



SMU
Sikkim Manipal University



SMU Medical Journal



ISSN : 2349 – 1604 (Volume – 3, No. 1, January 2016) Review article

Indexed in SIS (USA), ASI (Germany), I2OR & i-Scholar (India) and SJIF (Morocco)
databases

Impact Factor: 3.835 (SJIF)

Problems and Proceedings to Antiretroviral Therapy for the Treatment of AIDS

Chinmaya keshari sahuo¹, Surepalli Ram Mohan Rao², Muvvala Sudhakar³

¹Department of pharmaceutics,Osmania University College of Technology,OsmaniaUniversity,Hyderabad,Telangana,India. ²Mekelle Institute of Technology, Mekelle University, Mekelle, Ethiopia. ³Department of pharmaceutics, Malla Reddy College of Pharmacy,Maisammaguda,secunderabad,Telangana,India.

Corresponding author

Chinmaya Keshari Sahoo

Phone no.-+918688410043,+919553665627

Manuscript received : 17.11.2015

Manuscript accepted: 22.12.2015

Abstract

Long term use of antiretroviral therapy suppresses HIV infection in real. But due to mutation there is chance of drug resistance, drug toxicity, drug penetration, adherence to therapy, viral replication in cellular reservoirs and augmentation of host immune responses. Viral replication

causes the accumulation of drug resistance, mutations, increase in viral loads and disease progression. The optimization of antiviral therapy by the understanding of the pathogenesis of drug resistant HIV-1 is essential. In the present review outlines various problems to HIV therapy and discussions about the success of antiviral therapy.

Keywords: Replication, drug resistance, antiviral therapy, progression.

Introduction

Acquired immune deficiency syndrome (AIDS) is considered one of the most dangerous and a pandemic [1] disease which is present over a large demographic area of the world. The disease may cause Kaposi's sarcoma, pneumocystis carini pneumonia [2] and serious opportunistic infections. AIDS is the most serious infectious [3] disease and actively spreading worldwide among society. Human immunodeficiency virus infection/acquired immune deficiency syndrome (HIV/AIDS) is a disease of human immune system caused by infection with human immune deficiency virus. AIDS is called when a person infected with HIV has a CD4+ count of less than 200cells/ μ L or has an AIDS defining condition. While antiretroviral treatment [4,5] reduces the risk of death and complications from the disease these medications are expensive and may be associated with side effects. Antiviral agent has made HIV/AIDS a more manageable disease in some industrialized nations and several vaccines [6] are about to enter phase III clinical trials.HIV will doubtless continue to impose a terrible burden of morbidity and mortality. More importantly establishing the ground rules that underpin the evolution of HIV will lead to better vaccines and antiviral agents. The present review gives idea about targets of HIV by antiretroviral drugs, problems and proceedings to antiretroviral therapy and tests for HIV-1 drug resistance.

Current targets of HIV antiretroviral drugs

The drug resistance is due to monotherapy.Hence therapy with potent combinations of three or more antiretroviral [7] drugs show that rapidly reduce circulating levels of plasma HIV to below detectable levels for periods of several years or more(Table-1).

Table (1): Different classes of HIV-1 antiretroviral drugs

Class	Drugs	Viral target
Nucleoside reverse transcriptase inhibitors	Zidovudine, didanosine, zalcitabine, stavudine, lamivudine, abacavir, tenofovir	Reverse transcriptase
Non-nucleoside reverse transcriptase inhibitors	Nevirapine, delavirdine, efavirenz	Reverse transcriptase
Protease inhibitors	Saquinavir, indinavir, ritonavir, nelfinavir, Amprenavir, lopinavir, atazanavir	Protease
Fusion inhibitors	Enfuvirtide	Envelope gp41

Drug targets by Polymerase protein (Pol)

The polymerase protein (Pol) of HIV-1 is produced as a Gag pols (Pr¹⁶⁰ Gag-pol) fusion polyprotein [8]. The pol gene precursor is divided to produce three viral enzymes such as protease, reverse transcriptase and integrase. The HIV-1 protease enzyme plays a vital role acting to specifically cleave Gag and Pol precursor polypeptides into functionally active proteins. HIV protease is an aspartic proteinase and is responsible for break of the Gag (p55) and Gag-Pol (p160) polyprotein products yielding the functional core proteins (p17, p24, p7, p6) and essential enzymes (reverse transcriptase, integrase, protease) required to produce mature HIV [9]. When HIV protease is chemically blocked the formation of these core proteins is disrupted and assembled to form virions. These are immature and non infectious. The HIV-1 reverse transcriptase (RT) enzyme is a RNA-dependent DNA polymerase enzyme which is responsible for replicating the RNA genome [10]. The RT enzyme changes the single stranded virion into doubled stranded DNA for subsequent integration into the host cell genome. RT is obtained from a Gag-Pol precursor that is processed by protease to yield a heterodimeric enzyme composed of a 66kDa protein (p66) and a 51 kDa protein (p51) [11]. The 66 kDa protein (p66) is degraded to p51 and p15. Polymerase activity resides within the p51 fragment and RNAase activity is linked

with the presence of p15. Integrase is a 31kDa protein derived from the C terminal portion after the processing of Pr¹⁶⁰ Gag-Pol and is required for integration of a double stranded DNA copy of the viral RNA genome into the host chromosome.

Drug targets by envelope protein

HIV fusion and entry occur via the interaction of trimeric envelope gp160 spike (gp120 and gp41) with receptor molecules on the surface of target cells [11].The fusion process is a vital step in the viral replicative cycle making it an attractive target for antiretroviral drugs. Fusion inhibitor such as enfuvirtide (T-20) target is the crucial fusion step of the viral life cycle.Enfuvirtide is administered parenterally and inhibits fusion of the viral and cell membranes by binding to a portion of thegp41 molecule[12].

Drug resistance and antiretroviral therapy

Protease inhibitors (PIs) and drug resistance

The HIV protease enzyme is a dimeric aspartyl protease essential for the posttranslational cleavage of precursor Gag-Pol polyproteins during virion maturation to generate building blocks required for assembly of new virus particles [13,14].The activity of this protein is required for virus infectivity rendering it a major drug target. Resistance mutations in the protease gene result from amino acid substitutions at or near the active site interfering with binding of the inhibitor because of conformational perturbations or to amino acids lying outside the active region. The cleavage site mutations do not produce drug resistance themselves, but compensate for changes in protease that result from primary and secondary mutations. Resistance to PIs emerges rapidly when these inhibitors are administered at inadequate doses or as part of suboptimal regimens [15].Generally, high-level resistance to PIs results from the sequential accumulation of amino acid substitution in the Pr gene along pathways that usually vary between different PI drugs [16].The first approved PIs were saquinavir (SQV) and indinavir (IDV) [17].These drugs are potent and require the emergence of multiple mutations before high level resistance. The resistance to SQV often confers resistance to IDV and vice versa [18].When viral suppression to

below the limit of detection is not achieved in a dual NRTI plus PI regimen, early mutations often occur to lamivudine followed by mutations associated with resistance/cross resistance to other NRTIs. Nelfinavir (NFV) has a lower genetic barrier to resistance than SQV and IDV. It is susceptible to the single D30N and L90M mutations which develops quickly in PI naive patients. The presence of two or more mutations including D30N, G48V, 150V, V82A/F/T/S, 194V and L90M generally confers cross resistance to all the three of these PIs [19] whereas a single mutation at codon 30 does not infer cross resistance to other PIs. Amprenavir (APV) when used as a first-line PI is inclined to select for the 150V mutation conferring cross-resistance to lopinavir (LPV). LPV has a high genetic barrier to resistance [20], but a recent study has suggested that APV-selected resistance may confer cross-resistance to LPV44. Atazanavir (ATV) is susceptible to the 150L mutation. The patients who have taken PI-based therapy before commencing with ATV a number of both primary and secondary mutations may be associated with ATV resistance [21].

Nucleoside reverse-transcriptase inhibitors (NRTIs)

NRTIs inhibit the reverse transcriptase enzyme by competing with the endogenous nucleosides for the incorporation into the DNA chain generated by reverse transcription of HIV RNA. All nucleoside analogues must be triphosphorylated within the cell resulting inhibition of reverse transcriptase activity and premature chain termination. Resistance to NRTIs develops from nucleotide changes with the RT gene and the subsequent generation of amino acid substitutions in the RT enzyme [22]. NRTI induces a predictable set of genetic changes generally with primary mutations arriving first and secondary mutations developing during continued therapy. The resistance to zidovudine develops with the sequential selection of specific mutations in the RT gene including codons [23] 41(M41L), 67(D67N), 70(K70R), 210(L210W), 215(T215Y) and 219(K219Q). These mutations may result resistance to didanosine, zalcitabine, tenofovir and abacavir. Mutations linked with didanosine resistance involve codons 65, 74 and 184. Two sets of mutations such as Q151M complex and the T69S insertion mutations confer resistance to all the currently available NRTIs. The primary

codon change in Q151M complex [24] is linked with secondary mutations at codons 62(A62V),75(V75I),77(F77L) and 116(F116Y) which reduces the sensitivity to NRTIs. The T69S-S-S or T69S-S-A insertion mutations occur after prolonged treatment with multiple nucleosides which resistance to all NRTIs.

Non-nucleoside reverse transcriptase inhibitors (NNRTIs)

NNRTIs are non competitive inhibitors of HIV-1 RT which bind to a hydrophobic cavity near the active site of reverse transcriptase, causing a conformational change in the enzyme. NNRTI binding [25] sites are restricted to beta sheets consisting codons 100-110 and 180-190. The common mutations [26] in HIV selected by NNRTIs are L100I, K103N, V106A, V108I, Y181C/I, Y188C/L, G190A/E/S, P225H and P236L3. Y181C and K103N are linked with significant cross-resistance between nevirapine, delavirdine and efavirenz. The Y181C mutation is resistant to nevirapine and delaviridin, but not to efavirenz [27]. Y181C reduces NNRTI binding affinity leading drug resistance while the K103N mutation acts by preventing the formation of the binding pocket [28].

Resistance to the fusion inhibitor T-20

Resistance mutation to T-20 appears mainly in the Env gp 41 region. Substitutions occurred near the N-terminus of the HR-1 region in the highly conserve GIVQQQ sequence known to be critical for fusion [29].

Factors of HIV-1 drug resistance

HIV genetic heterogeneity

HIV-1 infection is characterized by a high degree of genetic variability within infected persons with the population present at a certain time point within an infected person consisting of a complex mixture of heterogeneous strains [30] termed quasispecies. Quasispecies differ in their antigenic and phenotypic properties and compete among themselves for survival [31] and propagation. The subsequent overgrowth or dominance of a certain viral strain over another is

largely determined by its relative adaptation to a given intra host environment a factor particularly relevant for the emergence of drug resistance variants.

Error in reverse transcriptase enzyme

HIV-1 mutation rate is nearly one hundred times higher than those of DNA viruses, bacteria or other eukaryotes [32]. The rate of nucleoside substitutions introduced by reverse transcriptase is approximately 10^{-4} per nucleotide per cycle of replication which is equal to one nucleotide substitution per genome [33] during a single replication cycle. Insertions, deletions and duplications contribute to the genetic heterogeneity [34] of HIV-1. HIV-1 turnover is rapid and it is estimated that approximately 10^9 virions per day are generated in an infected individual. The lifespan of plasma virus and virus producing cells is very short with a half-life of approximately two days and an almost complete replacement of wild type strains by drug resistant virus occurs in plasma within 2-4 week. During antiretroviral treatment rapid viral turnover in combination with a high mutation rate is a primary factor behind the emergences of HIV variants with antiretroviral drug resistance.

Genetic recombination

Each retrovirus particles contains a dimer RNA genome and a reverse transcriptase enzyme which can switch templates during proviral synthesis. Drug resistant mutations in HIV-1, leading to increased resistance to a particular drug by recombination or the generation of multi drug resistant variants. Recombination may cause to the acquisition of mutations that compensate for a loss in viral fitness or replicative capacity due to previous acquisition of resistance mutations. The recombination creates multiple drug resistant virus out of two single drug resistant strains, it is generally believed that the capacity of the virus to recombine facilitates the evolution of drug [35,36] resistance.

Selective pressures imposed by antiretroviral drugs

Drug resistant HIV strains are generally reduced fitness [37] compared to wild-type counterparts in the absence of drugs. The decrease in viral fitness is accompanied by the

emergence of primary mutations. Continued drug selective pressure then allows the virus to select secondary mutations which compensate for the primary mutations allowing restoration of wild type enzymatic activity of the enzyme (Pr or RT) targeted by the drug. Viriants exist within the population that are naturally resistant to some extent to a particular drug before the commencement of HIV antiretroviral therapy. The presence of an antiviral drug changes the selective pressure on the viral population. When drugs that only partially inhibit HIV-1 replication are administered the resulting evolutionary pressure selects for resistant strains. Resistance emerges at a rate that is proportional to the frequency of pre existing variants and their relative growth advantage in the presence of drug [38].

Drug pharmacokinetics

The pharmacological activity of antiretroviral drugs is dependent on unbound drug entering cells that harbor HIV and multi drug combination therapy. The pharmacokinetics of orally administered antiretroviral involves absorption, first-pass metabolism in the intestine and liver, systemic distribution, metabolism and removal (excretion). Many antiretroviral bind to plasma proteins [39], influencing uptake into cells, as only unbound drugs in plasma can pass across the cell membrane efficiently. Important pharmacokinetic [40] parameters to consider include the volume of distribution (concentration of drug in plasma for a given amount of drug in the body), rate of clearance (efficiency of drug excretion), drug half-life (determines the course of accumulation of the drug in the body in chronic dosing) and the degree of fluctuation within a dosing interval. These parameters can vary significantly between patients, resulting in differences in drug absorption, drug metabolic, excretory activity, drug distribution and the overall efficacy of drug regimens [41].

Suboptimal drug penetration in CNS

There is limit of drug penetration to central nervous system(CNS).The blood-brain barrier is located between the blood and brain tissue and the blood CSF barrier is formed by the choroid plexus. High plasma protein binding of protease inhibitors and their unidirectional efflux by P-

glycoprotein membrane proteins in the blood-brain barrier limit CNS penetration and absorption [42] of antiretrovirals. Thus the CNS represents a site where ongoing viral replication may occur in the absence of antiretroviral suppression.

Patient adherence

Adherence prevents the emergence of resistant strains, but incomplete patient adherence coupled with an array of other pharmacologic factors result in the presence of a heterogeneous population and the possibility of selecting for viral resistance. Poor access to healthcare providers and counseling and broader issues such as poverty and poor literacy add to difficulties in patient adherence [43] in these areas. Drug-use, high-risk behaviors and depression contribute to poor adherence. The incidence of adverse effects [44] associated with the administration of antiretroviral agents is dependent on a number of factors including the ethnic origin of the patient, use of additional medications and host factors. Adverse reactions to antiretroviral therapy may affect its clinical efficacy through adherence problems. Adverse effects linked with antiretroviral agents include mitochondrial toxicity, hypersensitivity, lipodystrophy, dyslipidaemia and type 2 diabetes. Other effects include cardiomyopathy, peripheral neuropathy and pancreatitis. All antiretroviral drugs are linked with liver dysfunction through either direct or indirect [45,46] mechanisms.

Viral reservoirs

The reservoirs of HIV occurs in various tissues e.g CNS, male genital tract etc. Some anatomical sites may be non-permissive to immune surveillance and effective drug penetration, thus serving as potential sites of persistent HIV replication e.g. the respiratory, gastrointestinal and reproductive tracts [47].

Cellular reservoirs of HIV-1

Cellular reservoirs of HIV-1 arise from the ability of HIV to infect a variety of immune cell types including monocytes, macrophages, NK cells and T lymphocytes in addition to other non-immune based cells. Cellular reservoirs of HIV include memory CD4+ T lymphocytes,

blood monocytes and macrophages /cells of macrophages [48] lineage.

Tests for HIV-1 drug resistance

Genotypic testing

Genotypic assays use nucleic acid sequencing methods to detect all mutations within the RT and Pr genes. It may use line probe or clip-based assays to identify mutations within the RT and Pr genes to be associated [49] with drug resistance. Genotypic assays depend upon amplification of the Pr and RT genes from viral RNA in plasma (by means of RTPCR). Genotypic testing is inexpensive and readily available in many laboratories, but there are limitations to this approach of assessing drug susceptibility which include variable reproducibility and increased potential for laboratory error. The interpretation of drug resistance profiles through sequence-based analysis is also an issue and inconsistencies in this area are evident between the various internet resources. In additions current assays only detect viruses representing 5-20 percent of the total population and resistance present in minor subpopulations is often missed [50].

Phenotypic testing

The functional characteristics and growth properties of a viral isolate are referred to as the viral phenotype. Phenotyping measures the susceptibility of the virus to inhibition by a particular drug. The commercial recombinant assays such as AntiVirogram (Virco) and Phenosense (Virologic) may be used for testing [51]. These assays relay on the incorporation of plasma derived HIV-1 RT and Pr genes into the backbone of an HIV-1 reference strain. The recombinant is then tested ex vivo to measure the IC50 and to measure a fold-change in susceptibility compared to wild type virus. Limitations of these assays include interpretation problems, as biological or virologic cut-offs measured by the assays do not incorporate achievable drug levels and can thus over or under-estimate the likelihood of a clinical response to a given drug.

Conclusion

AIDS is a pandemic disease which spreads globally in human society. Hence there is need for effective treatment for controlling HIV1 infection. There is control in the mortality and morbidity of HIV infected individuals by the implementation of antiviral therapy. Use of different combination therapies target HIV life cycle and facilitate the successful long term survival of patients.

Acknowledgements

The authors would like to acknowledge the contributions of Osmania University of Technology, Department of pharmaceutics, Osmania University, Hyderabad, Telangana, India for providing necessary facilities to carry out the review work. This study was part of a Ph.D thesis under Osmania University, Hyderabad.

References

- [1] Kallings LO (2008) The first postmodern pandemic 25 years of HIV/AIDS; *Journal of Internal Medicine*. 263, 218-43.
- [2] Centers for Disease Control (CDC) (2011) Kaposi's sarcoma and Pneumocystis pneumonia among homosexual men New York city and California; *Morbidity and mortality weekly report*. 30(25), 305-8.
- [3] Thomas Kerr WS (2011) HIV treatment as prevention and the role of applied social science research; *J AIDS Clinic Res*. 2, 102.
- [4] Thigpen MC, Kebaabetswe PM, Paxton LA et al.(2012) Antiretroviral preexposure prophylaxis for heterosexual HIV transmission in Botswana; *N Engl J Med*.367, 423-434,.
- [5] Velmurugan S., Ali MA and Kumar P (2014) Microparticulate drug carriers a promising approach for the delivery of anti HIV drugs; *Int J.Pharm. Pharm Sci*. 6(2), 31-39,
- [6] Barbara E., Aurelio C., Paolo M et al. (2014) Challenges in HIV vaccine research for treatment and prevention; *Frontiers in Immunology*. 5, 1-11.

- [7] Richman DD (2001) HIV chemotherapy; *Nature*. 410, 995-1001.
- [8] Jacks T., Power MD, Masiarz FR, Luciw PA, Barr PJ and Varmus HE (1988) Characterization of ribosomal frame shifting in HIV-1 gag-pol expression; *Nature*. 331, 280-3.
- [9] Peng C., Ho BK, Chang TW and Chang NT (1989) Role of human immunodeficiency virus type 1-specific protease in core protein maturation and viral infectivity; *J Virol*. 63, 2550-6.
- [10] Popovic M., Sarngadharan MG, Read E and Galloway RC (1984) Detection, isolation and continuous production of cytopathic, retroviruses (HTLV-III) from patients with AIDS and pre-AIDS; *Science*. 224, 497-500.
- [11] Earl PL, Doms RW and Moss B (1990) Oligomeric structure of the human immunodeficiency virus type I envelope glycoprotein; *Proc Natl Acad Sci*. 87, 648-52.
- [12] Wild C Greenwell T and Mathews T (1993) A synthetic peptide from HIV-1 gp41 is a potent inhibitor of virus mediator cell-cell fusion; *AIDS res hum retroviruses*. 9, 1051-3.
- [13] Todd S., Anderson C., Jolly DJ and Craik CS (2000) HIV protease as a target for retro viral vector-mediated gene therapy; *Biochim Biophys Acta*. 1477, 168-88.
- [14] Kaplan AH, Manchester M., Smith T., Yang YL and Swanstrom R (1996) Conditional human immunodeficiency virus type 1 protease mutants show no role for the viral protease early in viral replication; *J Virol*. 70, 5840-4.
- [15] Molla A., Korneyeva M., Goa Q., Vasavanonda S., Schipper PJ, Mo HM, et al. (1996) Ordered accumulation of mutation in HIV protease confers resistance to ritonavir; *Nat Med*. 2, 760-6.
- [16] Ridky T and Leis J (1995) development of drug resistance to HIV-I protease inhibitors; *J Biol Chem*. 270, 21-3.
- [17] Mammano F., Trouplin V., Zennou V and Clavel F (2000) Retracing the evolutionary pathways of human immunodeficiency virus type 1 resistance to protease inhibitors: virus fitness in the absence and in the presence of drug; *J Virol*. 74, 8524-31.
- [18] Hertogs K., Bloor S., Kemp SD, Van den Eynde C., Alcorn TH, Pauwels R., et al. (2000) Phenotypic and genotypic analysis of clinical HIV-1 isolates reveals extensive protease inhibitor cross-resistance: a survey of over 6000 samples; *AIDS*. 14, 1203-10.
- [19] Condra JH, Schleif WA, Blahy OM, Gabryelski LJ, Graham DJ, Quintero JC, et al. (1995) In vivo emergence of HIV-1 variants resistant to multiple protease inhibitors; *Nature*. 375, 569-71.

- [20] Parkin NT, Chappey C and Petropoulos CJ (2003) Improving lopinavir genotype correlations: novel mutation patterns and amprenavir cross-resistance; *AIDS*. 17, 955-61.
- [21] Colon RJ, Thiry A., Limoli K and Parkin N (2003) Activities of atazanavir (BMS-232632) against a large panel of human immunodeficiency virus type-1 clinical isolates resistant to one or more approved protease inhibitors; *Antimicrobe Agents Chemother.* 47, 1324-33.
- [22] Larder BA and Kemp SD (1989) Multiple mutations in HIV-1 reverse transcriptase confer high-level resistance to zidovudine (AZT); *Science*. 246, 1155-8.
- [23] Kellam P., Boucher CA and Larder BA (1982) Fifth mutation in human immunodeficiency virus type 1 reverse transcriptase contributes to the development of high-level resistance to zidovudine; *Proc Natl Acad Sci USA*. 89, 1934-8.
- [24] Iversen AK, Shafer RW, Wehrly K., Winters MA, Mullins JI, Chesebro B, et al. (1996) Multidrug-resistance human immunodeficiency virus type 1 strains resulting from combination antiretroviral therapy; *J Virol.* 70, 1086-90.
- [25] Kohlstaedt LA, Wang J., Friedman JM, Rice PA and Steitz TA (1992) Crystal structure at 3.5 Å resolution of HIV-1 reverse transcriptase complexed with an inhibitor; *Science*. 256, 1783-90.
- [26] Deeks SG (2001) Durable HIV treatment benefit despite low-level viremia: reassessing definitions of success or failure; *JAMA*. 286, 224-6.
- [27] Baldanti F., Paolucci S., Maga G, Labo N., Hubscher U., Skoblov Ay, et al. (2003) Nevirapine-selected mutations y181I/C of HIV -1 reverse transcriptase confer cross-resistance to stavudine; *AIDS*. 17, 1568-70.
- [28] Wei X., Ghosh SK, Taylor ME, Johnson VA, Emini EA, Deutsch P, et al. (1995) Viral dynamics in human immunodeficiency virus type 1 infection; *Nature*. 373, 117-22.
- [29] Horn T (2002) Entry and fusion inhibitors: an update. In: Cohen C. Moore J, editors; *The Prn notebook vol.7*. New York USA: Physicians Research Network. 16-22.
- [30] Meyerhans A., Cheynier R, Albert J., Seth M., Kwok S., Sninsky J, et al. (1989) Temporal fluctuations in HIV quasispecies in vivo are not reflected by sequential HIV isolation; *Cell*. 58901-10.
- [31] Fisher AG, Ensoli B., Looney D., Rose A., Gallo RC, Saag MS et al. (1988) Biologically diverse molecular variants within a single HIV-1 isolate; *Nature*. 334, 444-7.

- [32] Mansky LM and Temin H (1995) Lower in vivo mutation rate of human immunodeficiency virus type 1 than that predicted from the fidelity of purified reverse transcriptase; *J Virol.* 69, 5087-94.
- [33] Nowak M (1990) HIV mutation rate; *Nature.* 347,522.
- [34] Hu WS and Temin HM (1990) Retroviral recombination and reverse transcription; *Science.* 250,1227-33.
- [35] Yusa K., Kavlick MF, Kosalaraksa P and Mitsuya H (1997) HIV-1 acquires resistance to two classes of antiviral drugs through homologous recombination; *Antiviral Res.* 36, 179-89.
- [36] Burke DS (1997) Recombination in HIV: an important viral evolutionary strategy; *Emerg Infect Dis.* 3, 253-9.
- [37] Clavel F., Race E and Mammano F (2000) HIV drug resistance and viral fitness; *Adv pharmacol.* 49, 41-66.
- [38] Berkhout B (1999) HIV-1 evolution under pressure of protease inhibitors: climbing the stairs of viral fitness; *J Biomed Sci.* 6,298-305.
- [39] Boffito M., Back DJ, Blaschke TF, Rowland M, Bertz RJ, Gerber JG, et al. (2003) Protein binding in antiretroviral therapies; *AIDS Res Hum Retroviruses.* 19, 825-35.
- [40] Lesho EP and Gey DC (2003) Managing issues related to antiretroviral therapy; *Am Fam physician.* 68, 675-86.
- [41] Shafer RW and Vuitton DA (1999) Highly active antiretroviral therapy (HAART) for the treatment of infection with human immunodeficiency virus type 1; *Biomed Pharmacother.* 53,73-86.
- [42] Kim RB, Fromm MF, Wandel C., Leake B., Wood AJ, Roden DM, et al. (1998) The drug transporter P-glycoprotein limits oral absorption and brain entry of HIV-1 protease inhibitors; *J Clin Invest.* 101, 289-94.
- [43] Nieuwerk PT, Sprangers MA, Burger DM, Hoetelmans RM, Hungen PW, Danner SA, et al. (2001) Limited patient adherence to highly active antiretroviral therapy for HIV-1 infection in an observational cohort study; *Arch Intern Med.* 161,1962-8.
- [44] Lewis W and Dalakas MC (1995) Mitochondrial toxicity of antiviral drugs; *Nat Med.* 1,417-22.

- [45] Hadigan C., Meigs JB, CORcoran C., Rietschel P., Piecuch S., Basgoz N, et al.(2001)Metabolic abnormalities and cardiovascular disease risk factors in adults with human immunodeficiency virus infection and lipodystrophy; Clin Infect Dis. 32,130-9.
- [46] Noor MA, Lo JC, Mulligan K., Schwarz JM, Halvorsen RA, Schambelan M,et al.(2001) Metabolic effects of Indinavir in healthy HIV-seronegative men; AIDS.15,11-8.
- [47] Saksena NK and Potter SJ (2003) Reservoirs of HIV-1 in vivo implications for antiretroviral therapy; AIDS Rev.5, 3-18.
- [48] Potter SJ, Dwyer DE and Saksena NK (2003) Differential cellular distribution of HIV-1 drug resistance in vivo:evidence for infection of CD8+T cells during HAART; Virology 305,339-52.
- [49] Wyseur LA, Rombout A., Louwagie J., Scarcez T., Verhofstede C,et al. (1997) Line probe assay for rapid detection of drug-selected mutations in the human immunodeficiency virus type 1 reverse transcriptase gene; Antimicrob Agents Chemother. 41,284-92.
- [50] Charpentier C., Dwyer DE, Mammano F., Lecossier D., Clavel F and Hance AJ(2004) Role minority populations of HIV-1 in the evolution of viral resistance to protease inhibitors; J Virol. In press.
- [51] Petropoulos CJ, Parkin NT, Limoli KL, Lie YS, Wrin T., Huang W et. al. (2000) A novel phenotypic drug susceptibility assay for human immunodeficiency virus type-1. Antimicrob Agents Chemother. 44,920-8.

Authors Column



Chinmaya Keshari Sahoo is an Assistant. Professor of Pharmaceutics at Princeton College of Pharmacy, Hyderabad. Mr. C.K.Sahoo did his M.Pharm in pharmaceutics from Sri Jayadev College of Pharmaceutical Sciences, Bhubaneswar under Biju Patnaik University of Technology, Odisha and is presently pursuing Ph.D. from Osmania University, Hyderabad. His main areas of interest in research include in controlled drug delivery system of various osmotic pump tablets. He has published 28 articles in various international journals as well as represented more than 5 papers in various international and national conferences.