# Calophyllum apetalum interceded blend of silver nanoparticles and their antimicrobial impact

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# Abstract

**Background/Objective:** synthesis of silver nanoparticles by the extract of *Calophyllum apetalum* and their characterization to find out size and morphology of the particles.

**Methodology**: The characterization was done by X-Ray diffractive (XRD), UV- Visible spectroscopy and scanning electron microscopy (SEM). X-Ray diffraction (XRD) was used to find out the particle size and SEM image was used to determine its morphology.

Findings: 94nm silver nanoparticles were synthesized.

Improvements/ Application: As antibacterial agents.

Key words: Calophyllum apetalum, UV-VIS, SEM, XRD, silver nanoparticles, antibacterial.

## **1. Introduction**

Nanobiotechnology is one of the alternative options to chemical and physical strategies for the arrangement of nanoparticles. In the blink of an eye, nanotechnology is applying in different circumstances, for example, biosensing, cell imaging, focused on medication conveyance, simulated inserts, as lithium battery, photograph catalysis, sunlight based cells etc [1,2]. Organic strategy for amalgamation incorporates decrease by the constituents of plants, growths, microbes, actinomycetes, yeast and algae [3-5]. Aside from such a large number of non-medicinal applications, silver nanoparticles got significance in light of their physico-chemical properties, i e., high electrical and warm conductivity, surface-improved Raman dispersing, synthetic solidness, synergist movement and non direct optical behavior [6]. Green union of silver nanoparticles has its own points of interest over compound or physical strategies because of its cost viability, environment friendliness [7,8].

In the range of medicinal sciences silver nanoparticles are being utilized as antimicrobial, anti-inflammatory, and antiviral specialists furthermore for refinement of water [9]. In Plant interceded amalgamation of silver nanoparticles, leaves have been effectively used [10]. In any case, no report has been issued on the creation of silver nanoparticles utilizing leaf concentrate of *Calophyllum apetalum* Willd.

Calophyllum apetalum (Calophyllaceae) is a folkloric herb known for its therapeutic values usually known as Alexandrian Laurel. *C. apetalum* seed oil is utilized ordinarily to treat ailment and infection. Xanthonoids and apetalinones were separated from the roots and stem barks of *Calophyllum apetalum* separated from known mixes like calozeyloxanthone and zeyloxanthonone [11]. Coumarins were rich in this spiecies. Dipyranocoumarin,  $\alpha$ -hydroxytomentolide was confined from the leaves of *Calophyllum apetalum* together with the known mixes friedelin, apetalactone, inophyllum and canophyllol [12].

Thus, we clarified an eco-accommodating strategy for orchestrating and balancing out silver nanoparticles by utilizing naturally dynamic leaf concentrate of *Calophyllum apetalum*. What's more, incorporated silver nanoparticles and their portrayal by UV-vis spectroscopy, Scanning Electron Microscope (SEM), X-Ray Diffraction (XRD) studies are additionally reported. Aside from this, the integrated silver chloride nanoparticles were subjected to bactericidal exercises. Assessment of antibacterial action of integrated silver nanoparticles is likewise reported as a natural use of our work.

# 2. Materials and Methods

#### 2.1 Materials

Silver nitrate was obtained from Sigma-Aldrich Chemicals. All glass products were washed with refined water and dried in an oven before use. Crisp leaves of *Calophyllum apetalum* were gathered from the Agumbeghats, Udupi locale, Karnataka, India.

# 2.2. Methods

## 2.2.1. Preparation of leaves concentrate

The gathered new leaves of *Calophyllum apetalum* were washed a few times with twofold refined water. 10 gm of leaves powder was bubbled in Erlenmeyer flask contained 30 mL of deionized water for 30 min and separated through the Whatman channel paper No 1. At that point the filtrate was gathered in 100 mL Erlenmeyer flask and put away at 4<sup>o</sup> C for further utilize.

# 2.2.2. Synthesis of silver nanoparticles

To create silver nanoparticles, 10 ml of concentrated leaf concentrate was added to 25 ml of crisply arranged 5mM silver nitrate and mixed constantly for 10 min at 30<sup>o</sup> C. Lessening happened quickly as demonstrated by a light yellow to dull cocoa shading after 40 min, showing the developments of silver nanoparticles.

The silver nanoparticles got were purified by centrifugation in Remi rotator at 10,000 rpm for 10 min. Supernatant was disposed of and the pellet was washed thrice with deionized water to expel unreacted  $AgNO_3$  and plant separate. The immaculate pellet was gathered, air dried and safeguarded for further portrayal.

# 2.2.3. Characterization of silver nanoparticles

## 2.2.3.1. UV-Vis Spectral Analysis.

The blended silver nanoparticels by leaf concentrate were commented by UV-vis spectroscopy. Spectra of assimilation were gotten by UV-vis spectrophotometer (UV3600 - Shimadzu UV-Vis-NIR Spectrophotometer) utilizing 10 mm optical-way length quartz cuvette at a resolution of 0.5 nm somewhere around 350 and 550 nm at Center for Nano science and Engineering, Indian Institute of Science, Bangaluru.

## 2.2.3.2. Scanning electron microscopy (SEM) examination

Scanning electron microscopepicture was gotten by utilizing ZEISS Ultra 55 SEM machine worked 5 kV at Center for Nano science and Engineering, Indian Institute of Science, Bangaluru. A dainty film of the specimen was set up on a carbon covered copper framework by simply keeping little measure of the sample on the lattice. The readings and photographic sweep were taken at 10000X, 25000X and 50000X amplification with steady voltage and at various points.

## 2.2.3.3. X-Ray Diffraction (XRD) study

X-Ray Diffraction (XRD) is a fast diagnostic system basically utilized for stage distinguishing proof of a crystalline structure of material and can give a data on unit cell measurements. The material to be analysed is finely ground, homogenized, and normal mass piece was resolved. The size and morphology of the silver nanoparticels were dictated by XRD (Rigaku, SmartLab X-beam Diffractometer) at Center for Nano science and Engineering, Indian Institute of Science, Bangaluru at a voltage of 40 keV and a current of 30 mA with Cu K $\alpha$  radiation, step size – 0.02, speed – 5°/min with a wavelength of 1.5418 Å and at 20 point.

## 2.2.3.4. Antibacterial action

Antibacterial impact of the silver nanoparticles was screened by streak plate strategy with three unique conentrations against *Staphylococcus aureus, Klebsiella pneumoniae, Escherichia coli* and *Pseudomonas aeruginosa* [13]. The three distinct concentraions, 10, 50 and 100  $\mu$ l of each of the test mixes were exchanged independently to the test tubes containing 1 ml of bacterial broth. Treated bacterial test tubes were incubated for zero h, 2 h and 4 h. At every interim of incubation period, one loop full of treated bacterial specimen was inoculated on Mueller Hinton Agar (MHA) plate and kept for incubation for another 18-24 h at 37<sup>o</sup> C. MHA plates were watched for bacterial development to survey the activity of nanoparticles.

#### 3. Results and discussion

#### 3.1. Synthesis of silver nanoparticles

The leaf of Calophyllumapetalum was initially blended with silver nitrate solution. At that point light yellow shade of fluid concentrates changed to dim chestnut shading after 40 min, showing the developments of silver nanoparticles (Figure 1). The shading change affirmed that mixes present in the leaf extract decreased silver metal particles to silver nanoparticles.

Figure 1. Calophyllum apetalum leaf extract, before and after treating with silver nitrate



## 3.2. UV-Vis Study of silver nanoparticles

UV–vis studies were completed to distinguish the development of silver nanoparticles and range demonstrated surface plasmon reverberation (SPR) for silver nanoparticles [14]. Blended silver nanoparticles demonstrated most extreme absorbance top at 410nm Figure 2, which was particular for the union of silver nanoparticles [15]. It is surely understood that because of Mie dissipating, the colloidal silver nanoparticles show assimilation at the wavelength from 390 to 420nm [16,17].





#### 3.3. SEM studies

Figure 3 indicates typical SEM pictures of silver nanoparticles incorporated by utilization of leaf concentrate of *Calophyllum apetalum*. The silver nanoparticles are of 94 - 114 nm quantifiable extent and circular fit as a fiddle. The SEM picture of silver nanoparticles was because of cooperations of hydrogen bond and electrostatic communications between the bioorganic topping atoms bound to the silver nanoparticles [18]. From the SEM studies it was watched that, the orchestrated silver nanoparticles are free scattered to each other (Figure 3).

Figure 3. SEM photographs of silver nanoparticles obtained using leaf extract of Caplophyllumapetalum



#### 3.4. X-Ray Diffraction (XRD) study

The crystalline way of the silver nanoparticles was further affirmed by XRD. Figure 4 demonstrates the XRD pattern of silver nanoparticles integrated by *Calophyllum apetalum* leaf separate after complete decrease of  $Ag^+$  to  $Ag^0$ . XRD investigation indicated six diverse diffraction tops at 20 estimations of 27.72°, 32.09°, 46.15°, 54.80°, 57.34° and 76.61° which recorded the planes (111), (200), (231), (311), (222) and (400) and contrasted and the information of JCPDS (Joint Committee on Powder Diffraction Standards), File No. 87-0720 and it demonstrated that the face focused cubic structure of metallic silver were made out of unadulterated crystalline silver.

The normal molecule sizes were figured by Scherrer equation [19]. These outcomes were demonstrated similarity with before aftereffects of silver nanoparticles orchestrated by leaf concentrate of geranium (*Pelargonium graveolens*) [20].





#### 3.5. Antibacterial study

Silver mixes and silver particles had been as of now verifiably perceived and connected as antibacterial specialists in avoidance of wound infections [21]. Bactericidal exercises of silver nanoparticles of *Calophyllum apetalum* were indicated huge results. *Escherichia coli* development was considerably repressed at all the convergences of tried silver nanoparticles after 4 h of incubation where as the development of *Pseudomonas aeruginosa* was totally hindered at all fixations (Figure 5 and 6). Be that as it may, the orchestrated silver nanoparticles were neglected to show development hindrance against *Staphylococcus aureus* and *Klebsiella pneumonia*.



Be that as it may, the pharmaco alterable and active properties of AgCINPs towards these microorganisms are yet to be known. It was expected that silver and silver nanoparticles stick to the surface proteins of bacterial cells. The ionic way of bacterial surface causes layer depolarization and structures porosity of cell membrane [22,23]. Silver nanoparticles are having a solid cooperation with the thiol gathering of key respiratory proteins in bacteria [24].

It was realized that, particularly Ag<sup>+</sup> ties to sulfur containing peptides and breaks the disulphide linkages in DNA and proteins and there by Ag<sup>+</sup> meddle in all cell capacities by hindering DNA-replication [25,26]. Aside from this, the silver nanoparticles meddle with the redox capability of cell layer and ribosomes driving the era of uncontrolled Reactive Oxygen Species (ROS) [27]. Again these free radicals destabilize the cell layer, cell proteins, causes oxidative anxiety and at the same time results in the loss of cell viability [28] (Figure 7).

In our study Calophyllum apetalum was taken for combination of silver nanoparticles in view of its restorative significance.



Figure 7. The general mechanism of antibacterial activity exhibited by silver chloride nanoparticles of Calophyllum apetalum

#### 4. Conclusions

The present study we infer that the amalgamation of silver nanoparticles by the fluid concentrate of leaves of Calophyllum apetalum. The delivered silver nanoparticles had with circular shapes and normal size of 86nm. UV-vis and SEM results affirmed that the silver nanoparticles created were steady, auxiliary examination by XRD affirmed the development of silver nanoparticles. Likewise, the antibacterial impact of the silver nanoparticles was measured by streak plate strategy. From our study it was plainly demonstrated that the silver nanoparticles smothered the www.iseeadyar.org

development of the tried microscopic organisms, for example, *Escherichia coli* and *Pseudomonas aeruginosa* at all convergences of blended silver nanoparticles. Thus, the consequences of our study are proposing that the orchestrated silver nanoparticles can be utilized for the advancement of antibacterial medications.

# 5. Acknowledgment

We express gratitude toward Dr MR Hulinaykar, Managing Trustee, Sri Shridevi Charitable Trust (R.) and Dr M.N. Eshwarappa, Principal, SIET, Tumakuru, India for support and proposals amid the examination, Dr K. G. Bhat, Taxonomist, Udupi, for his help with accumulation and recognizable proof of our plant. We express gratitude toward Center for Nano science and Engineering, Indian Institute of Science, Bangaluru for their stretched out administration in helping to portrayal of our sample by UV-vis, SEM and XRD.

# 6. References

- 1. Nagaraj Basavagowda, Lee Yong Rok. Synthesis of Gold and Silver Nanoparticles Using Leaf Extract of *Perilla frutescens* A Biogenic Approach. Journal of Nanoscience and Nanotechnology. 2014; 14(6), 4377-4382.
- 2. Zygmunt Sadowski. Biosynthesis and Application of Silver and Gold Nanoparticles, Silver Nanoparticles. Wroclaw University of Technology. Poland. 2010.
- 3. Mahendra Rai, Alka Yadav, Aniket Gade. Current trends in phytosyn thesis of metal nanoparticles. Critical Reviews in Biotechnology. 2008; 28, 277-284.
- 4. Kaushik N Thakkar, M.S., Snehit S. Mhatre, M.S., Rasesh Y. Parikh. Biological synthesis of metallic nanoparticles. Nanomedicine: Nanotechnology, Biology, and Medicine. 2010; 6(2), 257-262.
- 5. M. Stoytcheva. Pesticides Formulations, Effects, Fate. 9<sup>th</sup> edn. InTechJanezaTrdine, Croatia, 2011.
- 6. M. Linuma, T. Ito, H. Tosa, T. Tanaka, R. Miyake, V. Chelladurai. Prenylatedxanthonoides from *Calophyllum apetalum*. Phytochemistr. 1997; 46 (8), 1423-1429.
- 7. Murali Sastry, Absar Ahmad, M Islam Khan, Rajiv Kumar. Biosynthesis of metal nanoparticles using fungi and actinimycetes, Current science. 2003; 85(2), 162-170.
- 8. S. Gopalakrishnan, S. Yamini Sudha Lakshmi, Fouzia Banu. Antimicrobial activity of synthesized silver nanoparticles and phytochemical screening of the aqueous extract of *Mussaenda ferruginea*, Indian Journal of Nanoscience. 2015; 3(2), 1-4.
- 9. A.A. Revina, E.M. Egorova. International conference on advances in technology in 21<sup>st</sup> century (ICAT), Moscow, 1998, 411-413.
- Cheng-An J. Lin, Ting-Ya Yang, Chih-Hsien Lee, Sherry H. Huang, Ralph A. Sperling, Marco Zanella, Jimmy K. Li, Ji-Lin Shen, Hsueh-Hsiao Wang, Hung-I Yeh, Wolfgang J. Parak, Walter H. Chang. Synthesis, characterization and bioconjugation of flurescent gold nanoclusters toward biological labeling applications. American Chemical Society Nano. 2009; 24(2), 395-401.
- 11. Jagessar R.C, Masara A, Gomes G. Selective Antimicrobial properties of *Phyllanthus acidus* leaf extract against *Candida albicans, Escherichia coli* and *Staphylococcus aureus* using Stokes Disc diffusion, Well diffusion, Streak plate and a dilution method, Nature and science. 2008; 6(2), 24-38.
- 12. Kannaiah Paulkumar, Shanmugam Rajeshkumar, Gnanajobitha, Mahendran Vanaja, Chelladurai Malarkodi, Gurusamy Annadurai. Biosyntheis of silver chloride nanoparticles using *Bacillus subtilis* MTTC 3053 and assessment of its antifungal activity. International Scholarly Research Notices Nanomaterials. 2013; 2013, 1-8.
- 13. Morones J.R, Elechiguerra J.L, Camacho A, Holt K, Kouri J.B, Ramirez J.T, Yacaman M.J. The bacterial effect of silver nanoparticles. Nanotechnology. 2005; 16(10), 2346-2353.
- 14. Smitha S.L, Philip D, Gopchandran K.G. Green synthesis of gold nanoparticles using *Cinnamomum zeylanicum* leaf broth. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy. 2009; 74(3), 735-739.
- 15. Khushboo Singh, Manju Panghal, Sangeeta Kadyan, Uma Chaudhary' Jaya Parkash Yadav. Antibacterial activity of synthesized silver nanoparticles from *Tinosporacordifolia* against multi drug resistant strains of *Pseudomonas aeruginosa* isolated from burn patients. Journal of nanomedicine and nanotechnology. 2014;5(2), 1-6.
- 16. Kleemann W. Random field induced antiferromagnetic, ferromagnetic and structural domain states. International journal of modern physics B. 1993;7(13), 2469-2507.
- 17. Gopalakrishnan S, Philip Kaupa, Yamini Sudha Lakshmi S, Fouzia Banu. Antimicrobial activity of synthesized silver nanoparticles and phytochemical screening of the aqueous extract of Similax latifolia, Indian Journal of Nanoscience. 2015; 3 (2), 1-4.

- 18. Preetha Devaraj, Prachi Kumari, Chirom Aarti, Arun Renganathan. Synthesis and characterization of silver nanoparticles using *Cannonball* leaves and their cytotoxic activity against MCF-7 cell line. Journal of nanotechnology. 2013; 2013, 1-5.
- 19. Aryou Emamifar, Mahdi Kadivar, Mohammad Shahedi, Sabihe Soleimanian-Zad. Evaluation of nanocomposite packaging containing Ag and Zno on shelf life of fresh orange juice. Innovative food science and emerging technologies. 2010; 11(4), 742-748.
- 20. Nagaraj Basavegowda, Akbar Idhayadhulla, Yong RokLee. Tyrosinase inhibitory activity of silver nanoparticles treated with *Hovenia dulsis* fruit extract: An *in vitro* study. Materials letters. 2014; 12, 928-30.
- 21. Khushboo Singh, Manju Panghal, Sangeeta Kadyan, Uma Chaudhary Jaya Parkash Yadav. Antibacterial activity of synthesized silver nanoparticles from *Tinospora cordifolia* against multi drug resistant strains of *Pseudomonas aeruginosa* isolated from burn patients. Journal of nanomedicine and nanotechnology. 2014; 5(2), 1-6.
- 22. Gianluigi Franci, Annarita Falanga, Stefania Galdiero, Luciana Palomba, Mahendra Rai, Giancarlo Morelli, Massimiliano Galdiero. Silver nanoparticles as potential antibacterial agents. Molecules. 2015; 20(5), 8856-8874.
- 23. Rajeshkumar S, Malarkodi C. *In vitro* antibacterial and mechanism of silver nanoparicles against food borne pathogens, Bioorganic chemistry and applications. 2014; 2014, 1-10.
- 24. Oliver Gordon, Tunde VigSlenters, Priscilla S. Brunetto, Amer E. Villaruz, Daniel E. Sturdevant, Michael Otto, Regine Landmann, Katharina M. Fromm. Silver coordination polymers for prevention of implant infection: thiol interaction, impact on respiratory chain enzymes, and hydroxyl radical induction. Antimicrobial agents and chemotherapy. 2010; 54(10), 4208-4218.
- 25. Sunkar S, Nachiyar C.V. Biogenesis of antibacterial silver nanoparticles using the endophytic bacterium *Bacillus cereus* isolated from *Garcinia xanthochymus*. Asian Pacific Journal of Tropical Biomedicine. 2012; 2(12), 953-959.
- 26. Shakeel Ahmed, Mudasir Ahmad, BabuLal Swami, Saiqalkram. A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: A green expertise. Journal of Advanced Research. 2016; 7(1), 17-28.
- 27. Kenneth K. Y. Wong, Xuelai Liu. Silver nanoparticles the real "silver bullet" in clinical medicine?. Medicinal Chemical Communications. 2010; 1(2), 125-131.
- 28. Bogumila Reidy, Andrea Haase, Andreas Luch, Kenneth A. Dawson, Iseult Lynch. Mechanisms of silver nanoparticle release, transformation and toxicity: A critical review of current knowledge and recommendations for future studies and applications, Materials.2013; 6(6), 2295-2350.

The Publication fee is defrayed by Indian Society for Education and Environment (iSee). www.iseeadyar.org

#### Citation:

Chandrappa CP, Govindappa M, Chandrasekar N, Sonia Sarkar, Sepuri Ooha, Channabasava R, Vijay Kumar GS, Ramesh C P. *Calophyllum apetalum* interceded blend of silver nanoparticles and their antimicrobial impact. *Indian Journal of Nanoscience*. 2016; 4(2), September.