Nymphalidae	India (Karnataka) ¹⁵
<i>Hypolimnas misippus</i> (Linnaeus)	Nymphalidae	Pakistan ¹⁴ , Cameroon ¹⁶
<i>Papilio demoleus</i> Linnaeus	Papilionidae	India (Karnataka) ²³

needs 18.5°C temperature, 64% relative humidity, 1700 lx light intensity, and 20 mW/cm² solar radiation.

Foraging ecology of major pollinators visiting onion flowers

Foraging behaviour and population dynamics alone are not good indicators of any flower visitor as an effective pollinator. It is difficult to determine whether a flower visitor is an effective pollinator based solely on the above two factors. There are other factors that may play a role, including size and shape of the insect, whether it is thirsty for nectar or pollen, and whether it has the chance to come into contact with the stigmas of the flowers and deposit pollen on them. Considering these facts, *T. irridipennis*⁶ and honey bees of genus *Apis*, viz. *A. dorsata*^{23,24,34}, *A. cerana indica*^{35,36}, *A. mellifera*^{15,18} and *A. florea*^{34,37}, and syrphids²⁴ are reported to be effective pollinators in the onion ecosystem^{20,24,32,37} because of their body size, morphology and short tongue^{4,21}. Pollen gatherers in onion are very low or few and the majority are nectar gatherers. It is the small size of the anthers of the flowers which makes it tedious for pollen collection. The flowers get pollinated when the visitors dust-off the pollen smeared on the body while collecting nectar. For collecting nectar, they tend to visit the middle portion of the umbels rather than the top or bottom³⁸.

A. dorsata being the most efficient pollinator^{21,24,37,39} has an average visitation rate of 5.24 bees/m²/5 min (ref. 23), visiting an average of 3 umbels/min (ref. 35). A single bee tends to visit an average of 7.5 flower/umbel during a

single visit²⁴ with a time of 2.9 ± 0.52 sec on every flower³⁶ and 17.83 sec on individual umbels³⁵. Higher onion seed yield can be seen when *A. dorsata* is engaged in pollination services (340.7 g/100 umbels)⁴⁰. *A. cerana indica* has an average visitation rate of 2.93 bees/m²/5 min (ref. 23), with a single bee visiting 3.17 umbels/min (ref. 35) covering 5.42 flowers/umbel in a single visit²⁴ and spending 7.33 ± 2.41 sec/flower (ref. 36).

A single *A. mellifera* bee can visit an average of 21.64 flowers/min (ref. 21) with a minimum of 0.81 bees/m²/min (refs 15, 23). The population density of *A. mellifera* in the field sometimes reaches up to 1.78/plant/min (44.67 bees/25 plants/min) (ref. 18). *A. florea* has an average visitation rate of 1.79 bees/m²/min (ref. 23) with 18.83 sec spent per umbel³⁵ and 12.5 ± 0.40 sec stay time on each flower of an umbel³⁶. *Tetragonula* spp. has an average stay time of 5.33 ± 0.73 sec/flower (ref. 36) with a maximum of 27.5 sec spent on individual umbels³⁵. Syrphids are also efficient and major pollinators in the onion ecosystem, with a stay time of 8.8 sec/flower (ref. 24).

Honey-bee pollination studies in onion (excluding other flower visitors)

Many studies have been conducted on onions under caged conditions, allowing only single species of pollinators to check their efficiency in seed production. The honey bees and flies like *C. vomitaria*, *C. vicina*, *L. caesar*, *L. sericata* (Diptera: Calliphoridae), *M. domestica* (Diptera: Muscidae), *E. balteatus*, *E. aeneus* (Diptera: Syrphidae) and the

solitary bee *Osmia rufa* (Hymenoptera: Megachilidae) proved to be the better pollinators under caged conditions^{30,41-43} with a good seed set of more than 87.79% in *A. mellifera*-pollinated crops¹⁶. Even the seeds obtained from umbels pollinated by individual pollinators tend to have better germination, seedling emergence, survival and crossing rate³⁰. For example, the umbels pollinated with only *A. mellifera* recorded 80.84% seeds/fruit (ref. 21), seed yield of 457.72 kg/ha, germination of 76%, and test weight of 3.23 g/100 seeds, which is significantly superior over flower visitor-excluded crops (seed yield of 5.91 kg/ha, germination of 14% and test weight of 2.67 g/100 seeds)⁴⁴.

Some studies reported that flies and honey bees are equivalent in their suitability as pollinators for caged onion³⁰. Few studies suggested that calliphorid flies, *Eristalis* flies and *M. domestica* are more suitable than honey bees for pollinating onion in cages^{42,43}. To support this, an experiment was conducted where *M. domestica* and *C. vicina* pollinated crops recorded 87.8% and 85.5% germination respectively, and it was observed that an increased supply of flies increased the yield²⁹.

Effect of pollinators on seed yield of onion

If onion blooms are not visited by insects, they will not generate excellent seeds. The scarcity of natural pollinators seriously hinders seed production, especially in India's mountainous regions²⁴. Pollinators are crucial for onion seed production. Compared to bagged plants, open-pollinated plants show a 72–79% increase in seed setting⁴⁵. According to some reports, open pollination results in a 61.8% seed setting rate compared to 1.5% for self-pollinated onion crops⁴⁰. In open-pollinated plots, seed yield per bulb will be larger (5.74 g/flower) than in caged plots (1.29 g/flower)²⁵. When compared to the yields of open fields with honey-bee colonies, the number and weight of onion seeds per flower from the self-pollinated plant are low⁴⁶.

In comparison to self-pollinated plants, plots with induced bee pollination have higher seed set⁴⁷, maximum germination²⁴, and shoot and root length³². When onion blooms are pollinated by honey bees, the weight of 1000 seeds, germination capacity and energy are all high⁴⁸. Bee pollination boosts seed output from 175% (ref. 49) to 1000% (ref. 45). It was found that honey-bee pollination resulted in a 10- to 11-fold increase in seed output³³. Bees boost onion seed production by 8–10 times; therefore, seed production plots should include roughly two colonies per acre⁵⁰.

Effect of pesticide usage on pollinators visiting onion flowers

Although pesticides are often required to control pests attacking onions during the flowering stage, these have undesirable effects on the health and foraging visits of the pollinators (Table 5). Insecticides are known to repel or kill honey

bees and lower crop yield²⁷. Systemic insecticides, which are used for seed coating, translocate from the roots to the entire plant, including the flowers and may harm non-target pollinators. The susceptibility of the pollinators and other non-target organisms to the pesticides is mainly associated with the chemical structure of the pesticides rather than the body mass of the pollinators⁵¹. Pesticides like fipronil, imidacloprid, thiamethoxam, dimethoate, mancozeb, captan and azoxystrobin are some of the widely used chemicals in the onion ecosystem, which have been proven to be highly toxic to pollinators.

Fipronil is one of the important insecticides used to manage thrips of onion, to which majority of the pollinators show high susceptibility. Exposure to pollinators like *Melipona scutellaris*, *A. mellifera* and *A. cerana indica* causes reduced flight velocity, lethargy^{52,53}, locomotion difficulty, olfactory and memory learning impairment⁵⁴, loss of ability to learn and memorize, and causes the paralysis of wings, legs and digestive system, hyperexcitation accompanied by the release of alarm pheromones and individual attacks⁵⁵, genetic instability, imbalance in reactive oxygen species⁵⁶ and antioxidants⁵⁷. Up- and down-regulation in protein synthesis, which are associated with pathogen susceptibility, apoptosis, visual impairment, ischaemia and brain degeneration, have also been reported in *A. mellifera* exposed to fipronil⁵⁸. Decreased secretory cell height and reduced reservoir volume of the mandibular glands, and decreased royal jelly secretion have also been observed in nurse bees⁵³.

Neonicotinoids like imidacloprid, thiacloprid, acetamiprid and thiamethoxam are reported to have harmful effects like impairment in brood development, cognitive and neurological impairment, reduced adult longevity, immunity suppression, abnormal foraging behaviour with delayed trips and a negative impact on the feeding and learning behaviour in honey bees, *Osmia lignaria* and *Bombus* species⁵⁹. The reduced life expectancy of *Bombus impatiens*, decline in nest numbers of *Megachile rotundata* up to 50%, memory loss and decline in navigation skills are also reported⁶⁰. Organophosphate compounds like dimethoate are reported to affect the olfactory and learning performances in *A. mellifera*⁶⁰.

Fungicides like amistar and azoxystrobin are known to affect the reproductive output of bumblebee colonies with reduced fecundity, the number of workers and sexuals, weight, and low foraging activity⁶¹. Fungicide captan used for seed treatment inhibits cell division and is acutely toxic to *O. lignaria* at field-relevant concentrations⁶². It can also enhance larval mortality and developmental deformities in honey bees^{63,64}.

Conclusion

The literature indicates that pollinators are essential for onion seed production. Cross-pollination occurs in majority

Table 5. Common pesticides used in onion and their effects on the pollinators

Insecticides	Effects on pollinators
Imidacloprid	<ul style="list-style-type: none"> Abnormalities or death during development⁷⁰, decreased foraging activity and negative effects on olfactory learned discrimination task⁷¹, impaired olfactory learning and memory formation^{72,73}, adverse effect on foraging behaviour^{74,75}, cognition/neurological impairment⁷¹, increased AChE activity⁷⁶ and decreased survival⁷³, increased susceptibility of the colony to microsporidia <i>Nosema</i>⁷⁷, increased incidence of colony collapse disorder^{78,79} in <i>Apis mellifera</i>. Reduction in feeding rate⁸⁰, reduction in mean daily locomotory activity⁷⁹, reduction in queen survival, worker movement, colony consumption and colony weight^{81,82}, decreased birth rates and increased death rates in the colony⁸³, impairment in pollen foraging efficiency⁸⁴ in bumble bees, <i>Bombus</i> spp. Sub-lethal effects on larval development and longer developmental time for <i>Osmia lignaria</i>⁸⁵. Negatively affects the development of mushroom bodies in the brain^{86,87} and impairs the walking behaviour of newly emerged adult workers for stingless bees (Hymenoptera: Apidae: Meliponinae)⁸⁸.
Thiacloprid	<ul style="list-style-type: none"> Interference with navigation^{89,90}, immune suppression with <i>Nosema</i>⁹¹ and additive interaction with black queen cell virus leading to increased larval mortality in <i>A. mellifera</i>⁹⁰.
Acetamiprid	<ul style="list-style-type: none"> Impaired long-term retention of olfactory learning in <i>A. mellifera</i>⁹².
Thiamethoxam	<ul style="list-style-type: none"> Foraging difficulty⁹³, decreased colony performance and productivity, decelerated colony growth⁹⁰, behaviour and locomotive impairment⁹⁴, malpighian tubules show pronounced alterations for Africanized <i>A. mellifera</i>⁹⁵. Fewer total brood cells, higher offspring mortality, and male-biased offspring sex ratio for solitary bee <i>Osmia bicornis</i> (red mason bee)⁹⁰.
Clothianidin	<ul style="list-style-type: none"> Decreased colony performance and productivity, decelerated colony growth⁷¹, behaviour and locomotive impairment⁹⁰, cognition/neurological impairment⁷¹ in <i>A. mellifera</i>. Reduction in queen survival, worker movement, colony consumption and colony weight⁸¹, reduced foraging activity, increased worker mortality, delayed weight gain and no new queens produced⁹⁶ in <i>Bombus</i> spp. Fewer total brood cells, higher offspring mortality, and male-biased offspring sex ratio for solitary bee <i>O. bicornis</i> (red mason bee)⁹⁰.
Fipronil	<ul style="list-style-type: none"> Reduced flight velocity, lethargy^{38,53}, motor and locomotion difficulty, olfactory and memory learning impairment⁵⁴, reduced learning and memory capacities leading to paralysis of wings, legs and digestive system, hyperexcitation accompanied by emission of alarm pheromone thus leading to individual attack⁵⁵, genetic instability, imbalance in reactive oxygen species⁵⁶ and antioxidants⁵⁷ in <i>A. mellifera</i> and <i>Apis cerana indica</i>. Up- and down-regulation in protein synthesis which is related to pathogen susceptibility, apoptosis, ischaemia, visual impairment and brain degeneration in <i>A. mellifera</i>⁶⁰. Decreased secretory cell height of mandibular glands, reduced reservoir volume of mandibular glands and decreased royal jelly secretion in nurse bees of <i>A. mellifera</i>⁵³.
Dimethoate	<ul style="list-style-type: none"> Affects the olfactory and learning performances in <i>A. mellifera</i>⁶¹.
Captan	<ul style="list-style-type: none"> Inhibits cell division and is acutely toxic to <i>O. lignaria</i> at field relevant concentrations⁶³ and can induce larval mortality and developmental malformations in honey bees^{64,71}.
Mancozeb, azoxystrobin	<ul style="list-style-type: none"> Affect the reproductive output of bumble-bee colonies with reduced number of eggs laid, reduced number of workers and sexuals, reduced weight and low foraging activity⁶².

of crops, including onion and insects play a key role in this process. Both pollinating and nectar-gathering insects are attracted to onion inflorescence. When the flowers are pollinated by insects, the quantity and quality of the seeds increase in value. Pollinators are attracted to flowers because of their colour, fragrance, form, pollen and nectar. However, they require a certain ecological threshold for normal functioning, below which no activity occurs. Indiscriminate use of pesticides affects the survival and services of pollinators. Studies on bee-friendly pesticides are needed in onion crop agroecosystems. Detailed studies are also required on this reveals the scope for conserving flower visitors in the onion crop ecosystem.

Conflict of interest: The authors declare that there is no conflict of interest.

1. Brewster, J. L., Flowering and seed production. In *Onions and other Vegetable Alliums*, CAB International, United Kingdom, 1994, pp. 122–145.

2. Anon., Agricultural development in India. Crop growth statistics. Indiastat, 2022; <http://www.indiastat.com/data/agriculture>

3. Anon., Seed production – onion. Directorate of Onion and Garlic Research, Pune, 2022; https://dogr.icar.gov.in/index.php?option=com_content&view=article&id=55&Itemid=155&lang=en

4. Kavitha, S. J. and Rami Reddy, P. V., Floral biology and pollination ecology of onion (*Allium cepa* L.). *J. Pharmacogn. Phytochem.*, 2018, 7, 2081–2084.

5. Karuppaiah, V., Soumia, P. S., Thangasamy, A. and Singh, M., Tapping insect pollinators for quality seed production in onion. *Indian Hortic.*, 2017, 44–45.

6. Devi, S., Gulati, R., Tehri, K. and Poonia, A., The pollination biology of onion (*Allium cepa* L.) – a review. *Agric. Rev.*, 2015, 36, 1–13.

7. Gaviola, J. C., Technical sheet, production of onion seeds (*Allium cepa* L.), INTA-EEA the Consultation, 2007, pp. 1–18.

8. McGregor, S. E., *Insect Pollination of Cultivated Crop Plants*, Agricultural Handbook No. 496. Agricultural Research Service, U.S. Dept. Agric., 1976, pp. 268–273.

9. Moll, R. H., Receptivity of the individual onion flower and some factors affecting its duration. *Am. Soc. Hortic. Sci. Proc.*, 1952, 64, 399–404.

10. Hagler, J. R. and Robert, J., Basic aspects of onion pollination. Ph.D. thesis, University of Arizona, USA, 1988.

11. Gary, N. E., Witherell, P. C. and Marston, J., Foraging range and distribution of honey bees used for carrot and onion pollination. *Environ. Entomol.*, 1972, **1**, 71–78.
12. Gary, N. E., Witherell, P. C., Lorenzen, K. and Marston, J. M., The interfield distribution of honey bees foraging on carrots, onions, and safflower. *Environ. Entomol.*, 1977, **6**, 637–640.
13. Ockendon, D. J. and Gates, P. J., Variation in pollen viability in the onion (*Allium cepa* L.). *Euphytica*, 1976, **25**, 753–759.
14. Sajjad, A., Saeed, S. and Masood, A., Pollinator community of onion (*Allium cepa* L.) and its role in crop reproductive success. *Pak. J. Zool.*, 2008, **40**, 451–456.
15. Biradar, R., Das, U., Kariyanna, B. and Srivatsava, P., Diversity and abundance of insect visitors/pollinators on onion (*Allium cepa* L.). *Bull. Environ. Pharmacol. Life Sci.*, 2017, **6**(2), 201–204.
16. Tchindebe, G., Farda, D., Dounia, Douka, C., Clautin, N., Auguste, P. M. and Nestor, T. F. F., Diversity of insect pollinators of *Allium cepa* L. (Liliaceae) and assessment of its impact on yields at Gazawa (Cameroon). *J. Appl. Biol. Biotechnol.*, 2021, **9**(2), 85–92.
17. Howlett, B. G., Donovan, B. J., McCallum, J. A., Newstrom, L. E. and Teulon, D. A. J., Between and within field variability of New Zealand indigenous flower visitors to onions. *N. Z. Plant Prot.*, 2005, **58**, 213–218.
18. Mazeed, A. R. and Marey, R. A., Pollinators activity on onion flowers and its effect on seeds yield at Sohag governorate, Egypt. *Egypt. J. Agric. Res.*, 2018, **96**(2), 465–474.
19. Korichi, Y., Malika, A., Karima, K. and Hassina, I., Foraging bees of the onion (*Allium cepa* L.) and their impact on seed production in Tizi-Ouzou area (Algeria). *J. Apicult. Res.*, 2021; doi:10.1080/00218839.2021.1897277.
20. Kevan, P. G. and Baker, H. G., Insects as flower visitors and pollinators. *Annu. Rev. Entomol.*, 1983, **28**, 407–453.
21. Tchindebe, G. and Fohouo, G. T., Foraging and pollination activity of *Apis mellifera adansonii* Latreille (Hymenoptera: Apidae) on flowers of *Allium cepa* L. (Liliaceae) at Maroua, Cameroon, 2014.
22. Al-Sahaf, F. H., Effect of planting method and rose water spray on seed production in onion (*Allium cepa* L.). *Emirates J. Food Agric.*, 2002, **14**, 14–23.
23. Devi, S., Gulati, R., Tehri, K. and Asha, Diversity and abundance of insect pollinators on *Allium cepa* L. *J. Entomol. Zool. Stud.*, 2014, **2**(6), 34–38.
24. Chandel, R. S., Thakur, R. K., Bhardwaj, N. R. and Pathania, N., Onion seed crop pollination: a missing dimension in mountain horticulture. *Acta Hort.*, 2004, **631**, 79–86.
25. Rao, G. M. and Suryanarayana, M. C., Effect of honeybee pollination on seed yield in onion (*Allium cepa* L.). *Indian Bee J.*, 1989, **51**, 9–11.
26. Lorenzon, M. C. A., Rodrigues, A. G. and Desouza, J. R. G. C., Comportamento polinizador de *Trigona spinipes* (Hymenoptera, Apidae) na florada da cebola (*Allium cepa* L.) h'ibrida. *Pesqui. Agropecu. Bras.*, 1993, **28**, 217–221.
27. Tolon, B. and Duman, Y., The effect of pollination by honeybee (*Apis mellifera*) on onion (*Allium cepa*) seed production and quality. In XXXVIII Apimondia International Apicultural Congress, Ljubljana, Solvenia, 2003, p. 554.
28. Kamal, M. M. and Akand, M. M., Foraging activity of different pollinators for seed setting and maximizing seed yield of onion. *IOSR J. Agric. Vet. Sci.*, 2017, **10**(5), 88–96.
29. Clement, S. L., Hellier, B. C., Elbersen, L. R., Staska, R. T. and Evans, M. A., Flies (Diptera: Muscidae: Calliphoridae) are efficient pollinators of *Allium ampeloprasum* L. (Alliaceae) in field cages. *J. Econ. Entomol.*, 2007, **100**, 131–135.
30. Currah, L. and Ockendon, D. J., Onion pollination by blowflies and honey bees in large cages. *Ann. Biol.*, 1983, **103**, 497–506.
31. Saleh, M. *et al.*, Onion flowers anthesis and insect pollinators preferences on onion (*Allium cepa* L.) crop. *Fresenius Environ. Bull.*, 2021, **30**(3), 2580–2585.
32. Kalmath, B. S. and Sattigi, H. N., Pollinator fauna and foraging behaviour of honey bees in onion ecosystem *Chang. Trends Pollen Spore Res.*, 2005, **22**, 25–28.
33. Sanduleac, E., The pollination of vegetable seed plants. *Apiculture*, 1961, **14**, 25–26.
34. Saeed, S., Sajjad, A. and Kown, J. Y., Fidelity of hymenoptera and diptera pollinators in onion (*Allium cepa* L.) pollination. *Entomol. Res.*, 2008, **38**(4), 276–280.
35. Karuppaiah, V., Soumia, P. S. and Wagh, P. D., Diversity and foraging behaviour of insect pollinators in onion. *Indian J. Entomol.*, 2018, **80**(4), 1366–1369.
36. Hosamani, V., Reddy, M. S., Venkateshalu, Hanumanthaswamy, B. C., Lingamurthi, K. R., Ravikumar, B. and Ashoka, N., Pollinator diversity, abundance and their stay time in onion, *Allium cepa* L. *J. Entomol. Zool. Stud.*, 2019, **7**, 158–161.
37. Priti, Abundance and pollination efficiency of insect visitors of onion bloom. *Indian Bee J.*, 1998, **60**, 75–78.
38. Benedek, P., Behaviour of honeybees (*Apis mellifera* L.) in relation to the pollination of onion (*Allium cepa* L.) inflorescences. *Z. Ang. Entomol.*, 1976, **82**, 414–420.
39. Lazic, B., Haker, D. and Markov, R., Contribution to the study of influenced peel on onion fertilization. In Yugoslav Symposium on Seeds of Semenarstvu. Piltvicka Lakes, Croatia, 1985.
40. Ewies, M. A. and El-Sahhar, K., Observation on the behaviour of honeybee on onion and their effect on seed yield. *J. Apic. Res.*, 1977, **16**, 194–196.
41. Moffett, J. O., Pollinating experimental onion varieties. *Am. Bee J.*, 1965, **105**, 378.
42. Bohart, G. E., Nye, W. P. and Hawthorn, I. R., Onion pollination as affected by different levels of pollinator activity. *Utah Agric. Exp. Sta. Bull.*, 1970, **482**, 60.
43. Free, J. B., *Insect Pollination of Crops*, Academic Press, London, UK, 1993, p. 684.
44. Thakur, R. K. and Kumaranag, K. M., Annual Report 2015–2016. ICAR-All India Coordinated Research Project on Honey Bees and Pollinators, Division of Entomology, Indian Agricultural Research Institute, New Delhi, 2016, pp. 15–78.
45. Singh, J. P. and Dharmwal, S. S., The role of honey bees in seed setting on onion at Pant Nagar. Dist. Nainital, Uttar Pradesh, India. *Indian Bee J.*, 1970, **32**, 23–27.
46. Woyke, H. W., Some aspects of the role of the honey bee in onion seed production in Poland. *Acta Hort.*, 1981, **111**, 91–98.
47. Prasad, R., Chand, H. and Singh, R., Effect of honey bee, *Apis mellifera* pollination on the yield and germination of onion, *Allium cepa* seeds. *Shashpa*, 2000, **7**, 53–55.
48. Deodikar, G. B. and Suryanaryana, M. C., Crop yields and bee pollination. *Indian Bee J.*, 1972, **4**, 53–64.
49. Munawar, M. S. and Muzaffar, N., Role of honeybee (*Apis mellifera* L.) pollination on onion (*Allium cepa*) seed production. In 36th Limondia Congress, Vancouver, Canada, 12–18 September 1999, p. 276.
50. Le Baron, F. C., Onion seed sample costs and production. California Agriculture Extension Service, Cost Data Sheet 22, Leaflet, 1962.
51. Yue, M., Luo, S., Liu, J. and Wu, J., *Apis cerana* is less sensitive to most neonicotinoids, despite of their smaller body mass. *J. Econ. Entomol.*, 2018, **111**(1), 39–42.
52. Zaluski, R., Justulin, L. A. and Orsi, R. O., Field-relevant doses of the systemic insecticide fipronil and fungicide pyraclostrobin impair mandibular and hypopharyngeal glands in nurse honeybees (*Apis mellifera*). *Sci. Rep.*, 2017; doi:10.1038/s41598-017-15581-5.
53. Morais, C. R. *et al.*, Eco-toxicological effects of the insecticide fipronil in Brazilian native stingless bees *Melipona scutellaris* (Apidae: Meliponini). *Chemosphere*, 2018, **206**, 632–642; <https://doi.org/10.1016/j.chemosphere.2018.04.153>.
54. Hassani, A. K., Dacher, M., Gauthier, M. and Armengaud, C., Effects of sub-lethal doses of fipronil on the behaviour of the honeybee (*Apis mellifera*). *Pharmacol., Biochem. Behav.*, 2005, **82**, 30–39.

55. Gunasekara, A. S., Truong, T., Goh, K. S., Spurlock, F. and Tjeerdema, R. S., Environmental fate and toxicology of fipronil. *J. Pestic. Sci.*, 2007, **32**, 189–199.
56. Park, M. G., Blitzer, E. J., Gibbs, J., Losey, J. E. and Danforth, B. N., Negative effects of pesticides on wild bee communities can be buffered by landscape context. *Proc. Biol. Sci.*, 2015, **282**, 1–9.
57. Paris, L., Roussel, M., Pereira, B., Delbac, F. and Diogon, M., Disruption of oxidative balance in the gut of the western honeybee *Apis mellifera* exposed to the intracellular parasite *Nosema ceranae* and to the insecticide fipronil. *Microb. Biotechnol.*, 2017, **10**(6), 1702–1717; doi:10.1111/1751-7915.12772.
58. Roat, T. C., dos Santos-Pinto, J. R. A., dos Santos, L. D., Santos, K. S., Malaspina, O. and Palma, M. S., Modification of the brain proteome of Africanized honeybees (*Apis mellifera*) exposed to a sub-lethal doses of the insecticide fipronil. *Ecotoxicology*, 2015; doi:10.1007/s10646-014-1305-8.
59. Lu, C., Hung, Y. and Cheng, Q., A review of sub-lethal neonicotinoid insecticides exposure and effects on pollinators. *Curr. Pollut. Rep.*, 2020; https://doi.org/10.1007/s40726-020-00142-8
60. Decourtye, A., Devillers, J., Genecque, E., Le Menach, K., Budzinski, H. and Cluzeau, S., Comparative sub-lethal toxicity of nine pesticides on olfactory learning performances of the honeybee *Apis mellifera*. *Arch. Environ. Contam. Toxicol.*, 2005, **48**, 242–250.
61. Siviter, H., Horner, J., Brown, M. J. F. and Leadbeater, E., Sulfoxaflo exposure reduces egg laying in bumblebees *Bombus terrestris*. *J. Appl. Ecol.*, 2020, **57**, 160–169; https://doi.org/10.1111/1365-2664.13519.
62. Ladurner, E., Bosch, J., Kemp, W. P. and Maini, S., Assessing delayed and acute toxicity of five formulated fungicides to *Osmia lignaria* Say and *Apis mellifera*. *Apidologie*, 2005, **36**, 449–460.
63. Atkins, E. L. and Kellum, D., Comparative morphogenic and toxicity studies on the effect of pesticides on honeybee brood. *J. Apic. Res.*, 1986, **25**, 242–255.
64. Mussen, E. C., Lopez, J. E. and Peng, C. Y., Effects of selected fungicides on growth and development of larval honey bees, *Apis mellifera* L. (Hymenoptera: Apidae). *Environ. Entomol.*, 2004, **33**(5), 1151–1154.
65. Heard, T. A., The role of stingless bees in crop pollination. *Annu. Rev. Entomol.*, 1999, **44**, 183–206.
66. Herrmann, J. D., Beye, H., de la Broise, C., Hartlep, H. and Diekötter, T., Positive effects of the pollinators *Osmia cornuta* (Megachilidae) and *Lucilia sericata* (Calliphoridae) on strawberry quality. *Arthropod – Plant Interact.*, 2019, **13**, 71–77.
67. Dag, A. and Gazit, S., Mango pollinators in Israel. *J. Appl. Hortic.*, 2000, **2**, 39–43.
68. Rader, R. *et al.*, Alternative pollinator taxa are equally efficient but not as effective as the honeybee in a mass flowering crop. *J. Appl. Ecol.*, 2009, **46**, 1080–1087.
69. Nye, W. P., Shasha'a, N. S., Campbell, W. F. and Hamson, A. R., Insect pollination and seed set of onions (*Allium cepa* L.). *Utah State Univ. Agric. Exp. Sta. Res. Rep.*, 1973, **6**, 1–15.
70. Derecka, K., Blythe, M. J., Malla, S., Genereux, D. P., Guffanti, A. and Pavan, P., Transient exposure to low levels of insecticide affects metabolic networks of honeybee larvae. *PLoS ONE*, 2013, **8**(7), e68191; https://doi.org/10.1371/journal.pone.0068191
71. Decourtye, A., Devillers, J., Cluzeau, S., Charreton, M. and Pham-Delègue, M. H., Effects of imidacloprid and deltamethrin on associative learning in honeybees under semi-field and laboratory conditions. *Ecotoxicol. Environ. Saf.*, 2004, **57**, 410–419.
72. Decourtye, A., Armengaud, C., Renou, M., Devillers, J., Cluzeau, S. and Gauthier, M., Imidacloprid impairs memory and brain metabolism in the honeybee (*Apis mellifera* L.). *Pestic. Biochem. Physiol.*, 2004, **78**, 83–92.
73. Williamson, S. M. and Wright, G. A., Exposure to multiple cholinergic pesticides impairs olfactory learning and memory in honeybees. *J. Exp. Biol.*, 2013, **216**(10), 1799–1807.
74. Yang, E. C., Chuang, Y. C., Chen, Y. L. and Chang, L. H., Abnormal foraging behaviour induced by sub-lethal dosage of imidacloprid in the honey bee (Hymenoptera: Apidae). *J. Econ. Entomol.*, 2008, **101**(6), 1743–1748.
75. Teeters, B. S., Johnson, R. M., Ellis, M. D. and Siegfried, B. D., Using video-tracking to assess sub-lethal effects of pesticides on honey bees (*Apis mellifera* L.). *Environ. Toxicol. Chem.*, 2012, **31**(6), 1349–1354.
76. Boily, M., Sarrasin, B., Deblois, C., Aras, P. and Chagnon, M., Acetylcholinesterase in honey bees (*Apis mellifera*) exposed to neonicotinoids, atrazine and glyphosate: laboratory and field experiments. *Environ. Sci. Pollut. Res. Int.*, 2013, **20**(8), 5603.
77. Alaux, C., Brunet, J. L., Dussaubat, C., Mondet, F., Tchamitchan, S. and Cousin, M., Interactions between *Nosema* microspores and a neonicotinoid weaken honeybees (*Apis mellifera*). *Environ. Microbiol.*, 2010, **12**(3), 774–782.
78. Lu, C., Warchol, K. M. and Callahan, R. A., Sub-lethal exposure to neonicotinoids impaired honey bees winterization before proceeding to colony collapse disorder. *Bull. Insectol.*, 2014, **67**(1), 125–130.
79. Cresswell, J. E., Robert, F. X., Florance, H. and Smirnov, N., Clearance of ingested neonicotinoid pesticide (imidacloprid) in honey bees (*Apis mellifera*) and bumblebees (*Bombus terrestris*). *Pest Manage. Sci.*, 2014, **70**(2), 332–337.
80. Cresswell, J. E., Page, C. J., Uygun, M. B., Holmbergh, M., Li, Y. and Wheeler, J. G., Differential sensitivity of honey bees and bumble bees to a dietary insecticide (imidacloprid). *Zoology (Jena)*, 2012, **115**(6), 365–371; https://doi.org/10.1016/j.zool.2012.05.003.
81. Scholer, J. and Krischik, V., Chronic exposure of imidacloprid and clothianidin reduce queen survival, foraging, and nectar storing in colonies of *Bombus impatiens*. *PLoS ONE*, 2014, **9**(3), e91573.
82. Whitehorn, P. R., O'Connor, S., Goulson, D. and Wackers, F. L., Neonicotinoid pesticide reduces bumble bee colony growth and queen production. *Science*, 2012, **336**(6079), 351–352; https://doi.org/10.1126/science.1215025.
83. Bryden, J., Gill, R. J., Mitton, R. A., Raine, N. E. and Jansen, V. A., Chronic sub-lethal stress causes bee colony failure. *Ecol. Lett.*, 2013, **16**, 1463–1469.
84. Feltham, H., Park, K. and Goulson, D., Field realistic doses of pesticide imidacloprid reduce bumblebee pollen foraging efficiency. *Ecotoxicology*, 2014, **23**, 317–323; https://doi.org/10.1007/s10646-014-1189-7.
85. Abbott, V. A., Nadeau, J. L., Higo, H. A. and Winston, M. L., Lethal and sub-lethal effects of imidacloprid on *Osmia lignaria* and clothianidin on *Megachile rotundata* (Hymenoptera: Megachilidae). *J. Econ. Entomol.*, 2008, **101**(3), 784–796.
86. Palmer, M. J., Moffat, C., Saranzewa, N., Harvey, J., Wright, G. A. and Connolly, C. N., Cholinergic pesticides cause mushroom body neuronal inactivation in honeybees. *Nature Commun.*, 2013, **4**, 1634; https://doi.org/10.1038/ncomms2648.
87. Tomé, H. V., Martins, G. F., Lima, M. A., Campos, L. A. and Guedes, R. N., Imidacloprid-induced impairment of mushroom bodies and behaviour of the native stingless bee *Melipona quadrifasciata anthidioides*. *PLoS ONE*, 2012, **7**(6), e38406.
88. Rossi Cde, A., Roat, T. C., Tavares, D. A., Cintra-Socolowski, P. and Malaspina, O., Effects of sub-lethal doses of imidacloprid in malpighian tubules of Africanized *Apis mellifera* (Hymenoptera, Apidae). *Microsc. Res. Tech.*, 2013, **76**(5), 552–558; https://doi.org/10.1002/jemt.22199.
89. Doublet, V., Labarussias, M., de Miranda, J. R., Moritz, R. F. A. and Paxton, R. J., Bees under stress: sublethal doses of a neonicotinoid pesticide and pathogens interact to elevate honey bee mortality across the life cycle. *Environ. Microbiol.*, 2014; https://doi.org/10.1111/1462-2920.12426.
90. Williamson, S. M., Willis, S. J. and Wright, G. A., Exposure to neonicotinoids influences the motor function of adult worker honeybees. *Ecotoxicology*, 2014, **23**(8), 1409–1418.

91. Vidau, C., Diogon, M., Aufauvre, J., Fontbonne, R., Viguès, B. and Brunet, J. L., Exposure to sub-lethal doses of fipronil and thiacloprid highly increases mortality of honeybees previously infected by *Nosema ceranae*. *PLoS ONE*, 2011, **6**(6), e21550; <https://doi.org/10.1371/journal.pone.0021550>.
92. Hassani, A. K., Dacher, M., Gary, V., Lambin, M., Gauthier, M. and Armengaud, C., Effects of sub-lethal doses of acetamiprid and thiamethoxam on the behaviour of the honeybee (*Apis mellifera*). *Arch. Environ. Contam. Toxicol.*, 2008, **54**, 653–661.
93. Henry, M., Rollin, O., Aptel, J., Tchamitchian, S., Beguin, M. and Requier, F., A common pesticide decreases foraging success and survival in honey bees. *Science*, 2012, **336**(6079), 348–350; <https://doi.org/10.1126/science.1215039>.
94. Sandrock, C., Tanadini, L. G., Pettis, J. S., Biesmeijer, J. C., Potts, S. G. and Neumann, P., Sublethal neonicotinoid insecticide exposure reduces solitary bee reproductive success. *Agric. For. Entomol.*, 2014, **16**, 119–128.
95. Fischer, J., Müller, T., Spatz, A. K., Greggers, U., Grünewald, B. and Menze, R., Neonicotinoids interfere with specific components of navigation in honeybees. *PLoS ONE*, 2014, **9**(3), e91364; <https://doi.org/10.1371/journal.pone.0091364>.
96. Larson, J. L., Redmond, C. T. and Potter, D. A., Assessing insecticide hazard to bumble bees foraging on flowering weeds in treated lawns. *PLoS ONE*, 2013, **8**(6), e66375.

Received 8 June 2022; revised accepted 3 October 2022

doi: 10.18520/cs/v124/i3/304-312
