Journal of Pharmaceutical Research Vol. 12, No. 4, October - December 2013: 122-127.

VALIDATED STABILITY INDICATING RP-HPLC METHOD FOR SIMULTANEOUS DETERMINATION OF METOPROLOL SUCCINATE AND OLMESARTAN MEDOXOMIL IN TABLET DOSAGE FORM

Mahaparale S P*1, Gonjari I D2, Jayaveera K N3

¹Padm. Dr. D. Y. Patil College of Pharmacy, Sector No. 29, Pradhikaran, Near ZSI Building, Akurdi, Pune - 411 044, Maharashtra, India.

²Government College of Pharmacy, Vidyanagar, Karad - 415 124, Maharashtra, India.

³ Jawaharlal Nehru Technological University, Anantapur - 515 002, Andhra Pradesh, India.

Received on: 02.08.2013 Revised: 07.10.2013 Accepted: 08.10.2013

ABSTRACT

The present paper describes simple, rapid, reproducible, accurate and precise stability indicating HPLC method developed for quantitative simultaneous estimation of metoprolol succinate and olmesartan medoxomil in bulk and combined pharmaceutical dosage form. A chromatographic separation of both drugs was achieved with Chromasil 250 x 4.6 mm, i.d 5 im C-18 column using methanol:0.05% v/v O-phosphoric acid in water (50:50 v/v) at the flow rate of 1ml/min. The measurements were made at 228.0 nm as detector wavelength. The described method showed excellent linearity over a range of 5-80 ig/ml for metoprolol succinate and 5-70 ig/ml for olmesartan medoxomil. The coefficient of correlation for metoprolol succinate and olmesartan medoxomil was found to be 0.9990 and 0.9993 respectively. The retention time for metoprolol succinate and olmesartan medoxomil was found to be 3.485 min and 7.085 min, respectively. The tailing factor for metoprolol succinate and olmesartan medoxomil was found to be 1.02 and 1.13 respectively. Both drugs and their combination drug product were found to be stable in neutral, thermal, oxidative and photolytic stress conditions but mild degradation was observed in acidic and alkaline conditions.

Keywords: *Metoprolol succinate; Olmesartan medoxomil; HPLC method validation; Stability-indicating method; ICH guidelines.*

INTRODUCTION

Chemically, Metoprolol succinate (MET) (Fig.1) is (±)-1-(Isopropylamino-3-[4-(2-methoxyethyl)phenoxy] propan-2-ol.1 It is used as an antianginal and antihypertensive.1 The official methods of assay like Potentiometric² and HPLC³ are reported for MET. Chemically, Olmesartan medoxomil (OLM) (Fig. 1) is 4-(1-Hydroxy-1-methylethyl)-2-propyl-1-[[22 -(1Htetrazol-5-yl)[1,12 -biphenyl]-4-yl]methyl]-1H-imidazole-5-carboxylic acid (5-methyl -2-oxo-1,-3-dioxol-4yl)methyl ester4. It is selective angiotensin II receptor antagonist and used as an antihypertensive.1 OLM lowers blood pressure and increases the supply of blood and oxygen to the heart.1 OLM and MET is a combination of medicines used to treat high blood pressure (hypertension). Literature survey revealed that various methods like UV spectrophotometric⁵, HPLC⁶⁻⁹ and GC-MS¹⁰ for the estimation of metoprolol succinate as single and HPLC in combination with other antihypertensive agents¹¹⁻¹³ are reported. The methods such as UV spectrophotometric^{14,15}, estimation of OLM in plasma, urine and tablet by HPLC 15-19 and LC-MS-MS 20 detection for olmesartan medoxomil as single and HPLC method in combination with other drugs are

Metoprolol succinate (MET)

Olmesartan Medoxomil (OLM)

Fig. 1: Chemical structure of MET and OLM

*Correspondence: sonalimahaparale@gmail.com Mob + 91 98506 53148

Metoprolol and Olmesartan determination

reported. ^{21,22} The purpose of stability testing is to provide evidence on how the quality of a drug substance or drug product varies with time under the influence of a variety of environmental factors such as temperature, humidity and light and enables recommendation of storage conditions, retest periods and shelf-life to be established. There is no report yet of stability indicating RP-HPLC method for these drugs in combination. The aim of the present study accordingly was to establish inherent stability of metoprolol succinate and olmesartan medoxomil in combined tablet dosage form through stress studies under ICH²³ recommended test conditions and to develop and validate stability indicating HPLC method.

EXPERIMENTAL

Chemicals and reagents

Reference standards of metoprolol succinate and olmesartan medoxomil were procured from Lupin Research Park, Pune and Emcure Pharmaceuticals Ltd., Pune respectively. Methanol (HPLC grade) was obtained from Qualigen Laboratories Pvt. Ltd., Mumbai. Analytical grade of hydrochloric acid (HCl), sodium hydroxide (NaOH), o-phosphoric acid and hydrogen peroxide were obtained from Merck Ltd., Mumbai. The tablets containing metoproplol succinate (25 mg) and olmesartan medoxomil (20 mg) were procured from local market.

Instrument

The chromatographic system used was an Agilent 1120 series, which comprised a degasser, gradient pump and photodiode array detector. The system was controlled through Ezchrome software using Chromasil C18 (4.6 x 250mm, 5 im) column maintained at 25°C temperature.

Chromatographic conditions

The separation was achieved using a mobile phase consisting methanol:0.05% v/v o-phosphoric acid in water (50:50 v/v) at a flow rate of 1.0 ml/min and the eluent was monitored using PDA detector at 228.0 nm. The mobile phase was kept in ultrasonicator for 30 min and filtered through a 0.45-ìm nylon membrane filter. The column was maintained at 25°C temperature and injection volume of 20 ìl was used. The peak homogeneity was expressed in terms of peak purity and was obtained directly from software.

Standard stock solutions

The stock solution (100 \(\text{ig/ml}\)) of MET and OLM were prepared separately by dissolving accurately 10 mg of each drug in 100 ml methanol HPLC grade in 100 ml volumetric flask.

Calibration curve

Appropriate aliquots of standard stock solutions of MET and OLM were diluted with mobile phase to obtain concentrations in the range of 5, 10, 20, 30, 40, 50, 60, 70 and 80 ig/ml of MET and 5, 10, 20, 30, 40, 50,

MAHAPARALE S P. GONJARI I D and JAYAVEERA K N

60 and 70 ig/ml of OLM respectively. The linearity of MET (Fig. 2) and OLM (Fig. 3) was found to be in the concentration ranges of 5-80 ig/ml and 5-70 ig/ml, respectively, at their respective maxima. The coefficients of correlation were found to be 0.9990 for MET and 0.9993 for OLM. The mixed standard solution containing 50 ig/ml of MET and 40 ig/ml of OLM was prepared from each standard stock solution and injected into HPLC system.

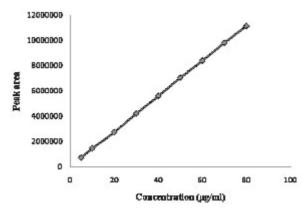


Fig. 2: Calibration curve of Metoprolol succinate

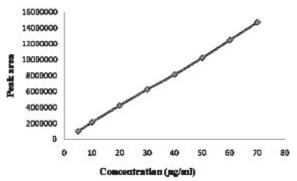


Fig. 3: Calibration curve of Olmesartan medoxomil

RESULTS

Optimization of the chromatographic conditions:

To develop a stability-indicating method, different stationary phases like C18, CN, different mobile phases containing buffers like phosphate, ammonium acetate, with different pH (3-7), and organic modifier (acetonitrile) were used. Our objective of chromatographic method development was to achieve peak tailing factor < 2, retention time between 3 to 10 min, along with resolution between MET and OLM > 2. The chromatographic separation was achieved using Chromasil C18 (250 x 4.6 mm i.d., 5 ìm) column, changing the composition of mobile phase and optimized the chromatographic method. To develop a stability-indicating method assessing the effect of change of proportion, MET and OLM were well-resolved from degradation products using mobile phase composition of methanol: 0.05% v/v o-phosphoric acid in water (50:50 v/v) at a flow rate of 1.0 ml/min. with UV detection at 228.0 nm wavelength and injection

Metoprolol and Olmesartan determination

volume 20 ì I. It was found to ideally resolve the peaks of MET (Rt 3.485 min) and OLM (Rt 7.085 min) (Fig.4). Resolution (Rs) between MET and OLM was found to be 10.28. ICH guidelines recommend 10 20 % degradation for establishing stability indicating nature of the assay method.

Resolution (Rs) = t_2 - t_1 / w_1 + w_2 t_1 , t_2 are retention time of MET and OLM respectively, w_1 , w_2 are width of peaks for MET and OLM respectively, Rs = 7.085 -3.485 / 0.15+ 0.2

Rs = 3.6 / 0.35 = 10.28

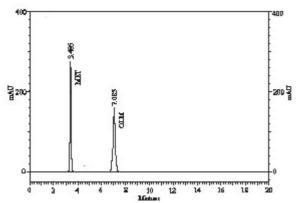


Fig. 4: Chromatogram of MET and OLM in standard mixture

Analysis of tablet formulation:

Twenty tablets of Rasotan Beta 25 (Emcure Pharma) each containing 25 mg of metoprolol succinate and 20 mg of olmesartan medoxomil were weighed and crushed in glass mortar to obtain fine powder. The powder sample equivalent to 25 mg of MET and 20 mg of OLM was transferred into a 100 ml volumetric flask and dissolved in 50 ml methanol HPLC grade. The flask was kept in an ultrasonic bath for 20 min. The volume was adjusted to 100 ml with methanol HPLC grade. The solution was filtered through 0.2 i nylon membrane filter. From this stock solution, 2 ml solution was pipetted out and transferred to 10 ml volumetric flask and made volume up to the mark with mobile phase to get the concentration 50 ig/ml of MET and 40 ig/ml of OLM. The solution was injected into HPLC system (Fig. 5). The results of the assay of tablet formulation and its statistical validation data is given in Table 1.

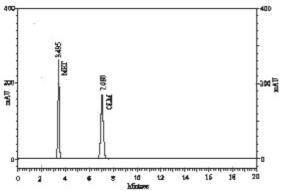


Fig. 5: Chromatogram of MET and OLM in Tablet mixture

MAHAPARALE S P. GONJARI I D and JAYAVEERA K N

Table 1: Analysis of Tablet Formulation

Tablet cample	Label olalm (mg/tablet)	Amount found (mg/tablet)	% Label olalm found*	± Standard deviation	Standard error
MET	25	24.96	99.84	0.4134	0.1688
O LM	20	19.96	99.80	0.5030	0.2053

*Average of six readings

MET and OLM denotes metoprolol succinate and olmesartan medoxomil respectively

Procedure for forced degradation study of drug substances

Forced degradation of each drug substances and the drug product was carried out under acid, base, neutral, oxidative, thermolytic and photolytic stress conditions.

Acidic degradation

The accurate quantity of 2.5 mg MET and 2.5 mg OLM was weighed and transferred to 25 ml volumetric flask separately; 10 ml of 0.1 N HCl was added into each flask separately and the flask was kept for 2 hrs at room temperature. Then it was neutralized with 0.1 N NaOH and solution was sonicated for 30 min with intermittent shaking in ultrasonicator. Then the volume was made up with methanol HPLC grade and each solution was filtered through 0.2 μm membrane filter. From the filtered stock solutions, 5 ml of MET and 4 ml of OLM was pipetted out separately and transferred into a 10 ml volumetric flask and diluted to volume with mobile phase to obtain final concentration of 50 $\mu g/ml$ of MET and 40 $\mu g/ml$ OLM. Then the solution was injected into system (Fig. 6).

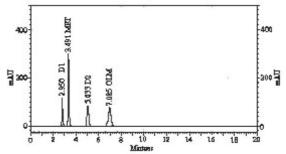


Fig. 6: Acidic degradation of MET and OLM in standard mixture

Alkali degradation

Alkali degradation was carried out by adding 10 ml of 1 N NaOH and the mixture was refluxed for 30 min at 60°C. Then it was neutralized with 1 N HCl and the solution was sonicated for 30 min with intermittent shaking in ultrasonicator (Fig. 7).

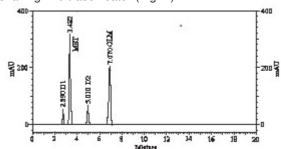


Fig. 7: Alkali degradation of MET and OLM in standard mixture

Metoprolol and Olmesartan determination Neutral degradation

10.0 ml distilled water was added to each stock solution of MET and OLM separately. Both solutions were refluxed at 60°C for 2 hrs and cooled at room temperature (Fig. 8).

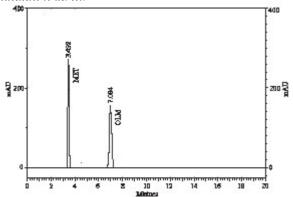


Fig. 8: Neutral degradation of MET and OLM in standard mixture

Oxidative degradation

Oxidative stress degradation of MET and OLM was conducted with 30% $\rm H_2O_2$ for 2 hrs at 60°C in a water bath (Fig. 9).

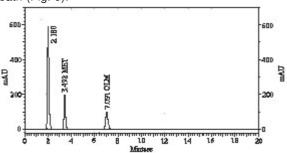


Fig. 9: Oxidative degradation of MET and OLM in standard mixture

Thermal degradation

About 100 mg of drug substances were placed in a controlled temperature oven at 80 °C for 48 hrs. (Fig.10)

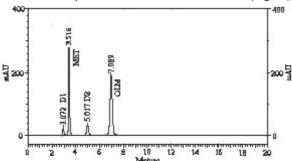


Fig. 10: Thermal degradation of MET and OLM in standard mixture

Photodegradation (UV light)

Photodegradation was performed by spreading the drug substance in petri dish as thin film and kept in

MAHAPARALE S P. GONJARI I D and JAYAVEERA K N

photostability chamber equipped with ultraviolet light with energy of not less than 200 watt hours/square meter (Fig. 11).

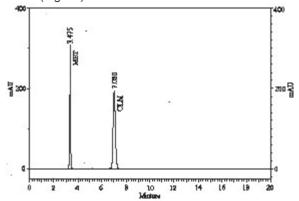


Fig. 11: Photolytic degradation under UV light of MET and OLM in standard mixture

Photodegradation (fluroscence light)

Photodegradation was performed by exposing the drug substance in photostability chamber equipped with fluroscence light illumination not less than 1.2 million lux hours. Sample was weighed, dissolved and diluted to obtain final concentration and injected into system. (Fig. 12)

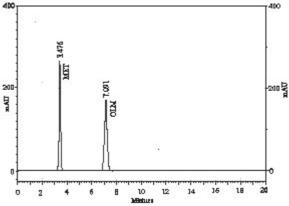


Fig. 12: Photolytic degradation under fluorescence light of MET and OLM in standard mixture

The results of forced degradation study of standard the proposed method are shown in Table 2.

Table 2: Results of forced degradation study

Stress condition	RIG T			OUM		
	Degradation (%)	Peakpurity		Degradation	Peak purity	
		Front	Tail	(%)	Front	Tail
Addict 0.1 NHC I/2 hr	19.76	994	991	19.53	99.5	9.92
Alkaline (1 N NaOH) příka 50/C 2hr	7.12	995	992	9.25	99.5	9.91
Neutrality Orie Bus 60°C 2 hr	0.50	997	990	0.15	99.6	9.92
Ouldraine 00 % HaCa (2hr	9.50	994	992	9.20	99.5	9.90
Dry head 60 °C (Tidaya.	3.00	992	990	3.00	.99.0	9.92
Photolysis UV 20 0 was thours aquaire motor	0.10	996	990	0.30	99.5	9.92
Fluorescence (1.2 million Lue.hrs	90.0	994	992	0.10	99.7	9.90

MET and OLM denotes metoprolol succinate and olmesartan medoxomil respectively.

Metoprolol and Olmesartan determination Method Validation

Linearity

Linearity for MET and OLM was selected at 5-80 ig/ml and 5-70 ig/ml. The correlation coefficients were selected at 0.9990 and 0.9993 for MET and OLM, respectively. The results are shown in Table 3.

Table 3: System Suitability Parameters

Para meter	MET	OLM
Line arity range* (µg/ml)	5-80	5-70
Correlation coefficient*	0.9990	0.9993
Slope*	139 49	20783
Limit of dietection (µg/ml)	0.020	0.024
Limit of quantitation (µg/ml)	0.080	0.073
Retention time* (min)	3.485	7.085
Tailing factor*	1.02	1.13
Theoretical plates*	8245	6432

^{*}Average of six readings

MET and OLM denotes metoprolol succinate and olmesartan medoxomil respectively.

Specificity

Photodiode array detection was used as an evidence of the specificity of the method and to evaluate the homogeneity of the drug peak. The peak purity values for analyte peaks, MET and OLM, were in the range of 999–1000 for drug substance and in the range of 998–1000 for tablets, indicating homogeneous peaks and thus establishing the specificity of assay method.

Determination of Limits of Quantification and

The limit of detection (LOD) and limit of quantitation (LOQ) for MET and OLM were determined at a signal-to-noise ratio of 3:1 and 10:1 respectively, by injecting a series of dilute solutions with known concentration. The LODs for MET and OLM were 0.020 ig/ml and 0.024 ig/ml, respectively and the LOQs were 0.060 and 0.073 ig/ml, respectively (Table 3).

Precision (repeatability)

The precision of the method was studied by determining the concentrations of each drug in the tablets six times. The results of the precision study indicate that the method is reliable (%RSD<2).

Accuracy (recovery test)

Accuracy of the method was studied by recovery experiments. The recovery was performed at three levels, 80 %, 100 %, and 120 % of the label claim of the tablet (25 mg of MET and 20 mg of OLM). The results are shown in Table 4.

Robustness

The robustness of a method is the ability of method to remain unaffected by small changes in parameters like mobile phase composition, flow rate, pH of mobile phase and temperature etc.

MAHAPARALE S P. GONJARI I D and JAYAVEERA K N

Table 4: Recovery studies

Drug	Level of % recovery	%Mean*	8 fand and de via flon	% R8 D	Stan dard error	
MET	80	99.95	0.1213	0.1207	0.0495	
OLM	80	99.72	0.23 17	0.2310	0.0946	
MET	100	99.48	0.1497	0.1489	0.0611	
OLM	100	99.85	0.4527	0.4520	0.1848	
MET	120	99.84	0.32 40	0.3234	0.1323	
OLM	120	99.75	0.1432	0.1428	0.0584	

*Average of six readings

MET and OLM denotes metoprolol succinate and olmesartan medoxomil respectively.

Both drugs and their combination drug product were found to be stable in neutral, thermal, oxidative and photolytic stress conditions but mild degradation was observed in acidic and alkaline conditions. The results obtained by the stress degradation conditions of the drugs show that the method is specific and stability-indicating.

DISCUSSION

The results obtained by the stress degradation conditions of both drugs showed that validated stability-indicating RP-HPLC method is specific, simple, rapid, reproducible, accurate and precise method developed for the quantitative simultaneous estimation of metoprolol succinate and olmesartan medoxomil in combined tablet dosage form. In the future, this method may be applied for routine analysis of both the drugs in API, formulations, dissolution studies, bioavailability and pharmacokinetic studies.

ACKNOWLEDGMENTS

The authors are thankful to the Principal, Padm. Dr. D. Y. Patil College of Pharmacy, Pune for providing excellent research facilities and Emcure Pharmaceuticals Ltd., Pune and Lupin Research Park, Pune for providing the gift sample of olmesartan medoxomil and metoprolol succinate respectively.

REFERENCES

- In Martindale, The extra pharmacopoeia, Sweetman SC, eds. 36th ed. London: Pharmaceutical Press, 2009, p.1338, 1361.
- 2. British Pharmacopoeia, Vol. 2, The controller of Her Majesty's Stationary Office for Department of Health Ministers, London, 2004, p.1303.
- 3. The United States Pharmacopoeia, Vol. 3, United States Pharmacopoeial Convention Inc. Rockville, 2008, p. 2695, 2697.
- In The Merck Index, Budhwari S, eds. 13th ed. Merck and Co; Inc., New Jersey: Whitehouse Station, 2001, p. 6910.
- Kulkarni MN, Kshirsagar RV, Sakarkar DM. Development and validation of spectrophotometric method for determination of metoprolol succinate. Int J Chem Tech Res. 2009;1 (4):1273-1277.

Metoprolol and Olmesartan determination

- Phale MD, Hamrapurkar PD. A validated and simplified RP-HPLC of metoprolol succinate from bulk drugs. Asian J Res Chem. 2009; 2(2): 119-122
- 7. Durga KB, Mounika IN, Shajhan SK, Srinivasa RN, Chandra reddy S, Subramanyam M. RP-HPLC method for estimation of metoprolol in bulk. Int J Sci Innov Discov. 2011; 1(2): 151-157.
- Aqil M, Ali A, Ahad A, Sultana Y, Najmi AK, Saha N. A validated HPLC method for estimation of metoprolol in human plasma. Acta Chromatographica. 2007; 19:130-140.
- Yilmaz B, Asci A, Arslan S. Determination of metoprolol in human plasma and urine by highperformance liquid chromatography with fluorescence detection. J Sep Sci. 2010; 33(13): 1904-1908.
- Yilmaz B. Development and validation of GC/FID and GC/MS methods for determination of metoprolol in pharmaceutical preparations. Alfa Universal Int J Chem. 2010; 1(1): 1-11.
- 11. Singh B, Patel DK, Ghosh SK. Development of Reverse-Phase HPLC method for Simultaneous analysis of metoprolol succinate and hydrochlorothiazide in a tablet formulation. Trop J Pharm Res. 2009; 8 (6):539-543.
- Gao F, Zhang M, Cui X, Wang Z, Sun Y, Jingkai G. Simultaneous quantitation of hydrochlorothiazide and metoprolol in human plasma by liquid chromatography-tandem mass spectrometry. J Pharm Biomed Anal. 2010; 52(1):149-154
- Dongre VG, Shah SB, Karmuse PP, Phadke M, Jadhav VK. Simultaneous determination of metoprolol succinate and amlodipine besylate in pharmaceutical dosage form by HPLC. J Pharm Biomed Anal. 2008; 46(3): 583-586.
- Verma PK, Kamboj VK, Ranjan S. Spectrophotometric estimation of olmesartan medoxomil in tablet dosage form with stability studies. Int J Chem Tech Res. 2010; 2(2):1129-1134.
- 15. Jain P, Jain A, Maliwal D, Jain V. Development and validation of spectrophotometric and RP-HPLC method for estimation of olmesartan medoxomil in tablet dosage form. Int J Pharm Bio Sci. 2010; 1(2): 1-7.

MAHAPARALE S P. GONJARI I D and JAYAVEERA K N

- Ganduri RB, Lanka RA, Pamidi S, Peddareddigari JR, Mohammed M. New RP-HPLC method for the determination of olmesartan medoxomil in tablet dosage form. Eurasian J Anal Chem. 2010; 5(2):145-151.
- Raghunathan K, Reddy JM. Stability indicating RP-HPLC method development and validation of Olmesartan medoxomil. Asian J Pharm Bio Res. 2011; 33-37.
- Sharma RN, Pancholi SS. Validated stability indicating LC-DAD method for determination of Olmesartan medoxomil in tablets exposed to stress conditions. Acta Pharm Sciencia. 2009; 51: 323-331.
- Lui D, Hu P, Matsushima N, Li X, Li L, Jiang J. Quantitative determination of Olmesartan in human plasma and urine by liquid chromatography coupled to tandem mass spectrometry. J Chromatogr B Analyt Technol Biomed Life Sci. 2007; 856:190–197.
- Vaidya VV, Shikha MN, Yetal SM, Joshi SS, Parekh SA. LC-MS-MS Determination of olmesartan in human plasma. J Chromatographia. 2008; 67 (1):147-150.
- Patil KR, Shinde DB. Stability indicating LC method for the simultaneous determination of olmesartan and ramipril in dosage form. Int J Ind Chem. 2011; 2 (2):102-111.
- Chouhan KS, Bhure MV, Hemke AT, Gupta KR, Wadodkar S. Development of RP-HPLC method for estimation of hydrochlorothiazide and olmesartan medoxomil in pharmaceutical formulation. Res J Pharm Bio Chem Sci. 2010; 1(4):78.
- 23. ICH, Q2B- Validation of Analytical Procedures: Methodology, International Conference on Harmonization, 1996; 137.