

Endophytes from marine macroalgae: promising sources of novel natural products

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Endophytic fungi are emerging as an excellent source of bioactive natural products. Though much of the research on endophytes and the natural products that they produce has focused on those isolated from terrestrial plants, endophytes of marine macroalgae have recently gained attention as an untapped source of biodiversity with the potential to yield novel bioactive metabolites. Recent work on the endophytic fungal assemblages of macroalgae has highlighted the scale of biodiversity and chemical diversity associated with fungal endophytes of macroalgae. The array of fungal species isolated and the discovery of new natural products exhibiting antimicrobial, anticancer and antiviral activities give a glimpse of the potential of macroalgal endophytes. The aim of this review is to highlight recent findings relating to endophytes of marine macroalgae with particular focus on the biodiversity of the endophytes associated with each macroalgal host, the geographical location of host alga and the biological activities exhibited by the natural products of these endophytic fungi.

Keywords: Bioactive metabolites, endophytic fungi, marine macroalgae, natural products.

Introduction

ENDOPHYTIC fungi are universally found in the plant kingdom¹⁻³. The number of endophytes that can colonize a single plant ranges from just a few to hundreds¹ with each association assumed to be unique but characterized as somewhere between symbiotic and pathogenic⁴.

While the relationship between plant host and endophyte is not always understood, many endophytes enhance host plant fitness through the production of bioactive compounds, improving survival against environmental stresses as well as promoting plant growth¹. These functional metabolites are the target of current research in drug discovery⁴⁻⁶.

With the knowledge that many terrestrial plants are associated with endophytes able to produce bioactive compounds, endophytes from marine plants and macroalgae are gaining special interest because of their existence in

an ecosystem distinguished by resource limitations such as temperature, light availability, salinity and osmotic stress^{4,7}. Indeed, endophytic fungi from various macroalgal hosts are increasingly being identified as sources of bioactive compounds with medicinal relevance (e.g. by exhibiting anticancer, antimicrobial and antifungal activities to name a few).

Despite this, our knowledge of the fundamental aspects of macroalgal endophyte biology is remarkably sparse and majority of the studies that have been performed on macroalgal endophytes provide little information that can meaningfully increase our understanding. This review will highlight the current state of knowledge relating to fungal endophytes of marine macroalgae with a particular focus on the geography and taxonomy of the algal hosts, the diversity and taxonomy of the fungal endophytes, and the reported bioactivity of extracts and natural products obtained from the fungi. Our aim is twofold: first to provide a comprehensive resource for researchers in the field and secondly, to highlight the deficiencies in the field to be addressed in future studies.

Geographic location of macroalgae

When evaluating the global distribution of algal endophytes, there exist discrepancies in the literature. While some studies are survey-based where researchers have attempted to maximize the number of hosts collected and diversity of endophytes⁸⁻¹², others are focused on the endophytes of a single macroalgal host or a single endophyte for the purpose of chemical elucidation and bioactivity assays¹³⁻²¹. Few studies have been performed screening marine macroalgae for their endophytic fungi. Most of the information on endophytic fungi in marine macroalgae comes from a study conducted by Suryanarayanan *et al.*⁹ in India. As a result of this work, studies concerning the diversity of fungal endophytes and their potential for bioactive natural products have also been performed in Canada, Malaysia and the United Kingdom (UK)¹⁰⁻¹².

Nearly 100 marine macroalgae have been investigated for their endophytic fungi (Table 1), though approximately 75% of them have been investigated from just six locations (Baltic Sea, Canada, China, India, North Sea and

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Table 1. Summary of associated fungal endophytes classified by location

Location	Algal host	Fungal endophyte	Reference
Northern hemisphere			
Azores	<i>Valonia utricularis</i> (g)	<i>Chaetomium</i> sp.	32
Baltic Sea	<i>Enteromorpha</i> sp. (g)	<i>Coniothyrium cereale</i>	20, 21
		<i>Wardomyces anomalus</i>	14
	<i>Fucus</i> sp. (b)	<i>Dendryphiella arenaria</i>	23
		<i>Phoma</i> sp.	
	<i>Polyides rotundus</i> (r)	Unidentified	
	<i>Polysiphonia</i> sp. (r)	<i>Colletotrichum</i> sp.	
		<i>Geniculosporium</i> sp.	61
		<i>Geniculosporium</i> sp.	23
	<i>Ulva linza</i> (g)	<i>Alternaria</i> sp.	
Canada (Atlantic coast)	<i>Ascophyllum nodosum</i> (b)	<i>Aspergillus</i> sp. II	
		<i>Mycosphaerella ascophylli</i>	37, 38, 52, 53, 54, 55, 56, 87
	<i>Chondrus crispus</i> (r)	<i>Aspergillus</i> sp. I	11
		<i>Penicillium crustosum</i> I	
	<i>Devaleraea ramentacea</i> (r)	<i>Aureobasidium pullulans</i> I	
		<i>Botrytis</i> sp. I	
		<i>Botrytis</i> sp. II	
		<i>Cladosporium</i> sp.	
		<i>Coniothyrium</i> sp.	
		<i>Penicillium decumbens</i> I	
		<i>Trametes versicolor</i>	
		Septate pigmented I	
		Septate pigmented II	
		Septate pigmented III	
		Sterile hyaline I	
		Sterile hyaline II	
		White fluffy II	
	<i>Fucus spiralis</i> (b)	Black hyaline I	
		Pigmented hyaline I	
		Sterile hyaline XV	
		Septate pigmented V	
	<i>Fucus vesiculosus</i> (b)	Sterile hyaline XXI	
	<i>Mastocarpus stellatus</i> (r)	Coelomycete I	
		<i>Penicillium decumbens</i> II	
		<i>Penicillium</i> sp.	
		Red yeast I	
		Red yeast II	
		Sterile hyaline III	
		Sterile hyaline IV	
		Septate pigmented IV	
	<i>Palmaria palmata</i> (r)	<i>Botrytis</i> sp. III	
		<i>Helicomyces</i> sp.	
		<i>Hypoxylon</i> sp.	
		<i>Penicillium chrysogenum</i> I	
		<i>Penicillium crustosum</i> II	
		<i>Penicillium decumbens</i> III	
		Sterile hyaline V	
		Sterile hyaline VI	
		Sterile hyaline VII	
	<i>Polysiphonia lanosa</i> (r)	<i>Aureobasidium pullulans</i> II	
		<i>Botryotinia fuckeliana</i>	
		Sterile hyaline VIII	
		Sterile hyaline IX	
		Sterile hyaline X	
	<i>Porphyra purpurea</i> (r)	Sterile hyaline XI	
		Sterile hyaline XII	
	<i>Porphyra umbilicalis</i> (r)	<i>Aspergillus sydowii</i>	
		Sterile hyaline XIII	
		Sterile hyaline XIV	
	<i>Saccharina latissima</i> (b)	<i>Aspergillus</i> sp. III	

(Contd)

Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference
		<i>Penicillium chrysogenum</i> II	
		<i>Penicillium chrysogenum</i> III	
		<i>Penicillium soppii</i> I	
		Black hyaline II	
		Pigmented hyaline IV(Red)	
		Sterile beige I	
		Sterile beige II	
		Sterile hyaline XVI	
		White hyaline I	
		White hyaline II	
		White hyaline IV	
	<i>Spongomorpha arcta</i> (g)	<i>Penicillium soppii</i> II	
		<i>Penicillium spinulosum</i>	
		<i>Penicillium</i> sp.	
		Black hyaline III	
		Pigmented hyaline V	
		Pigmented hyaline VI	
		Pigmented hyaline VII	
		Sterile beige III	
		Sterile hyaline XVII	
		Sterile hyaline XVIII	
		White hyaline III	
	<i>Ulva lactuca</i> (g)	<i>Penicillium chrysogenum</i> IV	
		Septate pigmented VII	
		Sterile hyaline XX	
	<i>Ulva intestinalis</i> (g)	<i>Aspergillus</i> sp. V	
		Pigmented hyaline VIII	
		Septate pigmented VI	
		Septate pigmented VIII	
		Sterile hyaline XIX	
China	<i>Chondrus ocellatus</i> (r)	<i>Penicillium echinulatum</i>	64
	<i>Codium fragile</i> (g)	<i>Aspergillus versicolor</i>	66
		<i>Gibberella zeae</i>	65
		<i>Trichoderma longibrachiatum</i>	69
	<i>Colpomenia sinuosa</i> (b)	<i>Aspergillus niger</i>	16, 17, 18, 19, 89
	<i>Corallina officinalis</i> (r)	<i>Aspergillus flavus</i>	31
	<i>Enteromorpha prolifera</i> (g)	Unidentified fungus 9	8
		Unidentified fungus 10	
	<i>Gelidium amansii</i> (r)	Unidentified fungus 2	
	<i>Gracilaria lemaneiformis</i> (r)	Unidentified fungus 3	
		Unidentified fungus 4	
	<i>Gracilariopsis</i> sp. (r)	Unidentified	71
	<i>Gymnogongrus flabelliformis</i> (r)	<i>Aspergillus wentii</i>	81
	<i>Heterosiphonia japonica</i> (r)	<i>Aspergillus oryzae</i>	29, 30
	<i>Laurencia similis</i> (r)	<i>Exophiala oligosperma</i>	62
	<i>Laurencia</i> sp. (r)	<i>Alternaria alternata</i>	48
	<i>Laurencia</i> sp. (r)	<i>Penicillium chrysogenum</i>	49, 50, 51
	<i>Polysiphonia urceolata</i> (r)	<i>Chaetomium globosum</i>	86
	<i>Rhodomela confervoides</i> (r)	Unidentified fungus 1	8
	<i>Sargassum fusiforme</i> (b)	<i>Aspergillus wentii</i>	70
	<i>Sargassum horneri</i> (b)	<i>Pestalotiopsis</i> sp.	26
	<i>Sargassum kjellmanianum</i> (b)	<i>Aspergillus ochraceus</i>	28
	<i>Sargassum palladium</i> (b)	<i>Penicillium chrysogenum</i>	34
	<i>Sargassum thunbergii</i> (b)	<i>Aspergillus versicolor</i>	68
		<i>Eurotium cristatum</i>	39
		Unidentified fungus 5	8
		Unidentified fungus 6	
		Unidentified fungus 7	
		<i>Penicillium glabrum</i>	
	<i>Sargassum</i> sp. (b)	<i>Aspergillus wentii</i>	63
		<i>Aspergillus wentii</i>	80, 82

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Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference	
Egypt France India	<i>Ulva pertusa</i> (g)	<i>Aspergillus wentii</i>	90	
		Unidentified fungus	88	
		<i>Chaetomium globosum</i>	27, 35	
		Unidentified fungus 11	8	
	<i>Undaria pinnatifida</i> (b)	<i>Ulva</i> sp. (g)	Unidentified fungus 12	
			<i>Guignardia</i> sp.	85
	France	<i>Ulva</i> sp. (g)	<i>Penicillium</i> sp.	47
			<i>Dendryphiella salina</i>	
	India	<i>Acanthophora spicifera</i> (r)	<i>Aspergillus sydowii</i>	84
			<i>Caulerpa racemosa</i> (g)	9
			<i>Aspergillus niger</i>	
			<i>Aspergillus terreus</i>	
			<i>Aspergillus</i> sp. 2	
			<i>Aspergillus</i> sp. 4	
			<i>Aureobasidium pullulans</i>	
			<i>Chaetomium</i> sp. 1	
			<i>Cladosporium</i> sp. 1	
			<i>Fusarium</i> sp. 2	
			<i>Fusarium</i> sp. 4	
			<i>Fusarium</i> sp. 5	
			<i>Mucor</i> sp.	
			<i>Penicillium</i> sp. 1	
			<i>Phoma</i> sp.	
			<i>Pyrenocheta</i> sp.	
			Sterile form 1	
			Yeast sp. 1	
		<i>Caulerpa scalpelliformis</i> (g)	<i>Aspergillus niger</i>	
	<i>Aspergillus terreus</i>			
	<i>Aspergillus</i> sp. 2			
	<i>Curvularia</i> sp. 1			
	<i>Caulerpa sertularioides</i> (g)	<i>Paecilomyces</i> sp. 1		
		<i>Penicillium</i> sp. 1		
		<i>Aspergillus niger</i>		
		<i>Aspergillus terreus</i>		
		<i>Aspergillus</i> sp. 2		
		<i>Chaetomium</i> sp. 1		
		<i>Cladosporium</i> sp. 1		
		<i>Fusarium</i> sp. 2		
		<i>Myrothecium</i> sp.		
		<i>Penicillium</i> sp. 1		
	<i>Dictyota dichotoma</i> (b)	<i>Phialophora</i> sp.		
		Sterile form 1		
		Yeast sp. 1		
		<i>Aspergillus niger</i>		
	<i>Gelidiella acerosa</i> (r)	<i>Aspergillus terreus</i>		
		<i>Trichoderma</i> sp.		
		Sterile form 1		
		<i>Alternaria</i> sp.		
	<i>Gelidiella acerosa</i> (r)	<i>Aspergillus niger</i>		
		<i>Aspergillus terreus</i>		
		<i>Aspergillus versicolor</i>		
		<i>Nigrospora</i> sp. 1		
	<i>Gelidiella acerosa</i> (r)	<i>Penicillium</i> sp. 1		
		<i>Phoma</i> sp.		
	<i>Gracilaria crassa</i> (r)	Sterile form 1		
		<i>Trichoderma</i> sp.		
	<i>Gracilaria edulis</i> (r)	Sterile form 1		
		<i>Aspergillus terreus</i>		
	<i>Gracilaria</i> sp. (r)	<i>Nigrospora</i> sp. 1		
		<i>Paecilomyces</i> sp. 1		
		<i>Aphanocladium</i> sp.		
		<i>Aspergillus niger</i>		
		<i>Aspergillus terreus</i>		

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Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference
		<i>Monilia</i> sp.	
		<i>Paecilomyces</i> sp. 2	
		Yeast sp. 2	
	<i>Grateloupia lithophila</i> (r)	<i>Aspergillus niger</i>	
		<i>Aspergillus terreus</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Chaetomium</i> sp. 1	
		<i>Cladosporium</i> sp. 1	
		<i>Emericella nidulans</i>	
		<i>Nigrospora</i> sp. 1	
		Sterile form 1	
		Yeast sp. 1	
	<i>Halimeda macroloba</i> (g)	<i>Aspergillus niger</i>	
		<i>Aspergillus terreus</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Aspergillus</i> sp. 5	
		<i>Paecilomyces</i> sp. 1	
		<i>Penicillium</i> sp. 1	
		<i>Trichoderma</i> sp.	
		Yeast sp. 1	
	<i>Halymenia</i> sp. 1 (r)	<i>Aspergillus niger</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Aspergillus terreus</i>	
		<i>Chaetomium</i> sp. 1	
		<i>Cladosporium</i> sp. 1	
		Coelomycete form 2	
		<i>Emericella nidulans</i>	
		<i>Paecilomyces</i> sp. 1	
		<i>Phoma</i> sp.	
	<i>Halymenia</i> sp. 2 (r)	<i>Aspergillus niger</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Aspergillus terreus</i>	
		<i>Cladosporium</i> sp. 1	
		<i>Drechslera</i> sp. 1	
		<i>Emericella nidulans</i>	
		<i>Penicillium</i> sp. 1	
		Yeast sp. 1	
	<i>Lobophora variegata</i> var. <i>indica</i> (b)	<i>Aspergillus niger</i>	
		<i>Aspergillus terreus</i>	
		<i>Chaetomium</i> sp. 1	
		<i>Emericella nidulans</i>	
		<i>Nigrospora</i> sp. 1	
		Sterile form 3	
		UNI 1	
	<i>Padina gymnospora</i> (b)	<i>Aspergillus terreus</i>	
		<i>Chaetomium</i> sp. 1	
		<i>Cladosporium</i> sp. 1	
		<i>Cladosporium</i> sp. 2	
		<i>Penicillium</i> sp. 1	
		<i>Phoma</i> sp.	
		<i>Trichoderma</i> sp.	
		Sterile form 1	
		Sterile form 2	
	<i>Padina tetrastromatica</i> (b)	<i>Acremoniella</i> sp.	
		<i>Ascotricha</i> sp.	
		<i>Aspergillus niger</i>	
		<i>Aspergillus terreus</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Aspergillus</i> sp. 4	
		<i>Aspergillus</i> sp. 5	
		<i>Chaetomium</i> sp. 1	
		<i>Monodictys</i> sp.	

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Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference
		<i>Paecilomyces</i> sp. 1	
		<i>Penicillium</i> sp. 1	
		<i>Phialophora</i> sp.	
		Sterile form 2	
	<i>Portieria hornemanii</i> (r)	<i>Aspergillus terreus</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Cladosporium</i> sp. 1	
		<i>Emericella nidulans</i>	
		<i>Memnoniella</i> sp.	
		<i>Penicillium</i> sp. 1	
		<i>Penicillium</i> sp. 2	
		<i>Phomopsis</i> sp.	
		<i>Trichophyton</i> like	
		<i>Trimmatostroma</i> sp.	
		Sterile form 1	
		Yeast sp. 2	
		Yeast sp. 4	
	<i>Sargassum ilicifolium</i> (b)	<i>Aspergillus terreus</i>	
		Sterile form 1	
	<i>Sargassum</i> sp. (b)	<i>Alternaria</i> sp. 5	
		<i>Aspergillus niger</i>	
		<i>Aspergillus terreus</i>	
		<i>Colletotrichum</i> sp. 1	
		<i>Curvularia lunata</i>	
		<i>Drechslera</i> sp. 1	
		<i>Helicosporium</i> sp.	
		<i>Nigrospora</i> sp. 1	
		<i>Oidiodendron</i> sp.	
		<i>Penicillium</i> sp. 1	
		<i>Taeniolella</i> sp.	
		<i>Varicosporium</i> sp.	
		Sterile form 1	
		Sterile form 2	
		Sterile form 4	
		Sterile form 12	
		Yeast sp. 1	
	<i>Sargassum wightii</i> (b)	<i>Aspergillus niger</i>	
		<i>Aspergillus terreus</i>	
		<i>Aspergillus</i> sp. 2	
		<i>Aspergillus</i> sp. 10	
		<i>Aspergillus</i> sp. 11	
		<i>Cladosporium</i> sp. 1	
		Coelomycete form 2	
		<i>Nigrospora</i> sp. 1	
		<i>Paecilomyces</i> sp. 1	
		<i>Penicillium</i> sp. 1	
		Sterile form 1	
		Sterile form 2	
		Yeast sp. 1	
		Yeast sp. 3	
	<i>Stoechospermum marginatum</i> (b)	<i>Alternaria</i> sp. 1	
		<i>Aspergillus terreus</i>	
		<i>Chaetomium</i> sp. 1	
		<i>Emericella nidulans</i>	
		<i>Nigrospora</i> sp. 1	
		Sterile form 3	
	<i>Turbinaria decurrens</i> (b)	<i>Alternaria</i> sp. 1	
		<i>Aspergillus terreus</i>	
		<i>Chaetomium</i> sp. 1	
		<i>Chaetomium</i> sp. 2	
		<i>Chaetomium</i> sp. 3	
		<i>Curvularia</i> sp. 1	

(Contd)

Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference
	<i>Turbinaria</i> sp. (b)	<i>Drechslera</i> sp. 1 <i>Emericella nidulans</i> <i>Nigrospora</i> sp. 1 <i>Phialophora</i> sp. <i>Pseudogymnoascus</i> -like Sterile form 1 UNI 2 <i>Alternaria</i> sp. 3 <i>Alternaria</i> sp. 4 <i>Alternaria</i> sp. 5 <i>Aspergillus terreus</i> <i>Aspergillus</i> sp. 2 <i>Aspergillus</i> sp. 4 <i>Aspergillus</i> sp. 12 <i>Chaetomium</i> sp. 3 <i>Cladosporium</i> sp. 1 <i>Cladosporium</i> sp. 3 <i>Colletotrichum</i> sp. 1 <i>Curvularia lunata</i> <i>Curvularia tuberculata</i> <i>Curvularia</i> sp. 3 <i>Drechslera papendorfii</i> <i>Drechslera</i> sp. 1 <i>Emericella nidulans</i> <i>Fusarium</i> sp. 5 <i>Monodictys</i> sp. <i>Paecilomyces</i> sp. 1 <i>Phaeotrichoconis</i> sp. Sterile form 1 UNI 3 <i>Varicosporium</i> sp. Yeast sp. 1	
	<i>Turbinaria conoides</i> (b)	<i>Aspergillus terreus</i> <i>Aspergillus</i> sp. 5 <i>Chaetomium</i> sp. 1 <i>Chaetomium</i> sp. 2 <i>Cladosporium</i> sp. 1 <i>Emericella nidulans</i> <i>Phoma</i> sp. Sterile form 2 Sterile form 3	
	<i>Ulva fasciata</i> (g)	<i>Aspergillus niger</i> <i>Aspergillus terreus</i> <i>Aspergillus</i> sp. 2 <i>Chaetomium</i> sp. 1 <i>Curvularia lunata</i> <i>Curvularia</i> sp. 3 <i>Paecilomyces</i> sp. 1 Sterile form 3 Yeast sp. 1 Yeast sp. 6	
	<i>Ulva lactuca</i> (g)	<i>Aspergillus niger</i> <i>Aspergillus terreus</i> <i>Aspergillus</i> sp. 2 <i>Chaetomium</i> sp. 1 <i>Cladosporium</i> sp. 1 <i>Nigrospora</i> sp. 1 <i>Penicillium</i> sp. 1	
Malaysia	<i>Caulerpa lentillifera</i> (g)	Unidentified fungus 3 Unidentified fungus 4 Unidentified fungus 5	10

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Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference	
Mediterranean Sea	<i>Caulerpa racemosa</i> (g)	Unidentified fungus 6		
		Unidentified fungus 7		
		Unidentified fungus 8		
	<i>Caulerpa racemosa</i> variant (g)	Unidentified fungus 11		
		Unidentified fungus 9		
	<i>Padina australis</i> (b)	Unidentified fungus 10		
		Unidentified fungus 12		
	<i>Sargassum oligocystum</i> (b)	Unidentified fungus 13		
		Unidentified fungus 14		
		Unidentified fungus 15		
		<i>Turbinaria conoides</i> (b)	Unidentified fungus 1	
			Unidentified fungus 2	
	North Sea	<i>Cladostephus spongius</i> (b)	<i>Acremonium</i> sp.	15
			<i>Nodulisporium</i> sp.	75
		<i>Plocamium</i> sp. (r)	<i>Monodictys putredinis</i>	25
Unidentified green alga		<i>Chaetomium</i> sp.	77	
Unidentified		<i>Emericella nidulans</i> var. <i>acristata</i>	60	
North Sea	<i>Ceramium</i> sp. (r)	<i>Nodulisporium</i> sp.	76	
		<i>Phaeosphaeria spartinae</i>	41, 42	
	<i>Fucus vesiculosus</i> (b)	<i>Penicillium</i> sp.	23	
		<i>Phoma</i> sp.		
	<i>Fucus</i> sp. (b)	<i>Tolyptocladium inflatum</i>		
		<i>Arthrimum</i> sp.		
	<i>Laminaria digita</i> (b)	<i>Dendryphiella salina</i>		
	<i>Plocamium</i> sp. (r)	<i>Acremonium</i> sp.	75	
	<i>Polysiphonia violacea</i> (r)	<i>Apiospora montagnei</i>	58	
	<i>Ulva</i> sp. (g)	<i>Ascochyta salicorniae</i>	72	
<i>Ascochyta salicorniae</i>		78		
Sri Lanka	<i>Laurencia ceylanica</i> (r)	<i>Aspergillus terreus</i>	57	
United Kingdom (Isle of Wight)	<i>Ascophyllum nodosum</i> (b)	<i>Mycosphaerella ascophylli</i>	79	
		<i>Lautitia danica</i>		
	<i>Dilsea carnosa</i> (r)	<i>Mycaureola dilseae</i>		
United Kingdom (Lulworth Coast)	<i>Fucus vesiculosus</i> (b)	<i>Dendryphiella salina</i>	23	
		<i>Dendryphiella salina</i>		
United Kingdom (Shetland Islands)	<i>Ascophyllum nodosum</i> (b)	<i>Aspergillus fumigatus</i> I	12	
		<i>Cladosporium</i> sp. I		
		<i>Dendryphiella salina</i>		
		<i>Lichtheimia corymbifera</i>		
		<i>Aspergillus fumigatus</i> V		
		<i>Aspergillus fumigatus</i> VI		
		<i>Coniothyrium</i> sp. II		
		<i>Coniothyrium</i> sp. III		
		<i>Penicillium</i> sp. VII		
		<i>Penicillium</i> sp. V	12, 44	
	<i>Fucus serratus</i> (b)	<i>Penicillium</i> sp. VI	12	
		Sterile beige VIII		
		Sterile beige IX		
		Sterile beige X		
		Sterile beige XI		
		Sterile pigmented VI		
		<i>Alternaria</i> sp.		
		<i>Aspergillus fumigatus</i> II		
		<i>Aspergillus fumigatus</i> III		
		<i>Aspergillus fumigatus</i> IV		
<i>Fucus spiralis</i> (b)	<i>Chalara</i> sp.			
	<i>Coniothyrium</i> sp. I			
	<i>Penicillium</i> sp. I			
	Sterile beige I			
	Sterile beige II			

(Contd)

Table 1. (Contd)

Location	Algal host	Fungal endophyte	Reference
	<i>Plocamium cartilagineum</i> (r)	<i>Acremonium</i> sp. <i>Cladosporium</i> sp. II <i>Metschnikowia</i> sp. <i>Penicillium biourgeianum</i> Sterile beige VI Sterile beige VII Sterile pigmented IV Sterile pigmented V	
	<i>Polysiphonia lanosa</i> (r)	<i>Phaeosphaeria</i> sp. Sterile beige XII Sterile beige XIII Sterile beige XIV Sterile beige XV Sterile beige XVI Sterile beige XVII Sterile pigmented VII Sterile pigmented VIII	
	<i>Porphyra</i> sp. (r)	<i>Microdochium</i> sp. <i>Rhodotorula mucilaginosa</i> Sterile beige III Sterile beige IV Sterile beige V	
	<i>Ulva intestinalis</i> (g)	<i>Bionectria ochroleuca</i> <i>Cordyceps</i> sp. <i>Leptosphaeria</i> sp. <i>Penicillium</i> sp. III <i>Penicillium</i> sp. IV <i>Penicillium</i> sp. VIII Pigmented filamentous Septate hyaline I Sterile pigmented II Sterile pigmented III Sterile pigmented IX	
	<i>Ulva lactuca</i> (g)	<i>Eurotium</i> sp. <i>Leotiomyces</i> sp. <i>Penicillium</i> sp. II <i>Pseudeurotium bakeri</i> <i>Stilbella fimetaria</i> Sterile pigmented I	
United States (California)	<i>Cystoseira</i> sp. (b) <i>Halidrys</i> sp. (b)	<i>Haloguignardia irritans</i> <i>Haloguignardia irritans</i>	24
United States (Florida)	<i>Ceramium</i> sp. (r) <i>Digenea simplex</i> (r) <i>Hypnea musciformis</i> (r) <i>Sargassum</i> sp. (b)	<i>Chaetomium</i> sp. <i>Dendryphiella arenaria</i> <i>Dendryphiella arenaria</i> <i>Dendryphiella arenaria</i> <i>Corollospora</i> sp.	23
Southern hemisphere			
Brazil	<i>Bostrychia radicans</i> (r)	<i>Phomopsis longicolla</i>	43
Chile	<i>Prasiola crispa</i> (g)	<i>Mastodia tessellata</i>	74
Indonesia	<i>Gracilaria</i> sp. (r)	<i>Daldinia eschscholzii</i>	83
New Zealand	<i>Apophlaea</i> sp. (r) <i>Xiphophora gladiata</i> (b)	<i>Mycosphaerella apophlaeae</i> <i>Penicillium</i> sp.	59 36
Unknown location	<i>Ascophyllum nodosum</i> (b)	<i>Mycosphaerella ascophylli</i> <i>Mycosphaerella ascophylli</i> <i>Phaeosphaeria spartinae</i> <i>Epicoccum</i> sp. <i>Drechslera dematioidea</i> <i>Chaetomium</i> sp.	45 46 40 13 73 67

(r) denotes red algae belonging to the division Rhodophyta; (b) denotes brown algae belonging to the class Phaeophyceae and (g) denotes green algae belonging to the division Chlorophyta.

Fungal endophytes – biology and bioprospecting

Table 2. Bioactivity of fungal endophytes isolated from marine macroalgal hosts

Bioactivity	Fungal endophyte	Algal host	Reference
AChE modulator	<i>Aspergillus flavus</i>	<i>Corallina officinalis</i> (r)	31
	<i>Aspergillus oryzae</i>	<i>Heterosiphonia japonica</i> (r)	29
Antialgal activity	<i>Aspergillus niger</i>	<i>Halimeda macroloba</i> (g)	
	<i>Aspergillus niger</i>	<i>Ulva lactuca</i> (g)	
	<i>Aspergillus terreus</i>	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Aspergillus terreus</i>	<i>Caulerpa sertularioides</i> (g)	
	<i>Aspergillus terreus</i>	<i>Gracilaria edulis</i> (r)	
	<i>Aspergillus terreus</i>	<i>Halimeda macroloba</i> (g)	
	<i>Aspergillus terreus</i>	<i>Ulva lactuca</i> (g)	
	<i>Aspergillus terreus</i>	<i>Sargassum ilicifolium</i> (b)	
	<i>Aspergillus</i> sp. 2	<i>Halimeda macroloba</i> (g)	
	<i>Aspergillus</i> sp. 4	<i>Caulerpa racemosa</i> (g)	
	<i>Aspergillus</i> sp. 4	<i>Sargassum wightii</i> (b)	
	<i>Chaetomium</i> sp.	<i>Ulva lactuca</i> (g)	
	<i>Chaetomium</i> sp. 1	<i>Padina tetrastromatica</i> (b)	
	<i>Chaetomium</i> sp. 1	<i>Halymenia</i> sp. (r)	
	<i>Fusarium</i> sp.	<i>Sargassum wightii</i> (b)	
	<i>Fusarium</i> sp. 2	<i>Caulerpa sertularioides</i> (g)	
	<i>Geniculosporium</i> sp.	<i>Polysiphonia</i> sp. (r)	61
	<i>Nigrospora</i> sp.	<i>Ulva lactuca</i> (g)	9
	<i>Nigrospora</i> sp.	<i>Gracilaria edulis</i> (r)	
	<i>Paecilomyces</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Penicillium</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Penicillium</i> sp. 1	<i>Halimeda macroloba</i> (g)	
	<i>Trichoderma</i> sp.	<i>Halimeda macroloba</i> (g)	
<i>Trichoderma</i> sp.	<i>Sargassum wightii</i> (b)		
Sterile form 1	<i>Portieria hornemanii</i> (r)		
Sterile form 2	<i>Sargassum wightii</i> (b)		
Sterile form 3	<i>Lobophora variegata</i> (b)		
Anticancer activity	<i>Alternaria</i> sp.	<i>Ulva linz</i> (g)	23
	<i>Apiospora montagnei</i>	<i>Polysiphonia violacea</i> (r)	58
	<i>Arthrimum</i> sp.	<i>Fucus</i> sp. (b)	23
	<i>Aspergillus niger</i>	<i>Colpomenia sinuosa</i> (b)	89
	<i>Aspergillus ochraceus</i>	<i>Sargassum kjellmanianum</i> (b)	27
	<i>Aspergillus versicolor</i>	<i>Codium fragile</i> (g)	66
	<i>Aspergillus wentii</i>	<i>Sargassum fusiforme</i> (b)	70
	<i>Chaetomium globosum</i>	<i>Ulva pertusa</i> (g)	35
	<i>Chaetomium</i> sp.	<i>Ceramium</i> sp. (r)	23
	<i>Colletotrichum</i> sp.	<i>Polysiphonia</i> sp. (r)	
	<i>Corollospora</i> sp.	<i>Sargassum</i> sp. (b)	
	<i>Emericella nidulans</i> var. <i>acristata</i>	Unidentified	60
	<i>Geniculosporium</i> sp.	<i>Polysiphonia</i> sp. (r)	23
	<i>Guignardia</i> sp.	<i>Undaria pinnatifida</i> (b)	85
	<i>Gibberella zeae</i>	<i>Codium fragile</i> (g)	65
	<i>Monodictys putredinis</i>	Unidentified green alga	25
	<i>Nodulisporium</i> sp.	Unidentified	76
	<i>Penicillium chrysogenum</i>	<i>Sargassum palladium</i> (b)	34
	<i>Penicillium chrysogenum</i>	<i>Laurencia</i> sp. (r)	50
	<i>Penicillium</i> sp.	<i>Ulva</i> sp. (g)	47
	<i>Penicillium</i> sp.	<i>Fucus vesiculosus</i> (b)	23
	<i>Phoma</i> sp.	<i>Fucus</i> sp. (b)	
	<i>Phoma</i> sp.	<i>Fucus vesiculosus</i> (b)	
<i>Penicillium</i> sp.	<i>Xiphophora gladiata</i> (b)	36	
<i>Tolypocladium inflatum</i>	<i>Fucus vesiculosus</i> (b)	23	
Unidentified fungus 1	<i>Turbinaria conoides</i> (b)	10	
Unidentified fungus 2	<i>Turbinaria conoides</i> (b)		
Unidentified fungus 4	<i>Caulerpa lentillifera</i> (g)		
Unidentified fungus 11	<i>Caulerpa racemosa</i> variant (g)		
Unidentified fungus 12	<i>Sargassum oligocystum</i> (b)		
Unidentified fungus 13	<i>Sargassum oligocystum</i> (b)		

(Contd)

Table 2. (Contd)

Bioactivity	Fungal endophyte	Algal host	Reference
	Unidentified fungus 14	<i>Sargassum oligocystum</i> (b)	
	Unidentified fungus 15	<i>Sargassum oligocystum</i> (b)	
Antimicrobial activity	<i>Alternaria</i> sp.	<i>Sargassum wighii</i> (b)	9
	<i>Aspergillus fumigatus</i> II	<i>Fucus vesiculosus</i> (b)	12
	<i>Aspergillus fumigatus</i> III	<i>Fucus vesiculosus</i> (b)	
	<i>Aspergillus fumigatus</i> IV	<i>Fucus vesiculosus</i> (b)	
	<i>Aspergillus fumigatus</i> V	<i>Fucus serratus</i> (b)	
	<i>Aspergillus fumigatus</i> VI	<i>Fucus serratus</i> (b)	12
	<i>Aspergillus niger</i>	<i>Colpomenia sinuosa</i> (b)	16, 17, 18, 19, 89
	<i>Aspergillus niger</i>	<i>Halimeda macroloba</i> (g)	9
	<i>Aspergillus oryzae</i>	<i>Heterosiphonia japonica</i> (r)	30
	<i>Aspergillus sydowii</i>	<i>Porphyra umbilicalis</i> (r)	11
	<i>Aspergillus terreus</i>	<i>Caulerpa sertularioides</i> (g)	9
	<i>Aspergillus terreus</i>	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Aspergillus terreus</i>	<i>Gracilaria edulis</i> (r)	
	<i>Aspergillus terreus</i>	<i>Ulva lactuca</i> (g)	
	<i>Aspergillus terreus</i>	<i>Halimeda macroloba</i> (g)	
	<i>Aspergillus versicolor</i>	<i>Codium fragile</i> (g)	66
	<i>Aspergillus versicolor</i>	<i>Sargassum thunbergii</i> (b)	68
	<i>Aspergillus wentii</i>	<i>Gymnogongrus flabelliformis</i> (r)	81
	<i>Aspergillus wentii</i>	<i>Sargassum</i> sp. (b)	80
	<i>Aspergillus</i> sp. 2	<i>Caulerpa scalpelliformis</i> (g)	9
	<i>Aspergillus</i> sp. 2	<i>Halimeda macroloba</i> (g)	
	<i>Aspergillus</i> sp. 4	<i>Sargassum wighii</i> (b)	
	<i>Aspergillus</i> sp. 10	<i>Sargassum wighii</i> (b)	
	<i>Aspergillus</i> sp. I	<i>Chondrus crispus</i> (r)	11
	<i>Aspergillus</i> sp. II	<i>Ascophyllum nodosum</i> (b)	
	<i>Aspergillus</i> sp. III	<i>Saccharina latissima</i> (b)	
	<i>Aureobasidium pullulans</i> I	<i>Devaleraea ramentacea</i> (r)	
	<i>Aureobasidium pullulans</i> II	<i>Polysiphonia lanosa</i> (r)	
	<i>Botryotinia fuckeliana</i>	<i>Polysiphonia lanosa</i> (r)	
	<i>Botrytis</i> sp. I	<i>Devaleraea ramentacea</i> (r)	
	<i>Botrytis</i> sp. II	<i>Devaleraea ramentacea</i> (r)	
	<i>Botrytis</i> sp. III	<i>Palmaria palmata</i> (r)	
	<i>Chaetomium</i> sp.	Unidentified	67
	<i>Chaetomium</i> sp. 1	<i>Halymenia</i> sp. (r)	9
	<i>Chaetomium</i> sp. 1	<i>Padina tetrastromatica</i> (b)	
	<i>Cladosporium</i> sp.	<i>Devaleraea ramentacea</i> (r)	11
	<i>Cladosporium</i> sp. 1	<i>Portieria hornemanii</i> (r)	9
	<i>Cladosporium</i> sp. 1	<i>Sargassum wighii</i> (b)	
	<i>Cladosporium</i> sp. 1	<i>Caulerpa racemosa</i> (g)	
	<i>Cladosporium</i> sp. II	<i>Plocamium cartilagineum</i> (r)	12
	<i>Coniothyrium cereale</i>	<i>Enteromorpha</i> sp. (g)	20
	<i>Coniothyrium</i> sp.	<i>Devaleraea ramentacea</i>	11
	<i>Coniothyrium</i> sp. I	<i>Fucus vesiculosus</i> (b)	
	<i>Coniothyrium</i> sp. II	<i>Fucus serratus</i> (b)	
	<i>Coniothyrium</i> sp. III	<i>Fucus serratus</i> (b)	
	<i>Curvularia</i> sp. 1	<i>Sargassum wighii</i> (b)	9
	<i>Curvularia</i> sp. 3	<i>Sargassum wighii</i> (b)	
	<i>Daldinia eschscholzii</i>	<i>Gracilaria</i> sp. (r)	83
	<i>Emericella nidulans</i>	<i>Sargassum wighii</i> (b)	9
	<i>Emericella nidulans</i>	<i>Halymenia</i> sp. (r)	
	<i>Eurotium cristatum</i>	<i>Sargassum thunbergii</i> (b)	39
	<i>Eurotium</i> sp.	<i>Ulva lactuca</i> (g)	12
	<i>Fusarium</i> sp.	<i>Sargassum wighii</i> (b)	9
	<i>Fusarium</i> sp. 2	<i>Caulerpa sertularioides</i> (g)	
	<i>Guignardia</i> sp.	<i>Undaria pinnatifida</i> (b)	85
	<i>Helicomycetes</i> sp.	<i>Palmaria palmata</i> (r)	11
	<i>Hypoxylon</i> sp.	<i>Palmaria palmata</i> (r)	
	<i>Leptosphaeria</i> sp.	<i>Ulva intestinalis</i> (g)	12
	<i>Microdochium</i> sp.	<i>Porphyra</i> sp. (r)	

(Contd)

Fungal endophytes – biology and bioprospecting

Table 2. (Contd)

Bioactivity	Fungal endophyte	Algal host	Reference
	<i>Nigrospora</i> sp.	<i>Gracilaria edulis</i> (r)	9
	<i>Paecilomyces</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Penicillium biourgeianum</i>	<i>Plocamium cartilagineum</i> (r)	12
	<i>Penicillium chrysogenum</i>	<i>Laurencia</i> sp. (r)	49, 50, 51
	<i>Penicillium chrysogenum</i> I	<i>Palmaria palmata</i> (r)	11
	<i>Penicillium chrysogenum</i> II	<i>Saccharina latissima</i> (b)	
	<i>Penicillium crustosum</i> I	<i>Chondrus crispus</i> (r)	
	<i>Penicillium crustosum</i> II	<i>Palmaria palmata</i> (r)	
	<i>Penicillium decumbens</i> I	<i>Devaleraea ramentacea</i> (r)	
	<i>Penicillium decumbens</i> II	<i>Mastocarpus stellatus</i> (r)	
	<i>Penicillium decumbens</i> III	<i>Palmaria palmata</i> (r)	
	<i>Penicillium echinulatum</i>	<i>Chondrus ocellatus</i> (r)	64
	<i>Penicillium glabrum</i>	<i>Sargassum thunbergii</i> (b)	8
	<i>Penicillium soppii</i> II	<i>Spongomorpha arcta</i> (g)	11
	<i>Penicillium spinulosum</i>	<i>Spongomorpha arcta</i> (g)	
	<i>Penicillium</i> sp.	<i>Xiphophora gladiata</i> (b)	36
	<i>Penicillium</i> sp.	<i>Caulerpa scalpelliformis</i> (g)	9
	<i>Penicillium</i> sp. 1	<i>Portieria hornemanii</i> (r)	
	<i>Penicillium</i> sp. 1	<i>Caulerpa racemosa</i> (g)	
	<i>Penicillium</i> sp. 1	<i>Caulerpa sertularioides</i> (g)	
	<i>Penicillium</i> sp. 1	<i>Halimeda macroloba</i> (g)	
	<i>Penicillium</i> sp. I	<i>Fucus vesiculosus</i> (b)	12
	<i>Penicillium</i> sp. II	<i>Ulva lactuca</i> (g)	
	<i>Penicillium</i> sp. III	<i>Ulva intestinalis</i> (g)	
	<i>Penicillium</i> sp. IV	<i>Ulva intestinalis</i> (g)	
	<i>Penicillium</i> sp. V	<i>Fucus spiralis</i> (b)	12, 44
	<i>Penicillium</i> sp. VII	<i>Fucus serratus</i> (b)	12
	<i>Penicillium</i> sp. VIII	<i>Ulva intestinalis</i> (g)	
	<i>Phaeosphaeria</i> sp.	<i>Polysiphonia lanosa</i> (r)	
	<i>Phomopsis</i> sp.	<i>Portieria hornemanii</i> (r)	9
	<i>Stilbella fimetaria</i>	<i>Ulva lactuca</i> (g)	12
	<i>Trametes versicolor</i>	<i>Devaleraea ramentacea</i> (r)	11
	<i>Trichoderma longibrachiatum</i>	<i>Codium fragile</i> (g)	69
	<i>Trichoderma</i> sp.	<i>Sargassum wightii</i> (b)	9
	Black hyaline I	<i>Fucus spiralis</i> (b)	11
	Black hyaline II	<i>Saccharina latissima</i> (b)	
	Pigmented filamentous	<i>Ulva intestinalis</i> (g)	12
	Pigmented hyaline I	<i>Fucus spiralis</i> (b)	11
	Pigmented hyaline IV (Red)	<i>Saccharina latissima</i> (b)	
	Pigmented hyaline V	<i>Spongomorpha arcta</i> (g)	
	Pigmented hyaline VII	<i>Spongomorpha arcta</i> (g)	
	Red yeast I	<i>Mastocarpus stellatus</i> (r)	
	Sterile beige II	<i>Saccharina latissima</i> (b)	
	Sterile beige III	<i>Porphyra</i> sp. (r)	12
	Sterile beige IV	<i>Porphyra</i> sp. (r)	
	Sterile beige VII	<i>Plocamium cartilagineum</i> (r)	
	Sterile beige VIII	<i>Fucus spiralis</i> (b)	
	Sterile beige X	<i>Fucus spiralis</i> (b)	
	Sterile beige XI	<i>Fucus spiralis</i> (b)	
	Sterile beige XII	<i>Polysiphonia lanosa</i> (r)	
	Sterile beige XIII	<i>Polysiphonia lanosa</i> (r)	
	Septate hyaline I	<i>Ulva intestinalis</i> (g)	
	Septate pigmented I	<i>Devaleraea ramentacea</i> (r)	11
	Septate pigmented III	<i>Devaleraea ramentacea</i> (r)	
	Septate pigmented IV	<i>Mastocarpus stellatus</i> (r)	
	Septate pigmented V	<i>Fucus spiralis</i> (b)	
	Septate pigmented VI	<i>Ulva intestinalis</i> (g)	
	Septate pigmented VII	<i>Ulva lactuca</i> (g)	
	Septate pigmented VIII	<i>Spongomorpha arcta</i> (g)	
	Sterile hyaline I	<i>Devaleraea ramentacea</i> (r)	
	Sterile hyaline V	<i>Palmaria palmata</i> (r)	

(Contd)

Table 2. (Contd)

Bioactivity	Fungal endophyte	Algal host	Reference
	Sterile hyaline VI	<i>Palmaria palmata</i> (r)	
	Sterile hyaline VII	<i>Palmaria palmata</i> (r)	
	Sterile hyaline IX	<i>Polysiphonia lanosa</i> (r)	
	Sterile hyaline X	<i>Polysiphonia lanosa</i> (r)	
	Sterile hyaline XI	<i>Porphyra purpurea</i> (r)	
	Sterile hyaline XII	<i>Porphyra purpurea</i> (r)	
	Sterile hyaline XIII	<i>Porphyra umbilicalis</i> (r)	
	Sterile hyaline XIV	<i>Porphyra umbilicalis</i> (r)	
	Sterile hyaline XVII	<i>Spongomorpha arcta</i> (g)	
	Sterile hyaline XIX	<i>Ulva intestinalis</i> (g)	
	Sterile hyaline XX	<i>Ulva lactuca</i> (g)	
	Sterile pigmented I	<i>Ulva lactuca</i> (g)	12
	Sterile pigmented II	<i>Ulva intestinalis</i> (g)	
	Sterile pigmented V	<i>Plocamium cartilagineum</i> (r)	
	Sterile pigmented VI	<i>Fucus spiralis</i> (b)	
	Sterile pigmented VII	<i>Polysiphonia lanosa</i> (r)	
	Sterile pigmented VIII	<i>Polysiphonia lanosa</i> (r)	
	Sterile pigmented IX	<i>Ulva intestinalis</i> (g)	
	White fluffy II	<i>Devaleraea ramentacea</i> (r)	11
	White hyaline II	<i>Saccharina latissima</i> (b)	
	White hyaline III	<i>Spongomorpha arcta</i> (g)	
	White hyaline IV	<i>Saccharina latissima</i> (b)	
	Sterile form 1	<i>Portieria hornemanii</i> (r)	9
	Sterile form 1	<i>Caulerpa racemosa</i> (g)	
	Sterile form 2	<i>Sargassum wightii</i> (b)	
	Sterile form 3	<i>Turbinaria decurrens</i> (b)	
	Unidentified fungus	<i>Sargassum</i> sp. (b)	88
	Unidentified fungus 1	<i>Rhodomela confervoides</i> (r)	8
	Unidentified fungus 2	<i>Gelidium amansii</i> (r)	
	Unidentified fungus 3	<i>Gracilaria lemaneiformis</i> (r)	
	Unidentified fungus 4	<i>Gracilaria lemaneiformis</i> (r)	
	Unidentified fungus 5	<i>Sargassum thunbergii</i> (b)	
	Unidentified fungus 6	<i>Sargassum thunbergii</i> (b)	
	Unidentified fungus 9	<i>Enteromorpha prolifera</i> (g)	
	Unidentified fungus 10	<i>Enteromorpha prolifera</i> (g)	
	Unidentified fungus 11	<i>Ulva pertusa</i> (g)	
	Unidentified fungus 12	<i>Ulva pertusa</i> (g)	
Antioxidant activity	<i>Acremonium</i> sp.	<i>Cladostephus spongius</i> (b)	15
	<i>Aspergillus niger</i>	<i>Caulerpa scalpelliformis</i> (g)	9
	<i>Aspergillus niger</i>	<i>Colpomenia sinuosa</i> (b)	17
	<i>Aspergillus ochraceus</i>	<i>Sargassum kjellmanianum</i> (b)	28
	<i>Aspergillus terreus</i>	<i>Caulerpa scalpelliformis</i> (g)	9
	<i>Aspergillus terreus</i>	<i>Gracilaria edulis</i> (r)	
	<i>Aspergillus terreus</i>	<i>Sargassum ilicifolium</i> (b)	
	<i>Aspergillus terreus</i>	<i>Ulva lacuca</i> (g)	
	<i>Aspergillus</i> sp. 2	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Aspergillus wentii</i>	<i>Sargassum</i> sp. (b)	63
	<i>Chaetomium globosum</i>	<i>Polysiphonia urceolata</i> (r)	86
	<i>Curvularia</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)	9
	<i>Epicoccum</i> sp.	<i>Fucus vesiculosus</i> (b)	13
	<i>Fusarium</i> sp. 4	<i>Caulerpa racemosa</i> (g)	9
	<i>Guignardia</i> sp.	<i>Undaria pinnatifida</i> (b)	85
	<i>Paecilomyces</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)	9
	<i>Penicillium</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)	
	<i>Penicillium</i> sp. 1	<i>Halimeda macroloba</i> (g)	
	Sterile form 1	<i>Caulerpa racemosa</i> (g)	
Antiplasmodial activity	<i>Ascochyta salicorniae</i>	<i>Ulva</i> sp. (g)	72
	<i>Chaetomium</i> sp.	Unidentified	77
	<i>Drechslera dematioidea</i>	<i>Liagora viscida</i> (r)	73
Inhibition of β -glucuronidase	<i>Aspergillus terreus</i>	<i>Laurencia ceylanica</i> (r)	57

(Contd)

Fungal endophytes – biology and bioprospecting

Table 2. (Contd)

Bioactivity	Fungal endophyte	Algal host	Reference	
Inhibition of HLE	<i>Coniothyrium cereale</i>	<i>Enteromorpha</i> sp. (g)	21	
	<i>Phaeosphaeria spartinae</i>	<i>Ceramium</i> sp. (r)	40, 41	
Insecticidal activity	<i>Aspergillus niger</i>	<i>Caulerpa racemosa</i> (g)	9	
	<i>Aspergillus niger</i>	<i>Ulva lactuca</i> (g)		
	<i>Aspergillus terreus</i>	<i>Caulerpa scalpelliformis</i> (g)		
	<i>Aspergillus terreus</i>	<i>Caulerpa sertularioides</i> (g)		
	<i>Aspergillus terreus</i>	<i>Gracilaria edulis</i> (r)		
	<i>Aspergillus terreus</i>	<i>Halimeda macroloba</i> (g)		
	<i>Aspergillus terreus</i>	<i>Sargassum ilicifolium</i> (b)		
	<i>Aspergillus terreus</i>	<i>Ulva lactuca</i> (g)		
	<i>Aspergillus</i> sp. 2	<i>Halimeda macroloba</i> (g)		
	<i>Chaetomium</i> sp. 1	<i>Ulva lactuca</i> (g)		
	<i>Cladosporium</i> sp. 1	<i>Caulerpa racemosa</i> (g)		
	<i>Fusarium</i> sp. 2	<i>Caulerpa sertularioides</i> (g)		
	<i>Fusarium</i> sp. 4	<i>Caulerpa racemosa</i> (g)		
	<i>Paecilomyces</i> sp. 1	<i>Caulerpa scalpelliformis</i> (g)		
Protein phosphatase inhibitor	<i>Ascochyta salicorniae</i>	<i>Ulva</i> sp. (g)	78	
	Tyrosine kinase inhibitor	<i>Chaetomium</i> sp.	<i>Valonia utricularis</i> (g)	32
		<i>Drechslera dematioidea</i>	<i>Liagora viscida</i> (r)	73
		<i>Pestalotiopsis</i> sp.	<i>Sargassum horneri</i> (b)	26
	<i>Wardomyces anomalus</i>	<i>Enteromorpha</i> sp. (g)	14	

(r), (b) and (g), same as in Table 1.

UK). It should also be noted that red algae represent the greatest proportion of hosts investigated, with 41 species studied (Table 1). The number of hosts studied decreases substantially for the brown and green algae (32 and 19 respectively; Table 1). In the context of the conservative estimates by Guiry²² (7000 red, 8000 green and 2000 brown algae worldwide), the 100 algal hosts investigated to date represent <1% of the available sources of endophytes. This suggests that much work remains to be done on the distribution of algal endophytes and their role as a promising source of chemical diversity.

It should be noted that several species have been studied in multiple locations (Table 1). Two studies performed by Flewelling *et al.*^{11,12} investigated *Ascophyllum nodosum*, *Fucus spiralis*, *Fucus vesiculosus*, *Polysiphonia lanosa*, *Ulva lactuca* and *Ulva intestinalis* from both the Bay of Fundy, Canada and the Shetland Islands, UK. *Sargassum* spp. have also been investigated in multiple locations, including China, India, USA and France (Table 1).

Fungal biodiversity

A screening of marine macroalgae from the southern Indian coast for their endophytic fungi by Suryanarayanan *et al.*⁹ indicated that from a total sample size of 281 fungal isolates (representing 72 distinct species), green

algae yielded the lowest diversity of endophytic fungal species, whereas brown algae yielded the highest diversity. This is in contrast to the findings of Flewelling *et al.*¹², where over one quarter of the distinct fungi isolated came from the green algae, *U. lactuca* and *U. intestinalis*. In order to continue analysing this trend, further work is required globally, through the isolation of a larger number of fungal isolates from a greater sampling of macroalgae, to identify if there are individual species differences within the groups of macroalgae (red, green, brown), or to see whether the global geographic, climatic or seasonal differences may account for differences in fungal diversity.

Of the green algae studied thus far, *Caulerpa* spp. and *Ulva* spp. have the most diverse endophytic fungal symbionts with the majority recorded by Suryanarayanan *et al.*⁹ and Flewelling *et al.*¹² in India and Shetland Islands respectively (Table 1). Forty-one red algae have been found to host fungal endophytes (Table 1). Of the red algae studied to date, *Portieria hornemanii*, *Gelidiella acerosa*, *Gracilaria* spp. and *Halymenia* spp. have displayed the greatest diversity of fungal endophytes, with the majority found in India⁹. Thirty-two species of brown algae have been shown to host fungal endophytes (Table 1). *Fucus* spp., *Padina* spp., *Sargassum* spp. and *Turbinaria* spp. have displayed the greatest diversity of endophytic fungi. *Alternaria* spp., *Chaetomium* spp. and *Cladosporium* spp. appear to lack host specificity, having been isolated as fungal endophytes from numerous locations and hosts

Table 3. Summary of the number of compounds isolated from endophytic fungi of marine macroalgae

Bioactivity	Fungal endophyte	Algal host	Number of compounds identified	Reference
AChE modulator	<i>Aspergillus flavus</i>	<i>Corallina officinalis</i> (r)	(8 <i>E</i> ,12 <i>Z</i>)-10,11-dihydroxyoctadeca-8,12-dienoic acid 3 β , 4 α -dihydroxy-26-methoxyergosta-7,24(28)-dien-6-one Episterol (22 <i>E</i> ,24 <i>R</i>)-ergosta-7,22-dien-3 β , 5 α ,6 α -triol (22 <i>E</i> ,24 <i>R</i>)-ergosta-5,22-dien-3 β -ol (22 <i>E</i> ,24 <i>R</i>)-ergosta-4,6,8(14),22-tetraen-3-one	31
	<i>Aspergillus oryzae</i>	<i>Heterosiphonia japonica</i> (r)	Asporyergosterol (22 <i>E</i> ,24 <i>R</i>)-ergosta-4,6,8(14),22-tetraen-3-one (22 <i>E</i> ,24 <i>R</i>)-3 β -hydroxyergosta-5,8,22-trien-7-one (22 <i>E</i> ,24 <i>R</i>)-ergosta-7,22-dien-3 β , 5 α ,6 β -triol (22 <i>E</i> ,24 <i>R</i>)-5 α ,8 α -epidioxyergosta-6,22-dien-3 β -ol	29
Antialgal	<i>Geniculosporium</i> sp.	<i>Polysiphonia</i> sp. (r)	7-Hydroxy-10-methoxydeacetyldihydrobotrydial 7-Hydroxy-10-oxodehydrodihydrobotrydial 7,10-Dihydroxydehydrodihydrobotrydial 7-Hydroxy-10-methoxydehydrodihydrobotrydial 7-Hydroxy-10-ethoxydehydrodihydrobotrydial 7-Hydroxy-10-dehydroxydehydrodihydrobotrydial 7-Hydroxydeacetylbotryenalol 7,10-Dihydroxydeacetyldihydrobotrydial-1(10)-ene 4,10-Didehydroxy-7-hydroxydeacetyldihydrobotrydial-1 (10),5(9)-diene 7-Hydroxy-10-dehydroxydeacetyldihydrobotrydial-1(10), 5(9)-diene 15 α -Hydroxy-14-aldehyde probotryan-4(5)-ene L-696,474 Cytochalasin U RKS-1778 Cytochalasin H	61
Anticancer	<i>Apiospora montagnei</i>	<i>Polysiphonia violacea</i> (r)	Myrocin A Apiosporic acid 9-Hydroxyhexylitaconic acid (-)-Hexylitaconic acid (+)-Epiepoxydon	58
	<i>Aspergillus niger</i>	<i>Colpomenia sinuosa</i> (b)	Isopyrophen Aspergillusol Pyrophen Cyclo-(L-Trp-L-Ile) Cyclo-(L-Trp-L-Phe) Cyclo-(L-Trp-L-Tyr)	89
	<i>Aspergillus ochraceus</i>	<i>Sargassum kjellmanianum</i> (b)	7-Nor-ergosterolide 3 β ,11 α -dihydroxyergosta-8,24(28)-dien-7-one 3 β -hydroxyergosta-8,24(28)-dien-7-one	27
	<i>Aspergillus wentii</i>	<i>Sargassum fusiforme</i> (b)	Aspewentin A Aspewentin B Aspewentin C	70
	<i>Aspergillus wentii</i>	<i>Sargassum</i> sp. (b)	Wentilactone B	90
	<i>Chaetomium globosum</i>	<i>Ulva pertusa</i> (g)	Cytoglobosin A Cytoglobosin B Cytoglobosin C Cytoglobosin D Cytoglobosin E Cytoglobosin F Cytoglobosin G Isochaetoglobosin D Chaetoglobosin F _{ex}	35
	<i>Emericella nidulans</i> var. <i>acristata</i>	Unknown green alga	Arugosin G Arugosin H Arugosin A Arugosin B	60

(Contd)

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Table 3. (Contd)

Bioactivity	Fungal endophyte	Algal host	Number of compounds identified	Reference
	<i>Gibberella zeae</i>	<i>Codium fragile</i> (g)	Shamixanthone Emericellin Emindole DA Microperfuraneone Sterigmatocystin 3-Hydroxy-5-(hydroxymethyl)-4-(4'-hydroxyphenoxy)pyrrolidin-2-one (22 <i>E</i> ,24 <i>R</i>)-7 <i>β</i> ,8 <i>β</i> -epoxy-3 <i>β</i> ,5 <i>α</i> ,9 <i>α</i> -trihydroxyergosta-22-en-6-one (22 <i>E</i> ,24 <i>R</i>)-3 <i>β</i> ,5 <i>α</i> ,9 <i>α</i> -trihydroxyergosta-7,22-dien-6-one (22 <i>E</i> ,24 <i>R</i>)-3 <i>β</i> ,5 <i>α</i> -dihydroxyergosta-7,22-dien-6-one (22 <i>E</i> ,24 <i>R</i>)-ergosta-7,22-dien-3 <i>β</i> ,5 <i>α</i> ,6 <i>β</i> -triol (22 <i>E</i> ,24 <i>R</i>)-ergosta-5,22-dien-3 <i>β</i> -ol (22 <i>E</i> ,24 <i>R</i>)-5 <i>α</i> ,8 <i>α</i> -epidioxyergosta-6,22-dien-3 <i>β</i> -ol (22 <i>E</i> ,24 <i>R</i>)-5 <i>α</i> ,8 <i>α</i> -epidioxyergosta-6,9(11),22-trien-3 <i>β</i> -ol (22 <i>E</i> ,24 <i>R</i>)-1(10 → 6)- <i>abeo</i> -ergosta-5,7,9,22-tetraen-3 <i>α</i> -ol	65
	<i>Monodictys putredinis</i>	Unknown green alga	Monodictysin A Monodictysin B Monodictysin C Monodictyxanthone Monodictyphenone	25
	<i>Nodulisporium</i> sp.	Unknown	Noduliprevenone	76
	<i>Penicillium chrysogenum</i>	<i>Sargassum palladium</i> (b)	Chrysotriazole A Chrysotriazole B 2-(4-Hydroxybenzoyl)-4(3 <i>H</i>)-quinazolinone 2-(4-Hydroxybenzyl)quinazolin-4(3 <i>H</i>)-one <i>N</i> -[2-(4-Hydroxyphenyl)acetyl]formamide 2-(4-Hydroxyphenyl)acetamide <i>N</i> -[(2 <i>E</i>)-(4-Hydroxyphenyl)ethenyl]formamide <i>N</i> -[(2 <i>Z</i>)-(4-Hydroxyphenyl)ethenyl]formamide	34
	<i>Penicillium</i> sp.	<i>Ulva</i> sp. (g)	Chromanone A	47
Anticancer; antimicrobial	<i>Aspergillus versicolor</i>	<i>Codium fragile</i> (g)	Albican-11,14-diol Sterigmatocystin 3-Hydroxy-5-(hydroxymethyl)-4-(4'-hydroxyphenoxy)pyrrolidin-2-one (1 <i>H</i> -Indol-3-yl) oxoacetamide Indole-3-carboxylic acid Indole-3-acetic acid Indole-3-carboxaldehyde Volemolide	66
	<i>Penicillium chrysogenum</i>	<i>Laurencia</i> sp. (r)	Penicisteroid A Penicisteroid B Anicequol (22 <i>E</i> ,24 <i>R</i>)-ergosta-4,6,8(14),22-tetraen-3-one (22 <i>E</i> ,24 <i>R</i>)-ergosta-7,22-dien-3,6-dione (22 <i>E</i> ,24 <i>R</i>)-5 <i>α</i> ,8 <i>α</i> -epidioxyergosta-6,22-dien-3 <i>β</i> -ol (22 <i>E</i> ,24 <i>R</i>)-ergosta-5 <i>α</i> ,6 <i>α</i> -epoxide-8,22-dien-3 <i>β</i> ,7 <i>α</i> -diol (22 <i>E</i> ,24 <i>R</i>)-ergosta-7,22-dien-3 <i>β</i> ,5 <i>α</i> ,6 <i>β</i> -triol (22 <i>E</i> ,24 <i>R</i>)-ergosta-7,22-dien-3 <i>β</i> ,6 <i>β</i> -diol	50
	<i>Penicillium</i> sp.	<i>Xiphophora gladiata</i> (b)	PF1140 Deoxy-PF1140	36
	<i>Guignardia</i> sp.	<i>Undaria pinnatifida</i> (b)	Deoxykanthomycin Ergosterol peroxide Ergosterol Cyclo-(Tyr-Leu) Cyclo-(Phe-Phe) Cyclo-(Val-Leu) Cyclo-(Phe-Pro) Cyclo-(Leu-Ile)	85

(Contd)

Table 3. (Contd)

Bioactivity	Fungal endophyte	Algal host	Number of compounds identified	Reference
Antimicrobial	<i>Aspergillus niger</i>	<i>Colpomenia sinuosa</i> (b)	Asperamide A	19
			Asperamide B	18
	<i>Aspergillus oryzae</i>	<i>Heterosiphonia japonica</i> (r)	5,7-Dihydroxy-2-[1-(4-methoxy-6-oxo-6H-pyran-2-yl)-2-phenylethylamino]-[1,4]naphthoquinone	
			Asporyzin A	30
	<i>Aspergillus versicolor</i>	<i>Sargassum thunbergii</i> (b)	Asporyzin B	
			Asporyzin C	
	<i>Aspergillus versicolor</i>	<i>Sargassum thunbergii</i> (b)	JBIR-03	
			Emindole SB	
			Emeniveol	
			Asperserin A	68
			9ξ-O-2(2,3-dimethylbut-3-enyl)brevianamide Q	
			Brevianamide K	
			Brevianamide M	
			Aversin	
			6,8-di-O-methylnidurufin	
			6,8-di-O-methylaverufin	
	<i>Aspergillus versicolor</i>	<i>Sargassum thunbergii</i> (b)	6-O-methylaverufin	
			5α,8α-Epidioxyergosta-6,22-dien-3β-ol	
	<i>Aspergillus wentii</i>	<i>Gymnogongrus flabelliformis</i> (r)	Ergosta-7,22-diene-3β,5α,6β-triol	68
			6β-methoxyergosta-7,22-diene-3β,5α-diol	81
	<i>Aspergillus wentii</i>	<i>Sargassum</i> sp. (b)	Yicathin A	
			Yicathin B	
			Yicathin C	
			Alatinone	
			1,5-Dihydroxy-3-methoxy-7-methylanthraquinone	
			5-Hydroxy-1,3-dimethoxy-7-methylanthraquinone	
			Asperolide A	80
			Asperolide B	
			Asperolide C	
			Tetranorditerpenoid derivative	
	<i>Chaetomium</i> sp.	Unknown	Wentilactone A	
			Wentilactone B	
			Botryosphaerin B	
			LL-Z1271-β	
	<i>Coniothyrium cereale</i>	<i>Enteromorpha</i> sp. (g)	Chaetocyclinone A	67
			Chaetocyclinone B	
			Chaetocyclinone C	
	<i>Coniothyrium cereale</i>	<i>Enteromorpha</i> sp. (g)	Coniosclerodin	20
			(Z)-Coniosclerodinol	
			(E)-Coniosclerodinol	
			(15S,17S)-(-)-Sclerodinol	
			Conioscleroderolide	
			Coniosclerodione	
			Coniolactone	
			(-)-7,8-Dihydro-3,6-dihydroxy-1,7,7,8-tetramethyl-5H-furo-[2',3':5,6]naphtho[1,8-bc]furan-5-one	
			(-)-Sclerodin A	
			Lamellicolic anhydride	20
			(-) Scleroderolide	
			(-) Sclerodione	
			<i>Daldinia eschscholzii</i>	<i>Gracilaria</i> sp. (r)
	<i>Eurotium cristatum</i>	<i>Sargassum thunbergii</i> (b)	Helicascolide A	
			Cristatumin A	39
			Cristatumin B	
			Cristatumin C	
			Cristatumin D	
			Neoechinulin A	
			Isoechinulin A	
			Variecolorin G	

(Contd)

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Table 3. (Contd)

Bioactivity	Fungal endophyte	Algal host	Number of compounds identified	Reference
	<i>Penicillium chrysogenum</i>	<i>Laurencia</i> sp. (r)	Preechinulin Tardioxopiperazine A Echinulin Penicitide A Penicitide B	49
	<i>Penicillium chrysogenum</i>	<i>Laurencia</i> sp. (r)	2-(2,4-Dihydroxy-6-methylbenzoyl)-glycerol 1-(2,4-Dihydroxy-6-methylbenzoyl)-glycerol Penicimonoterpene	51
	<i>Penicillium echinulatum</i>	<i>Chondrus ocellatus</i> (r)	Conidiogenone H Conidiogenone I Conidiogenone B Conidiogenone C Conidiogenone D Conidiogenone F Conidiogenol	64
	<i>Penicillium</i> sp.	<i>Fucus spiralis</i> (b)	Arisugacin K Arisugacin J Arisugacin G Arisugacin C Territrem C	44
	<i>Phomopsis longicolla</i>	<i>Bostrychia radicans</i> (r)	Cladosporin Epiepoformin Phyllostine Patulin	43
	<i>Trichoderma longibrachiatum</i>	<i>Codium fragile</i> (g)	18-deoxycytochalasin H Mycophenolic acid Dicerandrol C	69
	Unidentified fungus	<i>Sargassum</i> sp. (b)	Harziandione 6-Oxo-de- <i>O</i> -methylasiodiplodin (<i>E</i>)-9-Etheno-lasiodiplodin Lasiodiplodin de- <i>O</i> -Methylasiodiplodin 5-Hydroxy-de- <i>O</i> -methylasiodiplodin	88
Antimicrobial; Antioxidant	<i>Aspergillus niger</i>	<i>Colpomenia sinuosa</i> (b)	Nigerasperone A Nigerasperone B Nigerasperone C	17
Antioxidant	<i>Acremonium</i> sp.	<i>Cladostephus spongus</i> (b)	7-Isopropenylbicyclo[4.2.0]octa-1,3,5-triene-2,5-diol 7-Isopropenylbicyclo-[4.2.0]octa-1,3,5-triene-2,5-diol-5- β -D-glucopyranoside (3 <i>R</i> *,4 <i>S</i> *)-3,4-dihydroxy-7-methyl-3,4-dihydro-1(2 <i>H</i>)-naphthalenone (3 <i>S</i> *,4 <i>S</i> *)-3,4-Dihydroxy-7-methyl-3,4-dihydro-1(2 <i>H</i>)-naphthalenone 2-(1-Methylethylidene) pentanedioic acid pentanedioic acid 2-(1-methylethylidene)-5-methylester pentanedioic acid 2-(1-methylethylidene)-1-methyl ester Pentanedioic acid 2-(1-methylethenyl)-5-methyl ester 2-(1-Hydroxy-1-methyl)-2,3-dihydrobenzofuran-5-ol 2,2-Dimethylchroman-3, 6-diol 2-(3-Dihydroxy-3-methylbutyl)benzene-1,4-diol	15
	<i>Aspergillus ochraceus</i>	<i>Sargassum kjellmanianum</i> (b)	2-Hydroxycircumdatin C (1 <i>1aS</i>)-2,3-dihydro-7-methoxy-1 <i>H</i> -pyrrolo[2,1- <i>c</i>][1,4]benzodiazepine-5,11(10 <i>H</i> ,11 <i>aH</i>)-dione Circumdatin F Circumdatin C Circumdatin D Notoamide B Selerotiamide	28

(Contd)

Table 3. (Contd)

Bioactivity	Fungal endophyte	Algal host	Number of compounds identified	Reference
	<i>Aspergillus wentii</i>	<i>Sargassum</i> sp. (b)	Wentiquinone C Methyl 4-(3,4-dihydroxybenzamido)butanoate 5- <i>O</i> -Methylsulochine Methyl 2-(2,6-dimethoxy-4-methylbenzoyl)-3,5-dihydroxybenzoate Methyl-2-(2,6-dihydroxyl-4-methylbenzoyl)-3-hydroxy-5-methoxybenzoate Physcion 4-(3,4-Dihydroxybenzamido)butanoic acid (<i>E</i>)- <i>N</i> -(2-Hydroxy-2-(4-hydroxyphenyl)ethyl)-3-(3-hydroxy-4-methoxyphenyl)acrylamide	63
	<i>Chaetomium globosum</i>	<i>Polysiphonia urceolata</i> (r)	Chaetopyranin 2-(2',3'-epoxy-1',3'-heptadienyl)-6-hydroxy-5-(3-methyl-2-butenyl)benzaldehyde Isotetrahydroauroglaucin Erythroglaucin Parietin Asperentin 5'-Hydroxy-asperentin-8-methylether Asperentin-8-methyl ether 4'-Hydroxyasperentin 5'-Hydroxyasperentin Neoechinulin A	86
	<i>Epicoccum</i> sp.	<i>Fucus vesiculosus</i> (b)	4,5,6-Trihydroxy-7-methylphthalide 5-(Acetoxymethyl)-furan-2-carboxylic acid Furan-2-carboxylic acid 5-(Hydroxymethyl)-furan-2-carboxylic acid (-)-(3 <i>R</i> ,4 <i>S</i>)-4-hydroxymellein (-)-(3 <i>R</i>)-5-hydroxymellein	13
Antiplasmodial	<i>Ascochyta salicorniae</i>	<i>Ulva</i> sp. (g)	Ascosalipyrrolidinone A Ascosalipyrrolidinone B Ascosalipyrone Genistein 2,3-Dihydro-2-hydroxy-2,4-dimethyl-5- <i>trans</i> -propenylfuran-3-one	72
	<i>Chaetomium</i> sp.	Unknown	Chaetoxanthone A Chaetoxanthone B Chaetoxanthone C	77
Antiplasmodial; tyrosine kinase inhibitor	<i>Drechslera dematioidea</i>	<i>Liagora viscida</i> (r)	Isosativetriol Drechslerine A Drechslerine B Helminthosporol 9-Hydroxyhelminthosporol Drechslerine C Drechslerine D Drechslerine E Drechslerine F Drechslerine G <i>cis</i> -Sativenediol Sativene epoxide (+)-Secolongifolene diol Isocochlioquinone A Isocochlioquinone C Cochlioquinone B	73
Inhibition of β -glucuronidase	<i>Aspergillus terreus</i>	<i>Laurencia ceylanica</i> (r)	3-Hydroxy-4-(4-hydroxyphenyl)-5-methoxycarbonyl-5-(4-hydroxy-3-formylbenzyl)-2,5-dihydro-2-furanone Butyrolactone-1 6-Hydroxymellin	57

(Contd)

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Table 3. (Contd)

Bioactivity	Fungal endophyte	Algal host	Number of compounds identified	Reference
			(3 <i>R</i> ,4 <i>R</i>)-6,7-dimethoxy-4-hydroxymellin (+)-Territonin (+)-Territonin-A (+)-Asterrelenin (+)-Terrein Oleic acid Glucopyranosyl- β -sitosterol	
Inhibition of HLE	<i>Coniothyrium cereale</i>	<i>Enteromorpha</i> sp. (g)	(-)-Cereolactam (-)-Trypethelone (-)-Cereoaldomine	21
	<i>Phaeosphaeria spartinae</i>	<i>Ceramium</i> sp. (r)	Spartinol A Spartinol B Spartinol C Spartinol D Spartinoxide A82775C Iso-A82775C 4-Hydroxy-3-prenyl-benzoic acid Anofinic acid	40 41
Protein phosphatase inhibitor	<i>Ascochyta salicorniae</i>	<i>Ulva</i> sp. (g)	Ascolactone A Ascolactone B Hyalopyrone Ascochitine Ascochital Ascosalipyrene	78
Tyrosine kinase inhibitor	<i>Chaetomium</i> sp.	<i>Valonia utricularis</i> (g)	Chaetominedione 2-Furancarboxylic acid 5-(Hydroxymethyl)-2-furancarboxylic acid	32 26
	<i>Pestalotiopsis</i> sp.	<i>Sargassum horneri</i> (b)	1 β ,5 α ,6 α ,14-Tetraacetoxy-9 α -benzoyloxy-7 β H-eudesman-2 β ,11-diol 4 α ,5 α -Diacetoxy-9 α -benzoyloxy-7 β H-eudesman-1 β ,2 β ,11,14-tetraol	
	<i>Wardomyces anomalus</i>	<i>Enteromorpha</i> sp. (g)	2,3,6,8-Tetrahydroxy-1-methylxanthone 2,3,4,6,8-Pentahydroxy-1-methylxanthone 3,6,8-Trihydroxy-1-methylxanthone 5-(Hydroxymethyl)-2-furanocarboxylic acid	14

(r), (b) and (g), same as in Table 1.

belonging to each of the major algal groups (red, green, and brown; Tables 1 and 2). *Cladosporium* has also been found from the west coast of the Shetland Islands¹².

Many studies have included macroalgal hosts only reported to the genus level (Table 1). Such reporting creates obstacles to any potential comparisons of algal hosts and their associated endophytes. Despite this, many reports have documented the common and ubiquitous occurrence of several fungal species, such as *Aspergillus* (17%) and *Penicillium* (9%), which are the most commonly isolated fungal endophytes from marine macroalgae. These two genera (*Aspergillus* and *Penicillium*) have been reported from six and eight of the locations investigated respectively (Table 1). Interestingly, several fungal endophytes that were discovered in brown algae have not been previously reported from red or green algae. Among this group of endophytes are *Acremoniella* sp., *Arthrimum* sp., *Ascotricha* sp., *Chalara* sp., *Epicoccum* sp., *Haloguignardia irritans*, *Helicosporium* sp., *Lichtheimia corymbifera*,

Monodictys putredinis, *Oidiodendron* sp., *Pestalotiopsis* sp., *Phaeotrichoconis* sp., *Taeniolella* sp., *Tolypocladium inflatum* and *Varicosporium* sp.^{9,12,13,23–26}. Also noteworthy is the large proportion of unidentified and sterile forms of endophytic fungi (30%) designated morphology-based codes for identification (Table 1). Further work is required to improve procedures for molecular identification of sterile fungal isolates, as morphological descriptions do not allow for proper documentation of the fungal biodiversity present within marine macroalgae.

Bioactivity – screening

Few studies have performed large-scale screens on the bioactivity of endophytic fungi from marine macroalgae. The work by Suryanarayanan *et al.*⁹ in India represents one of the first studies that screened the chemical extracts of endophytic fungi from an array of macroalgal hosts for bioactivity (Tables 1 and 2). The majority of the

endophytic fungi tested possessed either antialgal, antimicrobial, insecticidal or antioxidant activity⁹ (Table 2). Several studies have since followed^{10–12}, which have shown endophytes to be promising sources of bioactive natural products. The work by Ariffin *et al.*¹⁰ resulted in the isolation of 15 endophytes from six species of macroalgae (Table 1), where over half of the fungal isolates exhibited anticancer properties, with some possessing antimicrobial activity (Table 2). Studies from the Atlantic coast of Canada¹¹ and the west coast of Shetland Islands¹² support the work by Suryanarayanan *et al.*⁹ and Ariffin *et al.*¹⁰, where 64 and 79 fungal endophytes were isolated respectively, and in each case, over half of the isolates obtained showed antimicrobial activity (Table 2).

Aspergillus and *Penicillium* spp. isolated from a range of hosts and locations have displayed an extensive variety of bioactivity, with numerous reports documenting antimicrobial, anticancer, anti-inflammatory, antialgal, anti-infective, antioxidant and insecticidal activity^{9,16–19,27–31} (Table 2). Numerous other species have been identified, including *Alternaria* spp., *Chaetomium* spp. and *Cladosporium* spp. (Tables 1 and 2). *Alternaria* spp. extracts have shown strong bioactivity in antimicrobial and anticancer tests^{9,12,23} (Table 2). Extracts from *Chaetomium* spp. have displayed a wide variety of bioactivity including antialgal, antimicrobial, anticancer and insecticidal^{9,23,32}. *Cladosporium* spp. extracts have been shown to produce secondary metabolites with antimicrobial, anticancer and insecticidal activity^{9,12}. *Nigrospora* spp. have been reported from a diverse array of macroalgal hosts (Table 1). Suryanarayanan *et al.*⁹ have found that several *Nigrospora* spp. possess antialgal and antimicrobial activity (Table 2).

Bioactivity – natural products

The screening of endophytes from marine macroalgae has shown them to be an excellent source of bioactive natural products against a range of biological targets (Table 2). Complementing these findings are over 50 studies focused on the isolation of new natural products from endophytes of macroalgae (Table 3). Over 300 natural products have been identified from 32 endophytic fungi, with 22% of the investigated fungi being from the genus *Aspergillus* (Table 3). These 32 endophytic fungi investigated were isolated from 35 marine macroalgae (13 red, 12 brown, 7 green and 3 unidentified) with *Sargassum* spp. being the predominant macroalgae studied (Table 3). Of the natural products published, 43% (139/327) were reported as new natural product entities suggesting there is a difference in chemical diversity between terrestrial and marines endophytes (Table 3) as well as overlap. Many of the identified natural products have shown a range of bioactivity, including but not limited to, antioxidant, anticancer and antimicrobial properties (Table 3).

Conclusions

According to Overy *et al.*³³ publishing lists of marine-derived fungi based on location is only sufficient when no relationship between the location and the fungus is known. In the case of macroalgal endophytes this is almost always the case, where fungi are isolated from the inside of marine algae and the relationship between host and fungus is not known. What has developed over the recent past are either lists of endophytes from a variable list of macroalgal hosts^{9,11,12} or bioassay data and chemical structures from extracts of endophytic fungi isolated from one or only a few algal hosts (Tables 2 and 3). This is not to imply that these approaches are somehow limited in impact or importance, but rather to state there is much to do in the study of endophytic fungi from macroalgae. This review attempts to summarize the current information on these two important facets of endophyte research from marine algae. Globally, endophytes have been isolated from less than 1% of the known macroalgal species resulting in a dearth of information on the abundance, distribution and species richness of endophytes from this source. Additionally, the effect of anthropomorphic influences and the nature of the relationship between macroalgae and their associated endophytes present important opportunities for research. These factors, and others, may impact a variety of endophytes and their capacity to produce novel chemical entities.

Bioassay data supports the notion that endophytes from marine algae are an important source of bioactive compounds (Tables 2 and 3), not only in terms of new natural products but also in relation to known compounds from different sources. As noted in the lack of data for distribution of endophytes, there is also a lot of work to be done in the bioassay and structure elucidation of compounds produced by algal endophytes. As a relatively new and understudied source of natural products, algal endophytes may open up a substantial vista of new chemistry. This review collates and organizes the latest literature with the aim to aid future research in the field. Clearly, there is much to do and much to discover.

1. Tan, R. X. and Zou, W. X., Endophytes: a rich source of functional metabolites. *Nat. Prod. Rep.*, 2001, **18**, 448–459.
2. Strobel, G. A. and Bryn, D., Bioprospecting for microbial endophytes and their natural products. *Microbiol. Mol. Biol. Rev.*, 2003, **67**, 491–502.
3. Zhang, H. W., Song, Y. C. and Tan, R. X., Biology and chemistry of endophytes. *Nat. Prod. Rep.*, 2006, **23**, 753–771.
4. Debbab, A., Aly, A. H. and Proksch, P., Bioactive secondary metabolites from endophytes and associated marine derived fungi. *Fungal Divers.*, 2011, **49**, 1–12.
5. Debbab, A., Aly, A. H. and Proksch, P., Endophytes and associated marine derived fungi – ecological and chemical perspectives. *Fungal Divers.*, 2012, **57**, 45–83.
6. Radic, N. and Strukelj, B., Endophytic fungi – the treasure chest of antibacterial substances. *Phytomedicine*, 2012, **19**, 1270–1284.

7. Oliveira, A. L. L. d., Felício, R. D. and Deboni, H. M., Marine natural products: chemical and biological potential of seaweeds and their endophytic fungi. *Rev. Bras. Farmacogn.*, 2012, **22**, 906–920.
8. Zhang, Y. *et al.*, Broad-spectrum antimicrobial epiphytic and endophytic fungi from marine organisms: isolation, bioassay and taxonomy. *Mar. Drugs*, 2009, **7**, 97–112.
9. Suryanarayanan, T. S., Venkatachalam, A., Thirunavukkarasu, N., Ravishankar, J. P., Doble, M. and Geetha, V., Internal mycobiota of marine macroalgae from the Tamil Nadu coast: distribution, diversity and biotechnological potential. *Bot. Mar.*, 2010, **53**, 457–468.
10. Ariffin, S. A., Davis, P. and Ramasamy, K., Cytotoxic and antimicrobial activities of Malaysian marine endophytic fungi. *Bot. Mar.*, 2011, **54**, 95–100.
11. Flewelling, A., Ellsworth, K., Sanford, J., Forward, E., Johnson, J. and Gray, C., Macroalgal endophytes from the Atlantic coast of Canada: a potential source of antibiotic natural products? *Microorganisms*, 2013, **1**, 175–187.
12. Flewelling, A. J., Johnson, J. A. and Gray, C. A., Isolation and bioassay screening of fungal endophytes from North Atlantic marine macroalgae. *Bot. Mar.*, 2013, **56**, 287–297.
13. Abdel-Lateff, A., Fisch, K. M., Wright, A. D. and König, G. M., A new antioxidant isobenzofuranone derivative from the algicolous marine fungus *Epicoccum* sp. *Planta Med.*, 2003, **69**, 831–834.
14. Abdel-Lateff, A., Klemke, C., König, G. M. and Wright, A. D., Two new xanthone derivatives from the algicolous marine fungus *Wardomyces anomalus*. *J. Nat. Prod.*, 2003, **66**, 706–708.
15. Abdel-Lateff, A., König, G. M., Fisch, K. M., Holler, U., Jones, P. G. and Wright, A. D., New antioxidant hydroquinone derivatives from the algicolous marine fungus *Acremonium* sp. *J. Nat. Prod.*, 2002, **65**, 1605–1611.
16. Zhang, Y., Li, X. M., Proksch, P. and Wang, B. G., Ergosterimide, a new natural Diels–Alder adduct of a steroid and maleimide in the fungus *Aspergillus niger*. *Steroids*, 2007, **72**, 723–727.
17. Zhang, Y., Li, X. M. and Wang, B. G., Nigerasperones A–C, new monomeric and dimeric naphtho- γ -pyrones from a marine alga-derived endophytic fungus *Aspergillus niger* EN-13. *J. Antibiot.*, 2007, **60**, 204–210.
18. Zhang, Y., Li, X. M., Wang, C. Y. and Wang, B. G., A new naphthoquinoneimine derivative from the marine algal-derived endophytic fungus *Aspergillus niger* EN-13. *Chin. Chem. Lett.*, 2007, **18**, 951–953.
19. Zhang, Y., Wang, S., Li, X. M., Cui, C. M., Feng, C. and Wang, B. G., New sphingolipids with a previously unreported 9-methyl-c20-sphingosine moiety from a marine algous endophytic fungus *Aspergillus niger* EN-13. *Lipids*, 2007, **42**, 759–764.
20. Elsebai, M. F. *et al.*, Antimicrobial phenalenone derivatives from the marine-derived fungus *Coniothyrium cereale*. *Org. Biomol. Chem.*, 2011, **9**, 802–808.
21. Elsebai, M. F. *et al.*, HLE-inhibitory alkaloids with a polyketide skeleton from the marine-derived fungus *Coniothyrium cereale*. *J. Nat. Prod.*, 2011, **74**, 2282–2285.
22. Guiry, M. D., How many species of algae are there? *J. Phycol.*, 2012, **48**, 1057–1063.
23. Schulz, B. *et al.*, Screening strategies for obtaining novel, biologically active, fungal secondary metabolites from marine habitats. *Bot. Mar.*, 2008, **51**, 219–234.
24. Harvey, J. B. J. and Goff, L. J., Genetic covariation of the marine fungal symbiont *Haloguignardia irritans* (Ascomycota, Pezizomycotina) with its algal hosts *Cystoseira* and *Halidryis* (Phaeophyceae, Fucales) along the west coast of north america. *Fungal Biol.*, 2010, **114**, 82–95.
25. Krick, A. *et al.*, Potential cancer chemopreventive *in vitro* activities of monomeric xanthone derivatives from the marine algicolous fungus *Monodictys putredinis*. *J. Nat. Prod.*, 2007, **70**, 353–360.
26. Wu, B., Wu, X., Sun, M. and Li, M., Two novel tyrosinase inhibitory sesquiterpenes induced by CuCl₂ from a marine-derived fungus *Pestalotiopsis* sp. Z233. *Mar. Drugs*, 2013, **11**, 2713–2721.
27. Cui, C. M., Li, X. M., Meng, L., Li, C. S., Huang, C. G. and Wang, B. G., 7-Nor-ergosterolide, a pentalactone-containing norsteroid and related steroids from the marine-derived endophytic *Aspergillus ochraceus* EN-31. *J. Nat. Prod.*, 2010, **73**, 1780–1784.
28. Cui, C.-M., Li, X.-M., Li, C.-S., Sun, H.-F., Gan, S.-S. and Wang, B.-G., Benzodiazepine alkaloids from marine-derived endophytic fungus *Aspergillus ochraceus*. *Helv. Chim. Acta*, 2009, **92**, 1366–1370.
29. Qiao, M. F., Ji, N. Y., Liu, X. H., Li, F. and Xue, Q. Z., Asporergosterol, a new steroid from an algicolous isolate of *Aspergillus oryzae*. *Nat. Prod. Commun.*, 2010, **5**, 1575–1578.
30. Qiao, M. F., Ji, N. Y., Liu, X. H., Li, K., Zhu, Q. M. and Xue, Q. Z., Indoloditerpenes from an algicolous isolate of *Aspergillus oryzae*. *Bioorg. Med. Chem. Lett.*, 2010, **20**, 5677–5680.
31. Qiao, M. F., Ji, N. Y., Miao, F. P. and Yin, X. L., Steroids and an oxylipin from an algicolous isolate of *Aspergillus flavus*. *Magn. Reson. Chem.*, 2011, **49**, 366–369.
32. Abdel-Lateff, A., Chaetominedione, a new tyrosine kinase inhibitor isolated from the algicolous marine fungus *Chaetomium* sp. *Tetrahedron Lett.*, 2008, **49**, 6398–6400.
33. Overy, D. P., Bayman, P., Kerr, R. G. and Bills, G. F., An assessment of natural product discovery from marine (*sensu strictu*) and marine-derived fungi. *Mycology*, 2014, **5**, 145–167.
34. An, C.-Y., Li, X.-M., Li, C.-S., Gao, S.-S., Shang, Z. and Wang, B.-G., Triazoles and other *n*-containing metabolites from the marine-derived endophytic fungus *Penicillium chrysogenum* EN-118. *Helv. Chim. Acta*, 2013, **96**, 682–687.
35. Cui, C. M., Li, X. M., Li, C. S., Proksch, P. and Wang, B. G., Cytoglobosins A–G, cytochalasans from a marine-derived endophytic fungus, *Chaetomium globosum* QEN-14. *J. Nat. Prod.*, 2010, **73**, 729–733.
36. de Silva, E. D., Geiermann, A.-S., Mitova, M. I., Kuegler, P., Blunt, J. W., Cole, A. L. J. and Munro, M. H. G., Isolation of 2-pyridone alkaloids from a New Zealand marine-derived *Penicillium* species. *J. Nat. Prod.*, 2009, **72**, 477–479.
37. Deckert, R. and Garbary, D., *Ascophyllum* and its symbionts. VI. Microscopic characterization of the *Ascophyllum nodosum* (Phaeophyceae), *Mycophycias ascophylli* (Ascomycetes) symbiotum. *Algae*, 2005, **20**, 225–232.
38. Deckert, R. J. and Garbary, D. J., *Ascophyllum* and its symbionts. VIII. Interactions among *Ascophyllum nodosum* (Phaeophyceae), *Mycophycias ascophylli* (Ascomycetes) and *Elachista fucicola* (Phaeophyceae). *Algae*, 2005, **20**, 363–368.
39. Du, F. Y., Li, X. M., Li, C. S., Shang, Z. and Wang, B. G., Cristatamins a-d, new indole alkaloids from the marine-derived endophytic fungus *Eurotium cristatum* EN-220. *Bioorg. Med. Chem. Lett.*, 2012, **22**, 4650–4653.
40. Elsebai, M. F., Kehraus, S., Gutschow, M. and König, G. M., New polyketides from the marine-derived fungus *Phaeosphaeria spartinae*. *Nat. Prod. Commun.*, 2009, **4**, 1463–1468.
41. Elsebai, M. F., Kehraus, S., Gutschow, M. and König, G. M., Spartinoxide, a new enantiomer of aZ2775c with inhibitory activity toward HLE from the marine-derived fungus *Phaeosphaeria spartinae*. *Nat. Prod. Commun.*, 2010, **5**, 1071–1076.
42. Elsebai, M. F., Kehraus, S. and König, G. M., Caught between triterpene- and steroid-metabolism: 4- α -carboxylic pregnane-derivative from the marine alga-derived fungus *Phaeosphaeria spartinae*. *Steroids*, 2013, **78**, 880–883.
43. Erbert, C. *et al.*, Antibacterial compound from the endophytic fungus *Phomopsis longicolla* isolated from the tropical red seaweed *Bostrychia radicans*. *Bot. Mar.*, 2012, **55**, 435–440.

44. Flewelling, A. J., Johnson, J. A. and Gray, C. A., Antimicrobials from the marine algal endophyte *Penicillium* sp. *Nat. Prod. Commun.*, 2013, **8**, 373–374.
45. Fries, N., Physiological characteristics of *Mycosphaerella ascophyllii*, a fungal endophyte of the marine brown alga *Ascophyllum nodosum*. *Physiol. Plant*, 1979, **45**, 117–121.
46. Fries, N. and Thorentolling, K., Identity of fungal endophyte of *Ascophyllum* with *Mycosphaerella ascophyllii* established by means of fluorescent antibody technique. *Bot. Mar.*, 1978, **21**, 409–411.
47. Gamal-Eldeen, A. M., Abdel-Lateff, A. and Okino, T., Modulation of carcinogen metabolizing enzymes by chromanone a; a new chromone derivative from algaliculous marine fungus *Penicillium* sp. *Environ. Toxicol. Pharmacol.*, 2009, **28**, 317–322.
48. Gao, S.-S., Li, X.-M. and Wang, B.-G., Perylene derivatives produced by *Alternaria alternata*, an endophytic fungus isolated from *Laurencia* species. *Nat. Prod. Commun.*, 2009, **4**, 1477–1480.
49. Gao, S. S., Li, X. M., Du, F. Y., Li, C. S., Proksch, P. and Wang, B. G., Secondary metabolites from a marine-derived endophytic fungus *Penicillium chrysogenum* QEN-24S. *Mar. Drugs*, 2011, **9**, 59–70.
50. Gao, S. S., Li, X. M., Li, C. S., Proksch, P. and Wang, B. G., Penicesteroids A and B, antifungal and cytotoxic polyoxygenated steroids from the marine alga-derived endophytic fungus *Penicillium chrysogenum* QEN-24S. *Bioorg. Med. Chem. Lett.*, 2011, **21**, 2894–2897.
51. Gao, S.-S., Li, X.-M., Zhang, Y., Li, C.-S. and Wang, B.-G., Conidiogenones H and I, two new diterpenes of cyclopiane class from a marine-derived endophytic fungus *Penicillium chrysogenum* QEN-24S. *Chem. Biodivers.*, 2011, **8**, 1748–1753.
52. Garbary, D. J. and Gautam, A., The *Ascophyllum*, *Polysiphonia*, *Mycosphaerella* symbiosis. 1. Population ecology of *Mycosphaerella* from Nova Scotia. *Bot. Mar.*, 1989, **32**, 181–186.
53. Garbary, D. J., Burke, J. and Tian, L. N., The *Ascophyllum* *Polysiphonia*, *Mycosphaerella* symbiosis. 2. Aspects of the ecology and distribution of *Polysiphonia lanosa* in Nova Scotia. *Bot. Mar.*, 1991, **34**, 391–401.
54. Garbary, D. J. and Macdonald, K. A., The *Ascophyllum*, *Polysiphonia*, *Mycosphaerella* symbiosis. 4. Mutualism in the *Ascophyllum* *Mycosphaerella* interaction. *Bot. Mar.*, 1995, **38**, 221–225.
55. Garbary, D. J. and London, J. F., The *Ascophyllum*, *Polysiphonia* *Mycosphaerella* symbiosis. 5. Fungal infection protects *A. nodosum* from desiccation. *Bot. Mar.*, 1995, **38**, 529–533.
56. Garbary, D. J., Deckert, R. J. and Hubbard, C. B., *Ascophyllum* and its symbionts. VII. Three-way interactions among *Ascophyllum nodosum* (Phaeophyceae), *Mycophycias ascophylli* (Ascomycetes) and *Vertebrata lanosa* (Rhodophyta). *Algae*, 2005, **20**, 353–361.
57. Haroon, M. H., Premaratne, S. R., Choudhry, M. I. and Dharmaratne, H. R., A new beta-glucuronidase inhibiting butyrolactone from the marine endophytic fungus *Aspergillus terreus*. *Nat. Prod. Res.*, 2013, **27**, 1060–1066.
58. Klemke, C., Kehraus, S., Wright, A. D. and König, G. M., New secondary metabolites from the marine endophytic fungus *Apiospora montagnei*. *J. Nat. Prod.*, 2004, **67**, 1058–1063.
59. Kohlmeyer, J. and Hawkes, M. W., A suspected case of mycophycobiosis between *Mycosphaerella apophlaeae* (Ascomycetes) and *Apophlaea* spp. (Rhodophyta). *J. Phycol.*, 1983, **19**, 257–260.
60. Kralj, A. *et al.*, Arugosins g and h: prenylated polyketides from the marine-derived fungus *Emericella nidulans* var. *Acristata*. *J. Nat. Prod.*, 2006, **69**, 995–1000.
61. Krohn, K., Dai, J., Florke, U., Aust, H. J., Dräger, S. and Schulz, B., Botryane metabolites from the fungus *Geniculosporium* sp. isolated from the marine red alga *Polysiphonia*. *J. Nat. Prod.*, 2005, **68**, 400–405.
62. Li, F., Li, K., Li, X. and Wang, B., Chemical constituents of marine algal-derived endophytic fungus *Exophiala oligosperma* EN-21. *Chin. J. Oceanol. Limnol.*, 2011, **29**, 63–67.
63. Li, X., Li, X.-M., Xu, G.-M., Li, C.-S. and Wang, B.-G., Antioxidant metabolites from marine alga-derived fungus *Aspergillus wentii* EN-48. *Phytochem. Lett.*, 2014, **7**, 120–123.
64. Li, X. D., Miao, F. P., Liang, X. R. and Ji, N. Y., Meroterpenes from an algaliculous strain of *Penicillium echinulatum*. *Magn. Reson. Chem.*, 2014, **52**, 247–250.
65. Liu, X.-H., Tang, X.-Z., Miao, F.-P. and Ji, N.-Y., A new pyrrolidine derivative and steroids from an algaliculous *Gibberella zeae* strain. *Nat. Prod. Commun.*, 2011, **6**, 1243–1246.
66. Liu, X.-H., Miao, F.-P., Li, X.-D., Yin, X.-L. and Ji, N.-Y., A new sesquiterpene from an endophytic *Aspergillus versicolor* strain. *Nat. Prod. Commun.*, 2012, **7**, 819–820.
67. Lösgen, S., Schlörke, O., Meindl, K., Herbst-Irmer, R. and Zeeck, A., Structure and biosynthesis of chaetocyclinones, new polyketides produced by an endosymbiotic fungus. *Eur. J. Org. Chem.*, 2007, **2007**, 2191–2196.
68. Miao, F. P., Li, X. D., Liu, X. H., Cichewicz, R. H. and Ji, N. Y., Secondary metabolites from an algaliculous *Aspergillus versicolor* strain. *Mar. Drugs*, 2012, **10**, 131–139.
69. Miao, F.-P., Liang, X.-R., Yin, X.-L., Wang, G. and Ji, N.-Y., Absolute configurations of unique harziane diterpenes from *Trichoderma* species. *Org. Lett.*, 2012, **14**, 3815–3817.
70. Miao, F. P., Liang, X. R., Liu, X. H. and Ji, N. Y., Aspewentins A–C, norditerpenes from a cryptic pathway in an algaliculous strain of *Aspergillus wentii*. *J. Nat. Prod.*, 2014, **77**, 429–432.
71. Nielsen, J. E., Yu, S. K., Bojko, M. and Marcussen, J., Alpha-1,4-glucan lyase-producing endophyte of *Gracilariopsis* sp (Rhodophyta) from China. *Eur. J. Phycol.*, 2000, **35**, 207–212.
72. Osterhage, C., Kaminsky, R., König, G. M. and Wright, A. D., Ascosalipyrrolidinone a, an antimicrobial alkaloid, from the obligate marine fungus *Ascochyta salicorniae*. *J. Org. Chem.*, 2000, **65**, 6412–6417.
73. Osterhage, C., König, G. M., Holler, U. and Wright, A. D., Rare sesquiterpenes from the algaliculous fungus *Drechslera dematioidea*. *J. Nat. Prod.*, 2002, **65**, 306–313.
74. Perez-Ortega, S., de los Rios, A., Crespo, A. and Sancho, L. G., Symbiotic lifestyle and phylogenetic relationships of the bionts of *Mastodia tessellata* (Ascomycota, *incertae sedis*). *Am. J. Bot.*, 2010, **97**, 738–752.
75. Pontius, A., Mohamed, I., Krick, A., Kehraus, S. and König, G. M., Aromatic polyketides from marine algaliculous fungi. *J. Nat. Prod.*, 2008, **71**, 272–274.
76. Pontius, A. *et al.*, Noduliprevenone: a novel heterodimeric chromanone with cancer chemopreventive potential. *Chemistry*, 2008, **14**, 9860–9863.
77. Pontius, A., Krick, A., Kehraus, S., Brun, R. and König, G. M., Antiprotozoal activities of heterocyclic-substituted xanthenes from the marine-derived fungus *Chaetomium* sp. *J. Nat. Prod.*, 2008, **71**, 1579–1584.
78. Seibert, S. F. *et al.*, Polyketides from the marine-derived fungus *Ascochyta salicorniae* and their potential to inhibit protein phosphatases. *Org. Biomol. Chem.*, 2006, **4**, 2233–2240.
79. Stanley, S. J., Observations on the seasonal occurrence of marine endophytic and parasitic fungi. *Can. J. Bot.*, 1992, **70**, 2089–2096.
80. Sun, H. F. *et al.*, Asperolides A–C, tetranorlabdane diterpenoids from the marine alga-derived endophytic fungus *Aspergillus wentii* EN-48. *J. Nat. Prod.*, 2012, **75**, 148–152.
81. Sun, R. R., Miao, F. P., Zhang, J., Wang, G., Yin, X. L. and Ji, N. Y., Three new xanthone derivatives from an algaliculous isolate of *Aspergillus wentii*. *Magn. Reson. Chem.*, 2013, **51**, 65–68.
82. Sun, H.-F., Li, X.-M., Meng, L.-H., Cui, C.-M., Gao, S.-S., Li, C.-S. and Wang, B.-G., Two new secoanthraquinone derivatives from the marine-derived endophytic fungus *Aspergillus wentii* EN-48. *Helv. Chim. Acta*, 2013, **96**, 458–462.

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83. Tarman, K. *et al.*, Helicascolide c, a new lactone from an Indonesian marine algal strain of *Daldinia eschscholzii* (Xylariaceae, Ascomycota). *Phytochem. Lett.*, 2012, **5**, 83–86.
 84. Teuscher, F., Lin, W., Wray, V., Edrada, R., Padmakumar, K., Proksch, P. and Ebel, R., Two new cyclopentanoids from the endophytic fungus *Aspergillus sydowii* associated with the marine alga *Acanthophora spicifera*. *Nat. Prod. Commun.*, 2006, **1**, 927–933.
 85. Wang, F.-W., Bioactive metabolites from *Guignardia* sp., an endophytic fungus residing in *Undaria pinnatifida*. *Chin. J. Nat. Med.*, 2012, **10**, 72–76.
 86. Wang, S. *et al.*, Chaetopyranin, a benzaldehyde derivative, and other related metabolites from *Chaetomium globosum*, an endophytic fungus derived from the marine red alga *Polysiphonia urceolata*. *J. Nat. Prod.*, 2006, **69**, 1622–1625.
 87. Xu, H., Deckert, R. J. and Garbary, D. J., *Ascophyllum* and its symbionts. X. Ultrastructure of the interaction between *A. nodosum* (Phaeophyceae) and *Mycophycias ascophylli* (Ascomycetes). *Botany*, 2008, **86**, 185–193.
 88. Yang, R. Y., Li, C. Y., Lin, Y. C., Peng, G. T., She, Z. G. and Zhou, S. N., Lactones from a brown alga endophytic fungus (No. ZZ36) from the South China Sea and their antimicrobial activities. *Bioorg. Med. Chem. Lett.*, 2006, **16**, 4205–4208.
 89. Zhang, Y., Li, X.-M., Feng, Y. and Wang, B.-G., Phenethyl-alpha-pyrone derivatives and cyclodipeptides from a marine algal endophytic fungus *Aspergillus niger* EN-13. *Nat. Prod. Res.*, 2010, **24**, 1036–1043.
 90. Zhang, Z. *et al.*, Wentilactone B induces G2/M phase arrest and apoptosis via the Ras/Raf/MAPK signaling pathway in human hepatoma SMMC-7721 cells. *Cell Death Dis.*, 2013, **4**, e657.
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